REVIEW

Perioperative fluid management in renal transplantation: a narrative review of the literature

Peter Schnuelle and Fokko Johannes van der Woude

Medical Clinic V, Medical Faculty of the University of Heidelberg, University Hospital Mannheim, Mannheim, Germany

Keywords

colloids, crystalloids, delayed graft function, fluid therapy, kidney transplantation.

Correspondence

Dr Peter Schnuelle MD, Medical Clinic V, Medical Faculty of the University of Heidelberg, University Hospital Mannheim, Mannheim, Germany. Tel.: 0049621 383 2751; fax: 0049621 383 3804; e-mail: peter.schnuelle@med5.ma.uni-heidelberg.de

Received: 2 February 2006 Revision requested: 4 March 2006 Accepted: 20 May 2006

doi:10.1111/j.1432-2277.2006.00356.x

Summary

Adequate volume maintenance is essential to prevent acute renal failure during major surgery or to ensure graft function after renal transplantation. The various recommendations on the optimum fluid therapy are based, at best, on sparse evidence only from observational studies. This article reviews the literature on perioperative fluid management in renal transplantation. Crystalloid solutions not exerting any specific side-effects are the first choice for volume replacement in kidney transplantation. The use of colloids should be restricted to patients with severe intravascular volume deficits necessitating high volume restoration. The routine application of albumin, dopamine, and high dose diuretics is no longer warranted. Mannitol given immediately before removal of the vessel clamps reduces the requirement of post-transplant dialysis, but has no effects on graft function in the long term. There is insufficient evidence on the best use of dialysis, but it seems peritoneal dialysis pretransplant is associated with less delayed graft function, whereas the preference of dialysis posttransplant is not yet well-founded. This review article should provide better guidance for fluid management in kidney transplantation until best-evidence guidelines can be established based upon more research.

Introduction

Delayed graft function (DGF), a term interchangeably used for acute renal failure (ARF) after transplantation, exerts an enduring and powerful effect on the subsequent clinical course after kidney transplantation [1]. DGF is grossly defined by the need for dialysis in the first week post-transplant. It may be considered as the result of an accumulation of various deleterious factors for the kidney graft. Donor related characteristics such as age, tissue quality, and brain death play a key role in transplant success. Factors related to procurement, cold storage, and reperfusion injury are crucial for the early performance of the graft and affect its long-term functioning [2-5]. Variables related to the recipients, including prerenal causes, immunosuppressive drugs, human leukocyte antigenmatching and sensitization have an impact on the risk of DGF. It is well recognized that DGF enhances the susceptibility for rejection [6-8]. The complex interrelationship of DGF and allograft immunogenicity has been summarized elsewhere [9].

Implementing an intervention before the damage has occurred is the best way to attenuate DGF. Fluid therapy has been shown to be effective for preventing ARF in certain clinical scenarios. However, in acute tubular necrosis (ATN), only supportive care has been shown to be efficacious. In kidney transplantation, partially self-contained modalities for perioperative fluid management have been developed in recent decades. These therapies must be evaluated on the basis of evidence from more recent clinical data, in regards to their efficacy, potential side-effects and patient outcomes. In this article, we review the available literature on fluid management in renal transplantation. PubMed was searched using the key-terms 'renal transplantation', 'fluid therapy', 'fluid management', 'crystalloids', 'colloids', 'albumin', 'mannitol', 'dopamine', 'dialysis', 'acute renal failure', and 'delayed graft function'.

Avoidance of hypovolemia

On points where there is insufficient evidence specifically on kidney transplantation, comparable studies on fluid management in the critically ill and in patients undergoing major surgery are reviewed. Perioperative fluid management must ensure the restoration and maintenance of the intravascular volume, in order to obtain an appropriate graft function. In experimental animal models of ischemic ATN, renal perfusion is linearly dependent on the mean arterial pressure, even in the normal blood pressure range. Paradoxical renal vasoconstriction occurs at a low mean arterial pressure [10,11]. In transplantation, denervation adds to a deteriorated hemodynamic autoregulation of the kidney graft [12-15]. Thus, mild or severe decreases in blood pressure can further reduce renal perfusion and thereby result in repeat ischemia to the transplanted kidneys. Under physiologic conditions, the intravascular volume is tightly regulated by various mechanisms, including the transmembrane filtration pressure, interstitial hydrostatic pressure, colloid osmotic pressure, lymphatic transport, sympathoadrenergic system, and renin-angiotensin system [16,17]. Severe intravascular volume deficiency can overextend the compensatory capacity of the effector mechanisms. Consequently, maldistribution of the nutritional blood flow and tissue hypoxemia may occur [18-22]. Rectification of intravascular volume deficiencies is therefore essential to obtain an adequate systemic circulation and microcirculation [23,24]. Ensuring adequate volume status must be part of any treatment strategy.

In general, any volume replacement is better than none in the hypovolemic state, and an appropriate volume amount is probably more important than the kind of fluid [25,26]. Furthermore, there is only sparse evidence to suggest that the fluid type in patients with risk for ARF should be different from that for other critically ill patients [27]. Crystalloid solutions are usually the first choice to correct for fluid and electrolyte imbalances in these patients [28]. However, in instances of severe hypovolemia colloid solutions may be preferable for obtaining sufficient tissue perfusion, particularly in situations of enhanced capillary permeability [29]. In addition to restoration of the intravascular volume, colloid solutions may ameliorate impaired microcirculation [29–31]. Treatment of intravascular hypovolemia has changed significantly during recent decades. There has been a widespread shift in clinical practice from natural colloids, such as blood, albumin, and fresh-frozen-plasma, over to crystalloids and synthetic colloids, such as hydroxethyl starch (HES), bovine derived gelatin products, and dextrans. These crystalloids and synthetic colloids are now the preferred substitutes for the treatment of hypovolemia. Despite a large number of studies and recommendations from consensus meetings, the optimal type of resuscitation fluid in patients with impending ARF is still not yet well established.

The choice of a particular fluid in a given clinical situation can be guided by an understanding of the solutions' properties, but nonetheless there is still an ongoing debate on the relative merits of crystalloid and colloid solutions. Crystalloid therapy increases the formation of edemas, but colloids have known adverse side effects. There are experimental studies both that support [32,33] and that refute [34] the assertion that tissue oxygen extraction is disabled by the accumulation of interstitial fluid. Evidence from a randomized phase III clinical trial suggests that fluid resuscitation with colloids expedites recovery in the postoperative period after major surgery [35]. Patients assigned to receive Hextend, a physiologically balanced plasma expander for large volume use, had less nausea, vomiting, and severe pain. This was attributed to the lower degree of interstitial fluid accumulation in these patients. Nonetheless, until specific side effects of the colloids cannot be excluded at all, crystalloids probably remain preferable for most situations.

There is no clear evidence to date that the fluid type administered has an influence on mortality. Several randomized controlled trials (RCT) have been conducted comparing colloid and crystalloid fluid therapy in a variety of clinical settings. Of these, only a limited number of large-scale studies have evaluated the effects of the different fluid classes on patient outcomes. The vast majority of the studies were neither designed nor sufficiently powered to investigate mortality as an endpoint. Meta-anlyses found no survival benefit in favor of either treatment. This holds true for both the comparison of crystalloids with any colloid [36,37], as well as for comparisons within the group of the various colloid preparations, including albumin, dextrans, gelatins, and HES [26]. The most recent meta-analysis published by the Cochrane Collaboration [37] involved a total of 7576 patients from randomized and quasi-randomized trials of colloids compared with crystalloids. The pooled estimate of the relative risk was 1.02 (95%CI: 0.93-1.11). Thus, clinicians' main focus should be foremost on maintaining an adequate volume level, and only secondarily on which fluid they use to do it.

