

Novel *Divavirus* (the family *Betaflexiviridae*) and *Mitovirus* (the family *Narnaviridae*) species identified in basil (*Ocimum basilicum*)

C. J. GOH¹, D. PARK¹, H. KIM¹, F. SEBASTIANI², Y. HAHN^{1*}

¹Department of Life Science, Research Center for Biomolecules and Biosystems, Chung-Ang University, Seoul 06974, South Korea;

²Institute for Sustainable Plant Protection, Department of Biology, Agriculture and Food Sciences, The National Research Council of Italy, Sesto Fiorentino, Italy

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Summary. – Transcriptome data obtained from a plant sample often contain a large number of reads that are derived from associated RNA virus genomes that were co-isolated during RNA preparation. These virus-derived reads can be assembled into a novel plant RNA genome sequence. Here, a basil (*Ocimum basilicum*) transcriptome dataset was analyzed to identify two new RNA viruses, which were named *Ocimum basilicum* RNA virus 1 (ObRV1) and *Ocimum basilicum* RNA virus 2 (ObRV2). A phylogenetic analysis of the ObRV1 RNA-dependent RNA polymerase (RdRp) motif indicated that ObRV1 is a novel species of the genus *Divavirus* of the family *Betaflexiviridae*. ObRV1 is the fourth *divavirus* species to be identified. The ObRV2 RdRp motif showed sequence similarity to viruses of the genus *Mitovirus* of the family *Narnaviridae*, which infect fungal mitochondria. Although most of the known mitoviruses do not produce a functional RdRp using the plant mitochondrial genetic code, the ObRV2 encodes a full-length RdRp using both the fungal and plant mitochondrial genetic codes.

Keywords: basil; *Ocimum basilicum* RNA virus 1; *Ocimum basilicum* RNA virus 2; *Divavirus*; *Mitovirus*

Introduction

Basil (*Ocimum basilicum* L.) is a popular herb that belongs to the genus *Ocimum* of the family *Lamiaceae*. It is widely cultivated in Asia, the Mediterranean region, and the United States of America for commercial and industrial purposes (Dudai *et al.*, 2002; Suppakul *et al.*, 2003; Chiang *et al.*, 2005). Basil is also used as a medicinal, antiviral, antibacterial, and antimicrobial agent, with its essential oils being known to have many beneficial compounds, such as monoterpenoids, which act against pathogens such as virus, bacteria, and fungi (Edris and Farrag, 2003; Suppakul *et al.*, 2003; Chiang *et al.*, 2005).

Transcriptome analyses of basil have been conducted to investigate its genetic features, such as the genes involved in secondary metabolism and production of unique essential oils (Rastogi *et al.*, 2014; Torre *et al.*, 2016). The plant transcriptome data often contain sequences derived from RNA viruses, which can be identified to be novel plant RNA viruses (Nouri *et al.*, 2016; Visser *et al.*, 2016; Park and Hahn, 2017a,b). In this study, a previously reported transcriptome dataset collected from red and green morphs of sweet basil was analyzed and potential sequences of two novel RNA viruses, a *divavirus* and a *mitovirus*, were identified (Torre *et al.*, 2016).

Currently, only three species are reported in the genus *Divavirus* of the family *Betaflexiviridae* (Wylie and Jones, 2011; Wylie *et al.*, 2013). The *Betaflexiviridae* viruses have a linear, single-stranded, positive-sense RNA genomic segment 6.5–9.0 kb in size, which encodes 2–6 proteins, including the RNA-dependent RNA polymerase (RdRp) and several other proteins depending on the genus (Adams, 2012; Igori *et al.*, 2016).

*Corresponding author. E-mail: hahny@cau.ac.kr; phone: +82-2-820-5812.