Use of crystalloid solutions

Isotonic crystalloid solutions, such as 0.9% saline solution and Ringer's lactate solution, are the first choice for volume restoration and for correcting of imbalances in homeostasis. Unlike plasma expanders, crystalloid solutions have no nephrotoxic or other specific side-effects. Isotonic crystalloid solutions are distributed rapidly into

the interstitial compartment and have a half-life of 20–30 min in the intravascular space. Consequently, the effect on plasma volume expansion is limited and does not exceed 20% of the volume applied [17,23]. To compensate for blood loss, crystalloid solution require a quantity four to five times greater than colloid solutions to exert the same volume effect [28], but crystalloids alone are incapable of restoring microcirculation in cases of severe bleeding [38,39]. Kidney transplantation can usually be performed without the need for plasma expanders, because major blood losses are uncommon during this operation.

Balanced crystalloid solutions are often preferable to saline-based fluids in major surgery. Large-volume administration of 0.9% saline may result in hyperchloremic metabolic acidosis, because of the high chloride load. By contrast, balanced crystalloid solutions are not associated with the same disturbance of the acid-base status and electrolyte status [40-43]. Patients randomly assigned to balanced solutions, when compared with those receiving saline-based fluids, showed less impairment of hemostasis [35] and enhanced gastric perfusion [42]. Renal function may also be better preserved [42]. Balanced crystalloid solutions containing potassium should be avoided though during renal transplantation, because they can aggravate hyperkalemia in instances of impaired graft function. Hyperkalemia may be life threatening and require acute hemodialysis. Thus, clinicians who consider switching to balanced crystalloid solutions in patients with impending graft function must be aware of this complication. Close monitoring of serum electrolytes remains a cornerstone of care for guiding fluid therapy in kidney transplantation.

Use of colloid solution

Natural colloids such as albumin are being widely replaced by synthetic colloids such as dextrans, gelatins, and solutions of hetastarch. Colloids are retained in the intravascular compartment because of their content of macromolecules. The degree of plasma volume expansion exerted by colloids is determined by their concentration, molecular weight, and structure, as well as by the colloid osmotic pressure, metabolism, and elimination rate [17,28]. The rate of loss through the capillary endothelial barrier into the interstitial compartment and through the glomerular basement membrane into the proximal tubule obeys the molecular size and surface charge characteristics. The predominant effect on whole blood viscosity is mediated through simple hemodilution, thereby enhancing blood flow characteristics [44]. However, the semisynthetic colloids also affect red cell aggregation, which adds to their overall effect on the blood flow characteris-

tics. Furthermore, all of the semi-synthetic colloids may prolong coagulation. HES solutions exert varying effects on clotting characteristics, which depend on the size of the HES molecules and the degree of hydroxethyl substitution [45,46]. Impaired platelet function, a von Willebrand-like syndrome (with reduction in vWF and factor VIIIc), and impaired coagulation as measured by thromb-elastography have been reported to arise during the administration of HES [47-49]. This raises some concern for end stage renal disease (ESRD) patients undergoing kidney transplantation, because they are prone to bleeding complications because of uremic platelet dysfunction [50-52]. Although it is rare, severe and lifethreatening anaphylactic reactions have been observed in association with any of the commonly used semisynthetic colloids and with albumin. The incidence of severe anaphylactic reactions is probably more frequent for gelatins (0.35%) and for dextrans (0.27%) than for albumin (0.10%) or for starches (0.06%) [53]. This needs to be taken into account when balancing the merits for the use of plasma expanders with crystalloid solutions.

Human albumin has been widely used as the 'natural colloid' for the treatment of hypovolemia in critically ill patients in past decades. Albumin administration is costly though and does not provide any outcomes benefits for patients with hypovolemia or hypoalbuminemia [54,55]. In the clinical situation of capillary leakage, the administration of albumin may even expedite edema formation, because of an increased shift of plasma proteins to the interstitial compartment [23,24]. There is currently little evidence that warrants the use of albumin in the ICU setting. A systematic review of human albumin in the critically ill suggested that administration might even be associated with a higher mortality [56]. That review was widely criticized though for the heterogeneity of the studies included. More recent data from the SAFE Study involving 6997 patients in a large multicenter RCT suggested that there was no difference in mortality between patients managed with either 4% albumin or normal saline for fluid resuscitation [57]. Nonetheless the low-cost benefit ratio and associated risks argue against the further use of albumin in general surgery.

Although there is little evidence supporting the use of albumin for hypovolemia or hypoalbuminemia in critically ill patients, several observational studies have been published which suggest that volume expansion with human albumin improves the short-term and long-term outcomes of kidney transplant recipients. In particular, human albumin improves the onset and the extent of the urine volume output post-transplant, the renal function, and the 1-year graft survival rate [58–60]. The largest series, involving 438 recipients of renal transplants from deceased donors, revealed a statistically significant benefit

from the usage of albumin, though mannitol, furosemide, and electrolyte solutions were given concomitantly [60]. Protective properties have also been attributed to the intraoperative administration of mannitol during the vascular phase [61–63]. Apart from the induction of osmotic diuresis, the salutary effect is thought to be mediated through the antioxidant properties of sugar alcohols and chemically related substances [9,64]. Controlled clinical data investigating only the effect of albumin infusions in kidney transplantation are not available, so their use in this setting should proceed with caution until more research is available.

Two of the synthetic colloids that have widely replaced albumin in clinical practise - dextrans and gelatins - do not seem on the whole to be preferable to albumin. A randomized study comparing intraoperative albumin and dextran-40 in renal transplant recipients from a living related donor did not find any difference between the two treatments in regards to urine volume output and serial serum creatinine concentrations, post-transplant [65]. The value of this study may be limited, because with only 17 patients the researches may not have had enough statistical power to detect outcome differences. Dextran solutions have been associated with serious side-effects, such as coagulation disturbances [18,66,67], highly severe hypersensitivity reactions [53,68-70], and the onset of oliguric or anuric renal failure [71-75]. This has led to major concern on their usage for volume expansion in the critically ill and in kidney transplantation. Likewise, gelatin preparations do not fulfill the first-choice requirements in the ICU setting [28]. Gelatins exert a more limited effect on intravascular volume resuscitation, because they contain a high proportion of low-molecular weight components. A colloid fluid regimen confined to gelatin may be less effective for patients with severe volume deficiency [23,76]. To some extent, the absence of dose limitations outweighs the disadvantage of low efficacy for volume expansion [28]. The high potassium and calcium contents of 3.5% urea-cross-linked gelatin renders them inapplicable for ARF and for perioperative care during kidney transplantation [77]. Thus, there is no reason to use dextran or gelatin instead of albumin.

By contrast, the recent trend to using solutions of hetastarch instead of albumin seems generally founded in the evidence. Solutions of hetastarch (HES, hydroxyethyl-starch) are synthesized from natural polymers of amylopectin. The pharmacokinetics of HES depend on the degree of substitution at carbons 2, 3, and 6 in the glucose ring in combination with the molecular weight, because the C2/C6 hydroxyethylation ratio influences their degradation mainly by nonspecific plasma amylases [46,78]. The optimum HES solution combines the lowest *in vivo* molecular weight above the threshold for renal elimination

with a low degree of hydroxyethyl substitution [78]. Easily degradable HES solutions, dominated by medium molecular weight, meet these specifications. They are clinically applied for various indications including isovolemic hemodilution, perioperative volume substitution, cardiac surgery, trauma, and sepsis [55,79–85]. They do not have considerable side-effects regarding bleeding complications, the reticuloendothelial system, or renal function, if given below their upper dosage limits. With regard to safety considerations [84], HES solutions with a low to medium *in vivo* molecular weight may offer the best risk to benefit ratio among the available synthetic colloids [28].

There has been some debate about whether HES specifically impairs renal function. An 80% rate of osmotic, nephrosis-like lesions was reported in transplanted kidneys after routine administration of HES 200/0.62 to brain-dead donors [86]. This prompted a prospective randomized trial comparing HES 200/0.62 and gelatin for plasma-volume expansion in brain-dead organ donors. The study found that HES was associated with impaired immediate renal function in kidney transplant recipients, because of a more frequent necessity of hemodialysis and because of significantly higher serum-creatinine concentrations 10 days after transplantation. Furthermore, renal biopsies showed osmotic, nephrosis-like lesions only in the HES treated group, although starch was not found in the vacuoles of the proximal tubular cells [87]. The clinical relevance of these lesions has however been questioned [88], because tubular vacuolizations are not a specific morphological finding and have also been observed in association with dextran, 20% mannitol, and with intravenous immunoglobulin, with and without accompanying ARF [74,89-95]. Considering the pathogenesis of hyperoncotic renal failure [96], it may be hypothesized that all colloids can induce this kind of renal function impairment. In the absence of a direct chemical toxicity, the most likely mechanism for HES-induced renal dysfunction may be swelling and vacuolization of tubular cells and tubular obstruction due to the production of hyperviscous urine. The risk of high plasma colloid osmotic pressure and subsequent renal dysfunction presumably increases with repeated doses of highly concentrated, slowly degradable HES of high molecular weight and high degree of substitution [74,97]. A more recent retrospective study concluded that HES compounds given at a maximum dose of 15 ml/kg/day to organ donors have no detrimental influences on graft function in kidneys preserved in University of Wisconsin (UW) solution or histidin-tryptophan-ketoglutarate (HTK) solution [98].

Treatment with HES needs to be accompanied by sufficient amounts of crystalloid solution. Careful monitoring of kidney function and dose reductions are required in patients with renal function impairment. Although HES 200/0.5 is considered an effective, safe, and economically attractive colloid solution in the critically ill, a restricted usage has been recommended for kidney transplantation because of the potential of side-effects [28].

Table 1 provides a summary of the key investigations on fluid type in kidney transplantation, as well as in the critically ill or in patients undergoing major surgery.

Mannitol, loop diuretics, and low-dose dopamine

Mannitol is widely used in kidney transplantation, immediately before opening the vascular anastomoses. Mannitol, an inert sugar, confers protection against renal cortical ischemia by expanding the intravascular volume, diminishing the potential of tubular obstruction and increasing tubular flow rate through prevention of water reabsorption in the proximal tubule. Furthermore, mannitol enhances the release of vasodilatory prostaglandins in the kidney [99] and may act as a free radical scavenger [100,101]. Clinical, single-center studies have found salutary effects of mannitol infusions in kidney transplantation [63,102-105]. Some of these studies have been retrospective analyses or have involved only a limited number of patients. Nonetheless, the sparse controlled data available have clearly shown that 250 ml of mannitol 20% given immediately before vessel clamp removal reduces the incidence of ARF, as indicated by a lower requirement of post-transplant dialysis [61,106,107] [Table 2]. However, 3 months after transplantation no difference was found in kidney function compared with patients who did not receive mannitol [61]. The usage of mannitol also has risks, because of the potential to induce rapid intravascular volume expansion, which leads to pulmonary edema. Concomitant hydration is indispensable for the optimal prevention of ARF. Overzealous administration (>200 g/day) may be harmful and can result in hyperoncotic kidney failure [90-92], as mentioned previously. Thus, mannitol should be used before opening the vascular anastomoses but moderately and with accompanying hydration.

Loop diuretics are thought to counteract the increased response of antidiuretic hormone to surgical stress [108]. They exert their pharmacological effect in the ascending loop of Henle. In kidney transplantation, furosemide is commonly given during the vascular anastomosis to stimulate diuresis, although it is unknown whether it actually improves early function or simply enhances the amount of urine production from a functioning kidney [109]. Despite their frequent use, there is no evidence that loop diuretics shorten the duration of ARF, reduce the subsequent requirement for dialysis, or improve outcomes in patients with ARF [110–114] [Table 2]. Loop

rable 1. Overview of studies and key references on types of fluid therapy in kidney transplantation and in critically ill/major surgery

	Kidney transplantation	tion			Critically ill/major surgery			
Treatment modality Study design	Study design	Parameter of efficacy	Outcome	Reference	Study design	Parameter of efficacy	Outcome	Reference
Colloids versus crystalloids	No data	ı	1	ı	Meta-analysis of RCTs	Survival	No difference	Choi <i>et al.</i> [36]; Roberts <i>et al.</i> [37]
					Multi-center RCT	Postoperative recovery	Beneficial effect for colloids	Gan <i>et al.</i> [35]
Albumin versus	Observational/	Urine output,	Beneficial	Dawidson <i>et al.</i> [58];	Meta-analysis of RCTs	Survival	No difference	Roberts <i>et al.</i> [37]
crystalloids	uncontrolled retrospective studies	renal Tunction, 1 year graft survival	enect for albumin	vviilms et al. [59]; Dawidson e <i>t al.</i> [60]	Large multi-center KCI			FINTER E <i>t al.</i> [57]
Comparison of colloid types	Single-center RCT	Urine volume s-creatinine	No difference	Dawidson <i>et al.</i> [65]	Meta-analysis of RCTs	Survival	No difference	Bunn e <i>t al.</i> [26]

able 2. Overview of studies and key references on mannitol, loop diuretics, and dopamine in kidney transplantation and in critically ill/major surgery

	Kidney transplantation	tation			Critically ill/major surgery	r surgery		
Treatment modality	Study design	Parameter of efficacy	Outcome	Reference	Study design	Parameter of efficacy	Outcome	Reference
Use of mannitol	RCT	ARF need for dialysis	Beneficial effect	Weimar <i>et al.</i> [61]; Tiggeler <i>et al.</i> [106];	I	1	ı	1
				Van Valenberg <i>et al.</i> [107]				
Use of loop diuretics	Observational study	Need for dialysis	No effect	Lachance e <i>t al.</i> [109]	RCT	Recovery from ARF survival	No effect	Shilliday et al. [110]; Cantarovich et al. [114]
Use of renal	RCT	Need for	No effect	Grundmann et al. [120];	RCT	Prevention of ARF	No effect	Marik et al. [132];
dose dopamine		dialysis		Kadieva <i>et al.</i> [122]				Bellomo <i>et al.</i> [133];
								Lassnigg <i>et al.</i> [115]
			Beneficial	Carmellini <i>et al.</i> [121];	Meta-analysis			Kellum <i>et al.</i> [134];
			effect	Dalton <i>et al.</i> [124]*	of RCTs			Marik <i>et al.</i> [135]

RCT, randomized controlled trial; ARF, acute renal failure. *Study solely investigating renal hemodynamics and functional parameters. diuretics in extended dosages may even be harmful for the kidney [115], because they may disturb the protective corticomedullary redistribution of blood flow [116]. Thus, there are no indications for loop diuretics other than the removal of fluid overload that is contributing to organ dysfunction in the lung and heart.

Low-dose dopamine has been administered to increase renal blood flow, in the belief that this protects against renal failure. Studies on the efficacy of dopamine infusion in kidney transplantation are conflicting [117-119]. The majority of them failed to demonstrate any significant effect when dopamine was administered to the recipients following transplantation [120-123]. Low-dose dopamine given in the very early period after transplantation (3-6 h postoperatively) has recently been shown to significantly increase effective renal plasma flow, urine flow rate, creatinine clearance, and total urinary sodium excretion rate [124]. A persistent beneficial effect on kidney function could not be found for donors and recipients receiving dopamine during living donor nephrectomy [125]. Dopamine and chemically related catecholamines given to brain-dead organ donors to stabilize hemodynamics may improve the outcomes regarding acute rejection episodes, initial graft function, and graft survival [126-130]. Additionally, the antioxidant properties of catecholamines and chemically-related substances may protect endothelial cells from preservation injury during prolonged cold storage [131]. Nonetheless, the available evidence does not warrant the routine use of dopamine for perioperative care in kidney transplant recipients or in the critically ill with impending or overt renal failure. Dopamine has been administered in the belief that it reduces the risk of renal failure or ameliorates its severity and duration by increasing renal blood flow, but no clinical protection from this has been found [115,132,133]. It was concluded from meta-analyses that dopamine should not be given for these indications and should be eliminated from routine clinical use, given its potential side-effects [134,135] [Table 2].