Abbreviations: ObRV1 = *Ocimum basilicum* virus 1; ObRV2 = *Ocimum basilicum* virus 2; ORF = open reading frame; RdRp = RNA-dependent RNA polymerase; SRA = Sequence Read Archive

Table 1. Summary of RNA viruses identified in this study

Acronym	Full name	Acc. No.	Length (nt)	ORF	Position	Length (aa)
ObRV1	Ocimum basilicum RNA virus 1	MF196913	6930	Replicase/Coat protein	44–6721	2255
				Movement protein	5050–6219	389
ObRV2	Ocimum basilicum RNA virus 2	MF196914	2784	Replicase	347–2629	760

Viruses of the family *Narnaviridae*, to which the genus *Mitovirus* belongs, have an unencapsidated, linear, single-stranded, positive-sense RNA genome 2.5–2.9 kb in size, which encodes a single open reading frame (ORF) for the replicase containing the RdRp motif (Hillman and Cai, 2013). Mitoviruses replicate in the cytosol or mitochondria of infected fungi and have been mainly isolated from plant-pathogenic fungi (Ghabrial and Suzuki, 2009; Xie and Ghabrial, 2012).

Materials and Methods

The previously reported basil transcriptome dataset (a total of 13 gigabase pairs), which was isolated from red and green sweet basils, was analyzed in this study (Torre *et al.*, 2016). The transcriptome data is available in the Sequence Read Archive (SRA) of the National Center for Biotechnology Information (NCBI) under the accession number SRA313233. Sickle program (version 1.33; <https://github.com/najoshi/sickle>) was used to collect high-quality reads with the option “-q 30 -l 55.” The SPAdes Genome Assembler (version 3.10.1) was used to assemble the RNA-seq reads into contigs (Bankevich *et al.*, 2012).

The RdRp motif sequences of known RNA viruses were retrieved from the Pfam database (release 30.0; <http://pfam.xfam.org>). A total of 345 non-redundant RdRp motif sequences were collected from 19 RdRp families: their accession numbers are PF00602, PF00603, PF00604, PF00680, PF00946, PF00972, PF00978, PF00998, PF02123, PF03431, PF04196, PF04197, PF05788, PF05919, PF07925, PF08467, PF08716, PF08717, and PF12426.

The assembled basil transcript contigs were compared with the Pfam-derived RdRp sequences, using BLAST (Altschul *et al.*, 1990) with the parameter “-evalue e-5.” Matched contigs 1000 bp or longer were selected. Annotation of contigs and identification of closely related viruses were carried out by BLAST searches of all viral genomes and proteins at the NCBI website (<https://www.ncbi.nlm.nih.gov/BLAST>). The basil RNA-seq reads were mapped to the viral contigs, using BWA, and variants were called using Samtools/Bcftools (Li and Durbin, 2009; Li, 2011).

For phylogenetic analysis, the RdRp motif-containing sequences were multiply aligned using the MUSCLE program (Edgar, 2004). The aligned region spanning the core RdRp motif defined by a Pfam analysis was extracted and subjected to phylogenetic analysis. A phylogenetic tree was constructed by the neighbor-joining method implemented in the ClustalW2 software (version 2.1) (Larkin *et al.*, 2007).

Results and Discussion

Genome sequences of two new viruses (Table 1) were identified by analysis of the basil transcriptome data (SRA accession number SRA313233) (Torre *et al.*, 2016). One viral contig, named Ocimum basilicum RNA virus 1 (ObRV1), was most similar to those of divaviruses (Table 2) and considered a novel member of the genus *Divavirus*. The other contig, named Ocimum basilicum RNA virus 2 (ObRV2), showed sequence similarity to those of mitoviruses (Table 2), suggesting that it belongs to the genus *Mitovirus*.

Ocimum basilicum RNA virus 1 (ObRV1)

The first of the two viral contigs discovered in the basil transcriptome was named Ocimum basilicum RNA virus 1 (ObRV1) and showed sequence similarity to those of plant RNA viruses of the genus *Divavirus* of the family *Betaflexiviridae*. There were sequence variations at 33 positions when the basil RNA-seq reads mapped to the ObRV1 genome were analyzed, indicating that the contig is a composite sequence derived from at least two closely related viruses (Supplementary Table S1).

The ObRV1 RNA genome is 6,930 base pairs (bp) long and predicted to have two overlapping ORFs. ORF1 encodes a 2,255-amino acid (aa)-long RdRp motif-containing protein and ORF2 a 389-aa-long movement protein (Table 1). A domain analysis using Pfam showed that the ObRV1 ORF1 protein contains four known domains: viral methyltransferase (PF01660) at aa 43–357, viral RNA helicase (PF01443) at aa 858–1,119, RdRp (PF00978) at aa 1,255–1,653, and trichovirus coat protein (PF05892) at aa 2,039–2,223. These genome and domain organizations are the shared features of viruses in the genera *Divavirus* and *Capillovirus* (Jelkmann, 1995; Wylie and Jones, 2011).