Dialysis therapy and fluid overload

There is still an inadequate state of research on dialysis therapy before and after kidney transplantation. Before transplantation surgery, many patients present with a contracted volume, as they have been dialyzed to dry weight. It therefore seems reasonable to restrict fluid removal during preoperative dialysis to a target of 1–2 kg above the former dry weight. A Belgian case–control study [136] and a large American cohort investigation of nearly 23 000 transplant recipients [137] have shown that DGF occurs more frequently in hemodialysis patients than in those on peritoneal dialysis. Although the

causation could not be determined from the data, this finding can be taken as indicating that hypovolemia is a prerenal risk factor for DGF [9].

Post-transplant dialysis is required for transplant patients who develop DGF with oliguria. Hemodialysis is commonly preferred, but clinical criteria for the best use of dialysis after transplantation are not well established. Unlike for patients with acute ARF [138,139], studies investigating the effect of biocompatible dialysis membranes in DGF after transplantation failed to demonstrate any difference regarding the average number of hemodialysis treatment sessions, mean time to recovery, or graft outcomes [140-142] [Table 3]. Peritoneal dialysis can safely be continued in patients formerly on this treatment without major complications or increased frequency of peritonitis. Thus, the preference for hemodialysis posttransplant does not seem founded. Retrospective analyses of the existing clinical data on dialysis therapy after kidney transplantation could quickly illuminate this issue until more controlled prospective studies can be carried out.

Postoperative hyperkalemia and fluid overload are prevalent indications for acute hemodialysis. Overzealous fluid administration may increase the demand on cardiac function, leading to myocardial dysfunction and associated morbidity. A more intense fluid regimen in the critically ill does not reduce mortality [143-145] and even precipitates noncardiogenic pulmonary edema [146,147]. A dose-response relation was observed between complications and increasing intravenous fluid volumes as well as increasing body weight [148]. In a prospective study of 48 consecutive postoperative patients admitted to a surgical ICU, mortality in the patients who gained more than 10% body weight was 31.6% as compared with 10.3% in the group that gained <10% body weight [149]. Elderly patients may have accumulated substantial co-morbidities during their life-time and in association with long-term dialysis [150,151]. They constitute a growing population of the dialysis patients entering the waiting lists for transplantation. Thus in contrast to recommendations from very early studies [58,59,65,152], a zealous fluid replacement regimen is no longer warranted, given the clinical outcomes and changing patient demographics.

Recent studies in gastrointestinal surgery advocate a more restrictive regimen for perioperative fluid administration [153]. A restricted intravenous fluid substitution predisposes the patient to less edema formation of the gut. This may shorten postoperative hypomotility of the bowel and may facilitate the onset of enteric alimentation. The potential for bacterial translocation and development of sepsis is also reduced. Reduced edema may improve tissue oxygenation and wound healing [154].

able 3. Overview of studies and key references on the amount of volume substitution and renal replacement therapy in kidney transplantation and in critically ill/major surgery

	Kidney transplantation	ntation			Critically	Critically ill/major surgery		
Treatment modality	Study design	Parameter of efficacy	Outcome	Reference	Study design	Study Parameter of design efficacy	Outcome	Reference
Use of biocompatible membrane RCT for hemodialysis	RCT	Recovery from DGF No effect	No effect	Valeri <i>et al.</i> [140]; Romao <i>et al.</i> [141]; Woo <i>et al.</i> [142]	RCT	Recovery from ARF survival	Recovery from Beneficial effect ARF survival for biocompatible membranes	Hakim <i>et al.</i> [138]; Himmelfarb <i>et al.</i> [139]
Large volume therapy versus restricted volume therapy	Observational studies	Observational Not well-defined: studies onset and extent of urine output	Beneficial effect for large volume therapy	Luciani e <i>t al.</i> [152]; Dawidson e <i>t al.</i> [58]; Willms e <i>t al.</i> [59];	RCT	Survival	No effect or even adverse effect for large volume therapy	Hayes <i>et al.</i> [143]; Gattinoni <i>et al.</i> [144]
			:	Dawidson et al. [65]		Postoperative recovery	Adverse effect for large volume therapy	Lobo <i>et al.</i> [157]; Brandstrup <i>et al.</i> [148]

RCT, randomized controlled trial; DGF, delayed graft function; ARF, acute renal failure.

Retrospective analyses and prospective controlled clinical studies specifically addressing this issue have shown a clear benefit with regard to reductions of overall complications [148,155-157]. In particular, they have shown fewer pulmonary complications, a faster in-hospital recovery, and a trend towards a reduced perioperative mortality [148] [Table 3]. No deaths occurred in the fluid restricted group of the Danish multicenter RCT, whereas, four patients (4.7%) died in the standard group. The causes of death included pulmonary edema in two patients. pneumonia with septicemia, and pulmonary embolism [148]. Thus, simple clinical measures - such as fluid balance, arterial blood pressure, clinical assessment of peripheral edema, and carefully measured daily body weight – remain the key parameters for the monitoring of fluid therapy in surgery patients and transplant recipients.

Conclusions

Crystalloids without side-effects are the first choice for volume replacement in kidney transplantation. The routine use of various cocktails containing albumin, dopamine, and high dose diuretics is no longer warranted. Overzealous fluid administration should also be avoided, because it can be harmful or deadly. Adequate intravascular volume load should be maintained instead by restricted intravenous fluid substitution.

Acknowledgements

The authors would like to thank Michael Hanna, PhD, for his support in editing and proof-reading the manuscript.

References

- Halloran PF, Hunsicker LG. Delayed graft function: state of the art, November 10–11, 2000. Summit meeting, Scottsdale, Arizona, USA. Am J Transplant 2001; 1: 115.
- Cosio FG, Qiu W, Henry ML, et al. Factors related to donor organ are major determinants of renal allograft function and survival. *Transplantation* 1996; 62: 1571.
- 3. Halloran P, Aprile MA. Factors influencing early renal function in cadaver kidney transplants. A case control study. *Transplantation* 1988; **45**: 122.
- Gourishankar S, Jhangri GS, Cockfield SM, Halloran PF. Donor tissue characteristics influence cadaver kidney transplant function and graft survival but not rejection. *J Am Soc Nephrol* 2003; 14: 393.
- 5. Terasaki PI, Cecka JM, Gjertson DW, Takemoto S. High survival rates of kidney transplants from spousal and living unrelated donors. *N Engl J Med* 1995; **333**: 333.
- Gjertson DW. Impact of delayed graft function and acute rejection on kidney graft survival. Clin Transpl 2000; 6: 467.

- 7. Qureshi F, Rabb H, Kasiske BL. Silent acute rejection during prolonged delayed graft function reduces kidney graft survival. *Transplantation* 2002; **74**: 1400.
- Brennan TV, Freise CE, Fuller F, Bostrom A, Tomlanovich SJ, Feng S. Early graft function after living donor kidney transplantation predicts rejection but not outcomes.
 Am J Transplant 2004; 4: 971.
- Perico N, Cattaneo D, Sayegh MH, Remuzzi G. Delayed graft function in kidney transplantation. *Lancet* 2004; 364: 1814.
- Adams PL, Adams FF, Bell PD, Navar LG. Impaired renal blood flow autoregulation in ischemic acute renal failure. *Kidney Int* 1980; 18: 68.
- 11. Conger JD, Robinette JB, Schrier RW. Smooth muscle calcium and endothelium-derived relaxing factor in the abnormal vascular responses of acute renal failure. *J Clin Invest* 1988; **82**: 532.
- 12. Morita K, Seki T, Nonomura K, Koyanagi T, Yoshioka M, Saito H. Changes in renal blood flow in response to sympathomimetics in the rat transplanted and dernervated kidney. *Int J Urol* 1999; **6**: 24.
- 13. Shannon JL, Headland R, MacIver AG, Ferryman SR, Barber PC, Howie AJ. Studies on the innervation of human renal allografts. *J Pathol* 1998; **186**: 109.
- Momen A, Bower D, Leuenberger UA, et al. Renal vascular response to static handgrip exercise: sympathetic vs. autoregulatory control. Am J Physiol Heart Circ Physiol 2005; 289: H1770.
- 15. Thomas MC, Mathew TH, Russ GR, Rao MM, Moran J. Perioperative blood pressure control, delayed graft function and acute rejection after renal transplantation. *Transplantation* 2003; 75: 1989.
- Schadt JC, Ludbrook J. Hemodynamic and neurohumoral responses to acute hypovolemia in conscious mammals. *Am J Physiol* 1991; 260: H305.
- 17. Grocott MPW, Mythen MG, Gan TJ. Perioperative fluid management and clinical outcomes in adults. *Anesth Analg* 2005; **100**: 1093.
- Groeneveld ABJ, Thijs LG. Hypovolemic shock. In: Parillo JE, Bone RC, eds. Critical Care Medicine: Principles of Diagnosis and Management. St Louis, MO, Mosby, 1995: 387–418.
- 19. Parrillo JE. Pathogenetic mechanisms of septic shock. *N Engl J Med* 1993; **328**: 1471.
- Astiz ME, Rackow EC. Mechanisms and classification of shock. In: Fein AM, Abraham EM, Balk RA, Bernard GR, Bone RC, Dantzker DR, Fink MP, eds. Sepsis and Multiorgan Failure. Baltimore: Williams & Wilkins, 1997: 11–20.
- Hinshaw LB. Sepsis/septic shock: Participation of the microcirculation: An abbreviated review. *Crit Care Med* 1996; 24: 1072.
- 22. Jawa RS, Solomkin JS. Cellular effectors of the septic process. In: Fein AM, Abraham EM, Balk RA, Bernard GR, Bone RC, Dantzker DR, Fink MP, eds. *Sepsis and*

- Multiorgan Failure. Baltimore: Williams & Wilkins, 1997: 74–99
- 23. Kreimeier U, Peter K. Strategies of volume therapy in sepsis and systemic inflammatory response syndrome. *Kidney Int* 1998; **53** (Suppl. 64): S75.
- Boldt J. Human albumine on the intensive care unit: Can we live without it? In: Vincent JL, ed. *Yearbook of Inten*sive Care and Emergency Medicine. Berlin: Springer, 2000: 467–475.
- 25. Nolan J. Fluid resuscitation for the trauma patient. *Resuscitation* 2001; **48**: 57.
- Bunn F, Alderson P, Hawkins V. Colloid solutions for fluid resuscitation. Cochrane Database Syst Rev 2003: CD001319
- 27. Mehta RL, Clark WC, Schetz M. Techniques for assessing and achieving fluid balance in acute renal failure. *Curr Opin Crit Care* 2002; **8**: 535.
- 28. Ragaller MRJ, Theilen H, Koch T. Volume replacement in critically ill patients with acute renal failure. *J Am Soc Nephrol* 2001; **12**: S33.
- 29. Kreimeier U. Pathophysiology of fluid imbalance. *Crit Care* 2000; 4 (Suppl 2): S3.
- 30. Collis RE, Collins PW, Gutteridge CN, *et al.* The effect of hydroxyethyl starch and other plasma volume substitutes on endothelial cell activation; an in vitro study. *Intensive Care Med* 1994; **20**: 37.
- 31. Zikria BA, Subbarao C, Oz MC, *et al.* Macromolecules reduce abnormal microvascular permeability in rat limb ischemia-reperfusion injury. *Crit Care Med* 1989; **17**: 1306.
- 32. Baum TD, Wang H, Rothschild HR, Gang DL, Fink MP. Mesenteric oxygen metabolism, ileal mucosal hydrogen ion concentration, and tissue edema after crystalloid or colloid resuscitation in porcine endotoxic shock: comparison of Ringer's lactate and 6% hetastarch. *Circ Shock* 1990; 30: 385.
- 33. Gow KW, Phang PT, Tebbutt-Speirs SM, *et al.* Effect of crystalloid administration on oxygen extraction in endotoxemic pigs. *J Appl Physiol* 1998; **85**: 1667.
- 34. Ostgaard G, Reed RK. Interstitial fluid accumulation does not influence oxygen uptake in the rabbit small intestine. *Acta Anaesthesiol Scand* 1995; **39**: 167.
- 35. Gan TJ, Bennet-Guerrero E, Phillips-Bute B, *et al.* Hextend, a physiologically balanced plasma expander for large volume use in major surgery: a randomised phase III clinical trial. Hextend Study Group. *Anesth Analg* 1999; **88**: 992.
- 36. Choi PT, Yip G, Quinonez LG, Cok DJ. Crystalloid vs. colloids in fluid resuscitation: a systematic review. *Crit Care Med* 1999; 27: 200.
- Roberts I, Alderson P, Bunn F, Chinnock P, Ker K, Schierhout G. Colloids versus crystalloids for fluid resuscitation in critically ill patients. *Cochrane Database Syst Rev* 2004; CD000567.
- 38. Wang P, Ayala A, Dean RE, et al. Adequate crystalloid resuscitation restores but fails to maintain the active

- hepatocellular function following hemorrhagic shock. *J Trauma* 1991; **31**: 601.
- 39. Kreimeier U, Ruiz-Morales M, Messmer K. Comparison of the effects of volume resuscitation with dextran 60 vs. Ringer's lactate on central hemodynamics, regional blood flow, pulmonary function, and blood composition during hyperdynamic endotoxemia. *Circ Shock* 1993; **39**: 89.
- 40. McFarlane C, Lee A. A comparison of Plasmalyte 148 and 0.9% saline for intra-operative fluid replacement. *Anaesthesia* 1994; **49**: 779.
- 41. Scheingraber S, Rehm M, Sehmisch C, Finsterer U. Rapid saline infusion produces hyperchloremic acidosis in patients undergoing gynecologic surgery. *Anesthesiology* 1999; **90**: 1265.
- 42. Wilkes NJ, Woolf R, Mutch M, *et al.* The effects of balanced versus saline-based hetastarch and crystalloid solutions on acid-base and electrolyte status and gastric mucosal perfusion in elderly surgical patients. *Anesth Analg* 2001; **93**: 811.
- 43. Stillstrom A, Persson E, Vinnars E. Postoperative water and electrolyte changes in skeletal muscle: a clinical study with three different intravenous infusions. *Acta Anaesthesiol Scand* 1987; **31**: 284.
- 44. Audibert G, Donner M, Lefevre JC, Stoltz JF, Laxenaire MC. Rheologic effects of plasma substitutes used for preoperative hemodilution. *Anesth Analg* 1994; **78**: 740.
- 45. Strauss RG, Pennell BJ, Stump DC. A randomized, blinded trial comparing the hemostatic effects of pentastarch versus hetastarch. *Transfusion* 2002; **42**: 27.
- 46. Treib J, Haass A, Pindur G, Grauer MT, Wenzel E, Schimrigk K. All medium starches are not the same: influence of the degree of hydroxethyl substitution of hydroxyethyl starch on plasma volume, hemorrheologic conditions, and coagulation. *Transfusion* 1996; 36: 450.
- 47. Knutson JE, Deering JA, Hall FW, et al. Does intraoperative hetastarch administration increase blood loss and transfusion requirement after cardiac surgery? *Anesth Analg* 2000; **90**: 801.
- 48. de Jonge E, Levi M, Buller HR, Berends F, Kesecioglu J. Decreased circulating levels of van Willebrand factor after intravenous administration of a rapidly degradable hydroxyethyl starch (Haes 200/0.5/6) in healthy human subjects. *Intensive Care Med* 2001; 27: 1825.
- de Jonge E, Levi M. Effects of different plasma substitutes on blood coagulation: a comparative review. *Crit Care Med* 2001; 29: 1261.
- Hassan AA, Kroll MH. Acquired disorders of platelet function. Hematology (Am Soc Hematol Educ Program) 2005: 403.
- 51. Boccardo P, Remuzzi G, Galbusera M. Platelet dysfunction in renal failure. *Semin Thromb Hemost* 2004; **5**: 579
- 52. Moal V, Brunnet P, Dou L, Morange S, Sampol J, Berland Y. Impaired expression of glycoproteins on