The ObRV1 RdRp showed 30–60% aa sequence identities with RdRps of the viruses that belong to the family *Betaflexiviridae* (Table 2). Among them, Hardenbergia virus A (HarVA), Diuris virus A (DiVA), and Diuris virus B (DiVB) were the top three similar viruses with aa sequence identities of 50–60%. These three viruses are species in the genus *Divavirus*, suggesting that ObRV1 is a new divavirus.

A phylogenetic analysis using multiply aligned RdRp motif sequences confirmed that ObRV1 is a new species of the genus *Divavirus* (Supplementary Fig. S1 and Fig. 1a).

Table 2. Identities among the RdRp sequences of ObRV1, ObRV2, and their respective related viruses

	Acronym	Full name	Acc. No. ^a	Sequence identity ^b	
ObRV1	HarVA	Hardenbergia virus A	YP_004376201.1	1341/2237 (60%)	
	DiVA	Diuris virus A	YP_006905850.1	1243/2248 (55%)	
	DiVB	Diuris virus B	YP_006905848.1	1196/2256 (53%)	
	CVA	Cherry virus A	ANE06570.1	357/949 (38%)	
	CVA	Cherry virus A	APG53775.1	331/848 (39%)	
	PCMV	Peach chlorotic mottle virus	YP_001497153.1	336/928 (36%)	
	ApLV	Apricot latent virus	ADT91605.1	334/910 (37%)	
	NeLV	Nerine latent virus	YP_009174681.1	335/1030 (33%)	
	NSV	Narcissus symptomless virus	YP_842438.1	336/1030 (33%)	
	CTLaV	Cherry twisted leaf associated virus	AHJ80270.1	326/840 (39%)	
	CTLaV	Cherry twisted leaf associated virus	AHJ80314.1	326/859 (38%)	
	CNRMV	Cherry necrotic rusty mottle virus	ALP45953.1	338/909 (37%)	
	CRMaV	Cherry rusty mottle associated virus	AHA59466.2	323/846 (38%)	
	CRMaV	Cherry rusty mottle associated virus	AHJ80324.1	317/839 (38%)	
	CRMaV	Cherry rusty mottle associated virus	AKN20442.1	347/971 (36%)	
	CGRMV	Cherry green ring mottle virus	AFU54620.1	318/825 (39%)	
	BanMMV	Banana mild mosaic virus	NP_112029.1	321/870 (37%)	
	ObRV2	BcMV1	Botrytis cinerea mitovirus 1	CEZ26300.1	165/506 (33%)
		FpMV4	Fusarium poae mitovirus 4	YP_009272901.1	104/224 (46%)
		OMV3a	Ophiostoma mitovirus 3a	NP_660176.1	101/241 (42%)
SsMV3		Sclerotinia sclerotiorum mitovirus 3	YP_009182164.1	110/283 (39%)	
MpMV3		Macrophomina phaseolina mitovirus 3	AMM45292.1	100/273 (37%)	
RsMV-17		Rhizoctonia solani mitovirus 17	ANR02693.1	91/230 (40%)	
RsMV-15		Rhizoctonia solani mitovirus 15	ALD89120.1	100/276 (36%)	
RsMV-14		Rhizoctonia solani mitovirus 14	ALD89119.1	102/297 (34%)	
HetMV1		Heterobasidion mitovirus 1	AIF33766.2	158/547 (29%)	
COV		Clitocybe odora virus	YP_005352912.1	174/577 (30%)	
BcMV3		Botrytis cinerea mitovirus 3	YP_009182161.1	113/304 (37%)	
FpMV3		Fusarium poae mitovirus 3	YP_009272900.1	87/219 (40%)	
SlaMV5		Soybean leaf-associated mitovirus 5	ALM62240.1	87/233 (37%)	
RsMV-19		Rhizoctonia solani mitovirus 19	ANR02690.1	87/221 (39%)	
BsMV1		Buergenerula spartinae mitovirus 1	AHY03257.1	91/242 (38%)	
CpMV1		Cryphonectria parasitica mitovirus 1-NB631	NP_660174.1	79/216 (37%)	
RcMV		Rhizoctonia cerealis mitovirus	AIT71973.1	89/241 (37%)	
SsMV15		Sclerotinia sclerotiorum mitovirus 15	AHF48631.1	81/208 (39%)	
HfMV1		Hymenoscyphus fraxineus mitovirus 1	AIU44705.1	81/208 (39%)	
SsMV16		Sclerotinia sclerotiorum mitovirus 16	AHF48632.1	89/240 (37%)	
SsMV2		Sclerotinia sclerotiorum mitovirus 2	AHX84129.1	88/232 (38%)	
SlaMV3		Soybean leaf-associated mitovirus 3	ALM62243.1	76/203 (37%)	
OMV5		Ophiostoma mitovirus 5	NP_660180.1	82/213 (38%)	
SlaMV2		Soybean leaf-associated mitovirus 2	ALM62242.1	78/209 (37%)	
FgMV1		Fusarium globosum mitovirus 1	YP_009126872.1	110/367 (30%)	
TbMV		Thielaviopsis basicola mitovirus	AAZ99833.1	127/435 (29%)	