- resting and stimulated platelets in uraemic patients. *Nephrol Dial Transplant* 2003; **18**: 1834.
- 53. Laxenaire MC, Charpentier C, Feldman L. Anaphylactoid reactions to colloid plasma substitutes: incidence, risk factors, mechanism. A French multicenter prospective study. *Ann Fr Anesth Reanim* 1994; **13**: 301.
- 54. Gloub R, Sorrento JJ Jr, Cantu R Jr, Nierman DM, Moideen A, Stein HD. Efficacy of albumin supplementation in the surgical intensive care unit: A prospective, randomised study. *Crit Care Med* 1994; 22: 613.
- 55. Boldt J, Heesen M, Müller M, Pabsdorf M, Hempelmann G. The effects of albumin versus hydroxyethyl starch on cardiorespiratory and circulatory variables in critically ill patients. *Anesth Analg* 1996; **83**: 254.
- 56. Cochrane Injuries Group Albumin Reviewers. Human albumin administration in critically ill patients: systematic review of randomised controlled trials. *BMJ* 1998; **317**: 235.
- 57. Finfer S, Bellomo R, Boyce N, French J, Myburgh J, Norton R; The SAFE Study Investigators. A comparison of albumin and saline for fluid resuscitation in the intensive care unit. *N Engl J Med* 2004; **350**: 2247.
- 58. Dawidson I, Peters P, Sagalowsky A, Abshier D, Coorpender L. The effect of intraoperative fluid management on the incidence of acute tubular necrosis. *Transplant Proc* 1987; **19** (1 Pt3): 2056.
- Willms CD, Dawidson IJ, Dickerman R, Drake D, Sandor ZF, Trevino G. Intraoperative blood volume expansion induces primary function after renal transplantation: a study of 96 paired cadaver kidneys. *Transplant Proc* 1991; 23 (1 Pt2): 1338.
- 60. Dawidson IJ, Sandor ZF, Coorpender L, *et al.* Intraoperative albumin administration affects the outcome of cadaver renal transplantation. *Transplantation* 1992; **53**: 774.
- 61. Weimar W, Geerlings W, Bijnen AB, *et al.* A controlled study on the effect of mannitol on immediate renal function after cadaver donor kidney transplantation. *Transplantation* 1983; **35**: 99.
- 62. Hoitsma AJ, Groenewoud AF, Berden JH, van Lier HJ, Koene RA. Important role for mannitol in the prevention of acute renal failure after cadaveric kidney transplantation. *Transplant Proc* 1987; **19** (1 Pt3): 2063.
- Lauzurica R, Teixido J, Serra A, et al. Hydration and mannitol reduce the need for dialysis in cadaveric kidney transplant recipients treated with CyA. Transplant Proc 1992; 24: 46.
- 64. Antunes N, Martinusso CA, Takiya CM, *et al.* Fructose-1,6 diphosphate as a protective agent for experimental ischemic renal failure. *Kidney Int* 2006; **69**: 68.
- 65. Dawidson I, Berglin E, Brynger H, Reisch J. Intravascular volumes and colloid dynamics in relation to fluid management in living related kidney donors and recipients. *Crit Care Med* 1987; **15**: 631.
- 66. Messmer KF. The use of plasma substitutes with special attention to their side effects. World J Surg 1987; 11: 69.

- 67. Bergman A, Andreen M, Blomback M. Plasma substitution with 3% dextran-60 in orthopaesic surgery: influence on plasma colloid osmotic pressure, coagulation parameters, immunoglobulines and other plasma constituents. *Acta Anaesthesiol Scand* 1990; **34**: 21.
- Wang DY, Forslund C, Persson U, Wiholm BE. Drug attributed anaphylaxis. *Pharmacoepidemiol Drug Saf* 1998; 7: 269.
- Walters BA, Van Wyck DB. Benchmarking iron dextran sensitivity: reactions requiring resuscitative medication in incident and prevalent patients. *Nephrol Dial Transplant* 2005: 20: 1438.
- 70. Bailie GR, Clark JA, Lane CE, Lane PL. Hypersensitivity reactions and deaths associated with intravenous iron preparations. *Nephrol Dial Transplant* 2005; **20**: 1443.
- Mailloux L, Swartz CD, Capizzi R, et al. Acute renal failure after administration of low-molecular weight dextran. N Engl J Med 1967; 277: 1113.
- Matheson NA, Diomi P. Renal failure after the administration of dextran 40. Surg Gynecol Obstet 1970; 131:
- 73. Biesenbach G, Kaiser W, Zazgornik J. Incidence of acute oligoanuric renal failure in dextran 40 treated patients with acute ischemic stroke stage III or IV. *Ren Fail* 1997; 19: 69.
- 74. Baron JF. Adverse effects of colloids on renal function. In: Vincent JL, ed. *Yearbook of Intensive Care and Emergency Medicine*. Berlin: Springer, 2000: 486–493.
- Kato A, Yonemura K, Matsushima H, Ikegaya N, Hishida A. Complication of oliguric acute renal failure in patients treated with low-molecular weight dextran. *Ren Fail* 2001; 23: 679.
- Halijamae H, Lindgren S. Fluid therapy: present controversies. In: Vincent JL, ed. *Yearbook of Intensive Care and Emergency Medicine*. Berlin: Springer, 2000: 429–442.
- 77. Nearman HS, Herman ML. Toxic effects of colloids in the intensive care unit. *Crit Care Clin* 1991; 7: 713.
- 78. Treib J, Baron JF, Grauer MT, Strauss RG. An international review of hydroxyethyl starches. *Intensive Care Med* 1999; **25**: 258.
- 79. Ickx BE, Bepperling F, Melot C, Schulman C, Van der Linden PJ. Plasma substitution effects of a new hydroxyethyl starch 130/0.4 compared with HES 200/0.5 during and after acute normovolaemic haemodilution. *Br J Anaesth* 2003; **91**: 196.
- 80. Kasper SM, Meinert P, Kampe S, *et al.* Large dose hydroxyethyl starch 130/0.4 does not increase blood loss and transfusion requirements in coronary artery bypass surgery compared with hydroxyethyl starch 200/0.5 at recommended dose. *Anesthesiology* 2003; **99**: 42.
- 81. Langeron O, Doelberg M, Ang ET, Bonnet F, Capdevila X, Coriat P. Voluven, a lower substituted novel hydroxyethyl starch (HES 130/0.4), causes fewer effects on coagulation in major orthopedic surgery than HES 200/0.5.

 Anesth Analg 2001; 92: 855.

- 82. Gallandat Huet RC, Siemons AW, Baus D, *et al.* A novel hydroxyethyl starch (Voluven) for effective perioperative plasma volume substitution in cardiac surgery. *Can J Anaesth* 2000; **47**: 1207.
- 83. Vogt N, Bothner U, Brinkmann A, de Petriconi R, Geogieff M. Post-operative tolerance to large-dose 6% HES 200/0.5 in major urological procedures compared with 5% human albumin. *Anaesthesia* 1999; **54**: 121.
- 84. Bothner U, Georgieff M, Vogt NH. Assessment of the safety and tolerance of 6% hydroxyethyl starch (200/0.5) solution: a randomized, controlled epidemiology study. *Anesth Analg* 1998; **86**: 850.
- 85. Boldt J, Zickmann B, Rapin J, Hammermann H, Dapper F, Hempelmann G. Influence of volume replacement with different HES-solutions on microcirculatory blood flow in cardiac surgery. *Acta Anaesthesiol Scand* 1994; 38: 432.
- Legendre C, Thervet E, Page B, Percheron A, Noel LH, Kreis H. Hydroxyethylstarch and osmotic-nephrosis-like lesions in kidney transplantation. *Lancet* 1993; 342: 248.
- Cittanova ML, Leblanc I, Legendre C, Mouquet C, Riou B, Coriat P. Effect of hydroxyethylstarch in brain-dead kidney donors on renal function in kidney-transplant recipients. *Lancet* 1996; 348: 1620.
- 88. Coronel B, Mercatello A, Colon S, Martin X, Moskovtchenko J. Hydroxyethylstarch and osmotic nephrosislike lesions in the kidney transplants. *Lancet* 1996; **348**: 1595
- 89. Ferraboli R, Malheiro PS, Abdulkader RC, Yu L, Sabbaga E, Burdmann EA. Anuric acute renal failure caused by dextran 40 administration. *Ren Fail* 1997; **19**: 303.
- Biesenbach G, Zazgornik J, Kaiser W, et al. Severe tubulopathy and kidney graft rupture after coadministration of mannitol and ciclosporin. Nephron 1992;
 62: 93.
- 91. Visweswaran P, Massin EK, Dubose TD Jr. Mannitolinduced acute renal failure. *J Am Soc Nephrol* 1997; **8**: 1028
- 92. Prerez-Perez AJ, Pazos B, Sobrado J, Gonzalez L, Gandara A. Acute renal failure following massive mannitol infusion. *Am J Nephrol* 2002; **22**: 573.
- Ahsan N, Palmer BF, Wheeler D, Greenlee RG Jr, Toto RG. Intravenous immunoglobulin-induced osmotic nephrosis. Arch Intern Med 1994; 154: 1985.
- 94. Haas M, Sonnenday CJ, Cicone JS, Rabb H, Montgomery RA. Isometric tubular vacuolization in renal allograft biopsy specimens of patients receiving low-dose intravenous immunoglobulin for a positive cross match. *Transplantation* 2004; **78**: 549.
- 95. Cantu TG, Hoehn-Saric EW, Burgess KM, Racusen L, Scheel PJ. Acute renal failure associated with immunoglobulin therapy. *Am J Kidney Dis* 1995; **25**: 228.
- Moran M, Kapsner C. Acute renal failure associated with elevated plasma oncotic pressure. N Engl J Med 1987; 317: 150.

- 97. Suttner S, Boldt J. Volume replacement with hydroxyethyl starch: is there an influence on kidney function? *Anasthesiol Intensivmed Notfallmed Schmerzther* 2004; **39**: 71.
- 98. Deman A, Peeters P, Sennesael J. Hydroxyethyl starch does not impair immediate renal function in kidney transplant recipients: a retrospective, multicentre analysis. *Nephrol Dial Transplant* 1999; **14**: 1517.
- 99. Johnston PA, Bernard DB, Perrin NS, Levinsky NG. Prostaglandins mediate the vasodilatory effect of mannitol in the hypoperfused rat kidney. *J Clin Invest* 1981; **68**: 127.
- 100. Bratall S, Folmerz P, Hansson R, et al. Effects of oxygen free radical scavengers, xanthine oxidase inhibition and calcium entry-blockers on leakage of albumin after ischemia. An experimental study in rabbit kidneys. Acta Physiol Scand 1988; 134: 35.
- 101. Tay M, Cooper WD, Vassilou P, Glasgow EF, Baker MS, Pratt L. The inhibitory action of oxygen radical scavengers on proteinuria and glomerular heparan sulphate loss in the isolated perfused kidney. *Biochem Int* 1990; 20: 767.
- Richards KF, Belnap LP, Stevens LE. Mannitol reduces ATN in cadaveric allografts. *Transplant Proc* 1989; 21 (1 Pt2): 1228.
- 103. Grino JM, Miravittles R, Castelao AM, *et al.* Flush solution with mannitol in the prevention of post-transplant renal failure. *Transplant Proc* 1987; **19**: 4140.
- 104. Porras I, Gonzalez-Posada JM, Losada M, *et al.* A multivariate analysis of the risk factors for posttransplant renal failure: beneficial effect of a flush solution with mannitol. *Transplant Proc* 1992; **24**: 52.
- 105. Bugge JF, Hartmann A, Ones S, Bentdal O, Stentstrom J. Immediate and early renal function after living donor transplantation. *Nephrol Dial Transplant* 1999; **14**: 389.
- 106. Tiggeler RG, Berden JH, Hoitsma AJ, Koene RA. Prevention of acute tubular necrosis in cadaveric kidney transplantation by the combined use of mannitol and moderate hydration. *Ann Surg* 1985; 201: 246.
- 107. Van Valenberg PL, Hoitsma AJ, Tiggeler RG, Berden JH, van Lier HJ, Koene RA. Mannitol as an indispensable constituent of an intraoperative hydration protocol for the prevention of acute renal failure after renal cadaveric transplantation. *Transplantation* 1987; 44: 784.
- 108. Caldwell JE, Cook DR. Kidney transplantation. In: Cook DR, Davis PJ, eds. *Anesthetic Principles of Organ Transplantation*. New York: Raven Press, 1994.
- Lachance SL, Barry JM. Effect of furosemide on dialysis requirement following cadaveric kidney transplantation. *J Urol* 1985; 133: 950.
- Shilliday IR, Quinn KJ, Allison ME. Loop diuretics in the management of acute renal failure: a prospective, doubleblind, placebo-controlled, randomised study. *Nephrol Dial Transplant* 1997; 12: 2592.
- Elsasser S, Schachinger H, Strobel W. Adjunctive drug treatment in severe hypoxic respiratory failure. *Drugs* 1999; 58: 429.

- 112. Bellomo R, Raman J, Ronco C. Intensive care unit management of the critically ill patient with fluid overload after open heart surgery. *Cardiology* 2001; **96**: 169.
- 113. Venkataram R, Kellum JA. The role of diuretic agents in the management of acute renal failure. *Contrib Nephrol* 2001; **132**: 158.
- 114. Cantarovich F, Rangoonwala B, Lorenz H, Verho M, Esnault VL; High Dose Furasemide in Acute Renal Failure Study Group. High-dose furosemide for established ARF: a prospective, randomised double-blind, placebo-controlled. multicenter trial. *Am J Kidney Dis* 2004; **44**: 402.
- 115. Lassnigg A, Donner E, Grubhofer G, Presterl E, Druml W, Hiesmayr M. Lack of renoprotective effects of dopamine and furosemide during cardiac surgery. *J Am Soc Nephrol* 2000; **11**: 97.
- Birtch A, Zakheim RM, Jones LG, Barger AC. Redistribution of renal blood flow produced by furosemide and ethacrynic acid. *Circ Res* 1967; 21: 869.
- 117. Pienaar H, Schwartz I, Roncone A, Lotz Z, Hickman R. Function of kidney grafts from brain-dead donor pigs. The influence of dopamine and trijodthyronine. *Transplantation* 1990; **50**: 580.
- 118. Walaszewski J, Rowinski W, Chmura A, et al. Decreased incidence of acute tubular necrosis after cadaveric donor transplantation due to lidocaine donor pretreatment and low-dose dopamine infusion in the recipient. Transplant Proc 1988; 20: 913.
- 119. Carmellini M, Romagnoli J, Giulianotti PC, *et al.* Dopamine lowers the incidence of delayed graft function in transplanted kidney patients treated with cyclosporine A. *Transplant Proc* 1994; **26**: 2626.
- 120. Grundmann R, Kindler J, Meider G, Stowe H, Sieberth HG, Pichlmaier H. Dopamine treatment of human cadaver kidney graft recipients: a prospectively randomized trial. *Klin Wochenschr* 1982; **60**: 193.
- 121. Sandberg J, Tyden G, Groth CG. Low-dose dopamine infusion following cadaveric renal transplantation: no effect on the incidence of ATN. *Transplant Proc* 1992; **24**: 357.
- 122. Kadieva VS, Friedman L, Margolius LP, Jackson SA, Morrell DF. The effect of dopamine on graft function in patients undergoing renal transplantation. *Anesth Analg* 1993; **76**: 362.
- 123. Ferguson CJ, Hillis AN, Williams JD, Griffin PJ, Salaman JR. Calcium-channel blockers and other factors influencing delayed function in renal allografts. *Nephrol Dial Transplant* 1990; **5**: 816.
- Dalton RS, Webber JN, Cameron C, et al. Physiologic impact of low-dose dopamine on renal function in the early post renal transplant period. *Transplantation* 2005; 79: 1561.
- 125. O'Dair J, Evans L, Rigg KM, Shehata M. Routine use of renal-dose dopamine during living donor nephrectomy has no beneficial effect to either donor or recipient. *Transplant Proc* 2005; **37**: 637.