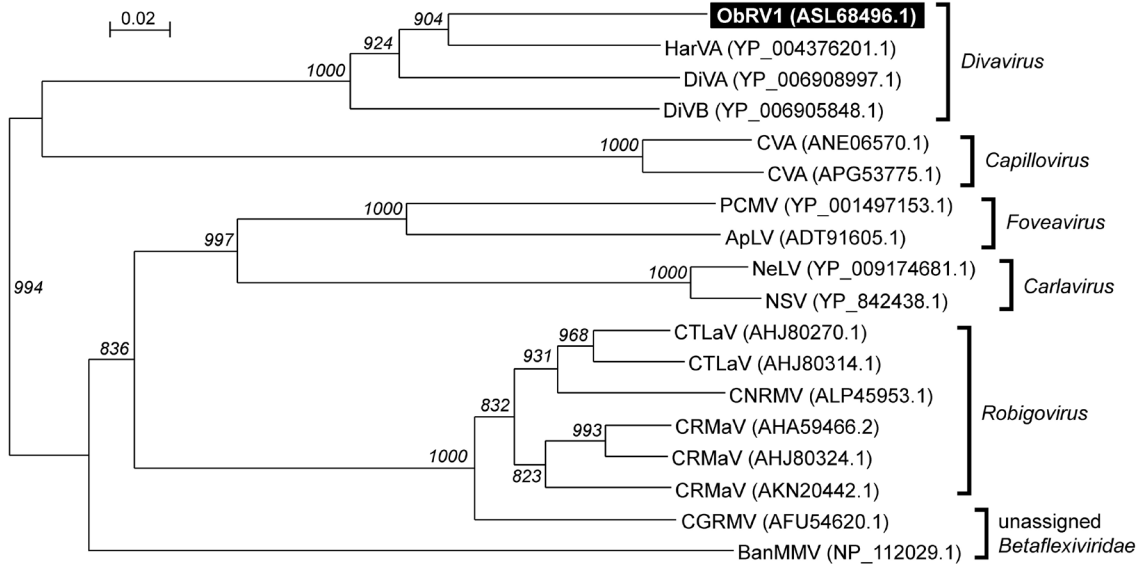
^aAcc. No. for the RdRp motif-containing protein sequence. ^bAmino acid sequence identities in a format of "identical residues/aligned length (% identity)."

The ObRV1 was clustered with the three known divaviruses, with HarVA being the closest species. Currently, there are three known species of the genus *Divavirus* (Wylie and Jones, 2011; Wylie *et al.*, 2013); ObRV1 is the fourth divavirus species to be reported.

Ocimum basilicum RNA virus 2 (ObRV2)

The second viral contig was named *Ocimum basilicum* RNA virus 2 (ObRV2) and showed sequence similarity to those of viruses of the genus *Mitovirus* of the family *Nar-*

(a) **ObRV1**



(b) **ObRV2**

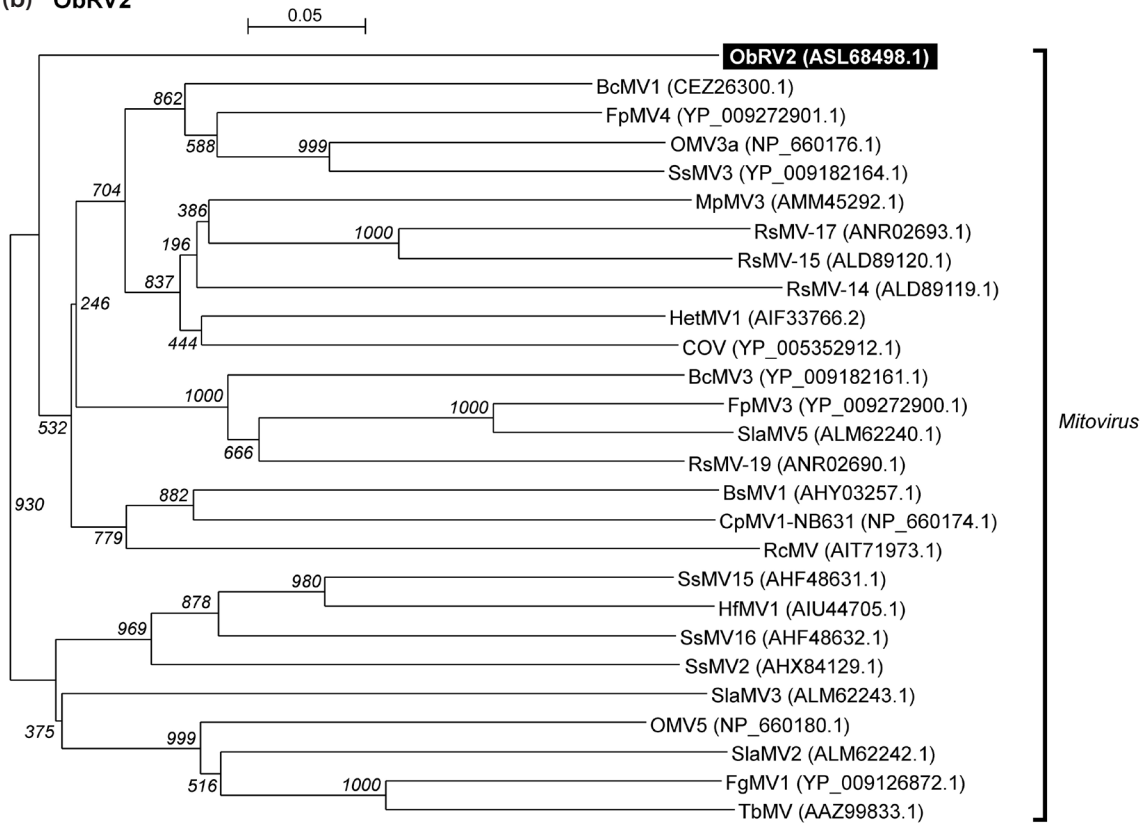


Fig. 1

Phylogenetic analysis of newly identified basil RNA viruses

Phylogenetic trees constructed based on the amino acid sequences of the RdRp motifs of (a) ObRV1 and (b) ObRV2 are presented. The bootstrap percentages calculated from 1,000 bootstrap replicates are shown at the nodes. See Table 2 for the full names of the viruses and Supplementary Figs. S1 (ObRV1) and S2 (ObRV2) for the multiple sequence alignments of the amino acid sequences of the RdRp motifs.

naviridae. Given that mitoviruses generally infect fungi, it was uncertain whether this mitovirus genome was isolated directly from basil cells or fungal cells that infected basil (Hillman and Cai, 2013).

The ObrV2 genome is 2,784-bp long and predicted to have a single ORF encoding a 760-aa-long protein when it was translated using both the fungal mitochondrial and plant (standard) genetic codes (Table 1). A Pfam analysis indicated that the ORF contains a mitovirus-type RdRp motif (PF05919) at aa 268–579. There were sequence variations at 61 positions, indicating that the ObrV2 genome contig is also a composite sequence derived from two or more clones (Supplementary Table S2).

The ObrV2 RdRp motif-containing protein showed about 30–40% similarity to those of other mitoviruses (Table 2). A phylogenetic analysis based on a multiple sequence alignment of the RdRp domain sequences placed ObrV2 as a distinct clade within the genus *Mitovirus* (Fig. 1b and Supplementary Fig. S2).