- 126. Schnuelle P, Lorenz D, Mueller A, Trede M, van der Woude FJ. Donor catecholamine use reduces acute allograft rejection and improves graft survival after renal cadaveric transplantation. *Kidney Int* 1999; **56**: 738.
- 127. Sutherland FR, Bloembergen W, Mohamed M, Ostbye T, Klar N, Lazarovits AI. Initial non function in cadaveric renal transplantation. *Can J Surg* 1993; **36**: 141.
- 128. Schnuelle P, Berger S, de Boer J, Persijn G, van der Woude FJ. Effects of catecholamine application to brain-dead donors on graft survival in solid organ transplantation. *Transplantation* 2001; **72**: 455.
- 129. Schnuelle P, Yard BA, Braun C, *et al.* Impact of donor dopamine on immediate graft function after kidney transplantation. *Am J Transplant* 2004; **4**: 419.
- 130. van der Hoeven JA, Molema G, Ter Horst GJ, et al. Relationship between duration of brain death and hemodynamic (in)stability on progressive dysfunction and increased immunologic activation of donor kidneys. Kidney Int 2003; 64: 1874.
- 131. Yard B, Beck G, Schnuelle P, *et al.* Prevention of cold-preservation injury of cultured endothelial cells by cate-cholamines and related compounds. *Am J Transplant* 2004; **4**: 22.
- 132. Marik PE, Iglesias J. Low-dose dopamine does not prevent acute renal failure in patients with septic shock and oliguria. NORASEPT II Study Investigators. *Am J Med* 1999; 107: 392.
- 133. Bellomo R, Chapman M, Finfer S, Hickling K, Myburgh J. Low dose dopamine in patients with early renal dysfunction: a placebo-controlled randomised trial. Australian and New Zealand Intensive Care Society (ANZICS) Clinical Trials Group. *Lancet* 2000; **356**: 2139.
- 134. Kellum JA, M Decker J. Use of dopamine in acute renal failure: a meta-analysis. *Crit Care Med* 2001; **29**: 1526.
- 135. Marik PE. Low-dose dopamine: a systemic review. *Intensive Care Med* 2002; **28**: 877.
- 136. Vanholder R, Heering P, Loo AV, et al. Reduced incidence of acute renal graft failure in patients treated with peritoneal dialysis compared with hemodialysis. Am J Kidney Dis 1999; 33: 934.
- 137. Snyder JJ, Kasiske BL, Gilbertson DT, Collins AJ. A comparison of transplant outcomes in peritoneal and hemodialysis patients. *Kidney Int* 2002; **62**: 1423.
- 138. Hakim RM, Wingard RL, Parker RA. Effect of the dialysis membrane in the treatment of patients with acute renal failure. *N Engl J Med* 1994; **331**: 1338.
- 139. Himmelfarb J, Tolkoff Rubin N, Chandran P, Parker RA, Wingard RL, Hakim R. A multicenter comparison of dialysis membranes in the treatment of acute renal failure requiring dialysis. *J Am Soc Nephrol* 1998; **9**: 257.
- 140. Valeri A, Radhakrishman J, Ryan R, Powell D. Biocompatible dialysis membranes and acute renal failure: a study in post-operative acute tubular necrosis in cadaveric renal transplant recipients. Clin Nephrol 1996; 46: 402.

- 141. Romao JE Jr, Abensur H, de Castro MC, Ianherz LE, Massola VC, Sabbaga E. Effect of dialyser biocompatibility on recovery from acute renal failure after cadaver renal transplantation. *Nephrol Dial Transplant* 1999; **14**: 709.
- 142. Woo YM, Craig AM, King BB, *et al.* Biocompatible membranes do not promote graft recovery following renal transplantation. *Clin Nephrol* 2002; **57**: 38.
- 143. Hayes MA, Timmins AC, Yau EH, Palazzo M, Hinds CJ, Watson D. Elevation of systemic oxygen delivery in the treatment of critically ill patients. *N Engl J Med* 1994; **330**: 1717.
- 144. Gattinoni L, Brazzi L, Pelosi P, *et al.* A trial of goal-oriented hemodynamic therapy in critically ill patients. SvO2 Collaborative Group. *N Engl J Med* 1995; **333**: 1025.
- 145. Heyland DK, Cook DJ, King D, Kernerman P, Brun-Buisson C. Maximizing oxygen delivery in critically ill patients: a methodologic appraisal of the evidence. Crit Care Med 1996; 24: 517.
- 146. Esson ML, Schrier RW. Diagnosis and treatment of acute tubular necrosis. *Ann Intern Med* 2002; **137**: 744.
- Arieff AI. Fatal postoperative pulmonary edema: pathogenesis and literature review. Chest 1999; 115: 1371.
- 148. Brandstrup B, Tonnesen H, Beier-Holgersen R, *et al.* Effects of intravenous fluid restriction on postoperative complications: comparison of two perioperative fluid regimens: a randomized assessor-blinded multicenter trial. *Ann Surg* 2003; **238**: 641.
- 149. Lowell JA, Schifferdecker C, Driscoll DF, Benotti PN, Bistrian BR. Postoperative fluid overload: not a benign problem. *Crit Care Med* 1990; **18**: 728.

- 150. Gill JS, Tonelli M, Johnson N, Kiberd B, Landsberg D, Peireira BJ. The impact of waiting time and comorbid conditions on the survival benefit of kidney transplantation. *Kidney Int* 2005; 68: 2345.
- 151. Gill JS, Peireira BJ. Death in the first year after kidney transplantation: implications for patients on the transplant waiting list. *Transplantation* 2003; **75**: 113.
- 152. Luciani J, Frantz P, Thibault P, *et al.* Early anuria prevention in human kidney transplantation. Advantage of fluid load under pulmonary arterial pressure monitoring during surgical period. *Transplantation* 1979; **28**: 308.
- 153. Joshi GP. Intraoperative fluid restriction improves outcome after major elective gastrointestinal surgery. *Anesth Analg* 2005; **101**: 601.
- 154. Holte K, Sharrock NE, Kehlet H. Pathophysiology and clinical implications of perioperative fluid excess. Br J Anaesth 2002; 89: 622.
- 155. Kita T, Mammoto T, Kishi Y. Fluid management and postoperative respiratory disturbances in patients with transthoracic esophagectomy for carcinoma. *J Clin Anesth* 2002; **14**: 252.
- 156. Neal JM, Wilcox RT, Allen HW, Low DE. Near total esophagectomy: the influence of standardized multimodal management and intraoperative fluid restriction. *Reg Anesth Pain Med* 2003; **28**: 328.
- 157. Lobo DN, Bostock KA, Neal KR, Perkins AC, Rowlands BJ, Allison SP. Effect of salt and water balance on recovery of gastrointestinal function after elective colonic resection: a randomized controlled trial. *Lancet* 2002; **359**: 1812