Most of the fungal mitoviruses replicate within the mitochondria of fungal cells and are unable to produce functional RdRps in plant cells because the genetic codes of fungal mitochondria and plants are different. For instance, the UGA codon, which is a stop codon in the standard genetic code, encodes for tryptophan in fungal mitochondria (Nibert, 2017). However, the ObrV2 ORF encodes a full-length RdRp using the plant nuclear and mitochondrial genetic codes, suggesting that ObrV2 may be able to replicate in both fungal and plant cells.

Interestingly, there are mitovirus-derived sequences in many plant nuclear and mitochondrial genomes, some of which are expressed as plant genes (Bruenn *et al.*, 2015; Xu *et al.*, 2015). Because the ObrV2 sequence was found in a plant transcriptome and encodes a full-length protein, it could also be derived from a mitovirus-like gene integrated in the basil nuclear or mitochondrial genome. However, most plant mitovirus-like genes are derived from the RdRp domain and are relatively shorter than those encoding the mitovirus RdRps; plant mitovirus-like genes encode for proteins about 500-aa long or shorter, while the mitovirus RdRps are 700-aa long or longer. The ObrV2 RdRp is 760-aa long. Therefore, it is highly likely that the ObrV2 sequence represents a mitovirus that infects the basil or a basil-associated fungus.

In conclusion, basil transcriptome data were analyzed to identify viral genome sequences representing two novel viral species of the genera *Divavirus* and *Mitovirus*. The method used in this study can be applied to other plant transcriptome data to discover plant viral genome sequences co-isolated with their host RNAs.

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Supplementary information is available in the online version of the paper.

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Supplementary information

Novel *Divavirus* (the family *Betaflexiviridae*) and *Mitovirus* (the family *Narnaviridae*) species identified in basil (*Ocimum basilicum*)

C. J. GOH¹, D. PARK¹, H. KIM¹, F. SEBASTIANI², Y. HAHN^{1*}

¹Department of Life Science, Research Center for Biomolecules and Biosystems, Chung-Ang University, Seoul 06974, South Korea;

²Institute for Sustainable Plant Protection, Department of Biology, Agriculture and Food Sciences, The National Research Council of Italy, Sesto Fiorentino, Italy

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Supplementary Table S1. Sequence variation of ObRV1

Position	Con ^a	Alt ^b	Quality ^c	Con #	Alt #	Con %	Alt %
274	A	G	222	2212	2792	44.2	55.8
535	G	A	222	7568	4917	60.6	39.4
709	T	C	222	6593	4802	57.9	42.1
871	G	A	222	6262	5003	55.6	44.4
904	A	G	222	5955	4258	58.3	41.7
907	A	T	222	5978	4217	58.6	41.4
1330	T	C	222	7713	4836	61.5	38.5
1421	T	C	222	8349	5928	58.5	41.5
1423	G	T	222	8496	5927	58.9	41.1
1912	G	A	222	7838	4337	64.4	35.6
2305	T	C	222	7486	4512	62.4	37.6
2407	C	T	222	8855	5501	61.7	38.3
2758	T	C	222	7397	4396	62.7	37.3
2761	A	G	222	7580	4294	63.8	36.2
3403	G	A	222	8320	6555	55.9	44.1
3689	T	C	222	8390	6545	56.2	43.8
4036	A	G	222	9544	7299	56.7	43.3
4222	G	A	222	9823	8396	53.9	46.1
4243	T	A	222	9350	7276	56.2	43.8
4576	A	G	222	9959	7574	56.8	43.2
4645	T	C	222	10088	8018	55.7	44.3
4978	A	G	222	11797	8422	58.3	41.7
4984	G	A	222	12145	8704	58.3	41.7
5130	T	C	222	11593	8383	58.0	42.0
5253	T	C	222	15217	11009	58.0	42.0
5307	C	T	222	15145	10481	59.1	40.9
5343	C	T	222	12969	9216	58.5	41.5
5511	G	A	222	13535	9896	57.8	42.2
5539	A	G	222	12512	9866	55.9	44.1
5785	C	T	222	15605	11631	57.3	42.7
5905	C	T	222	13495	11168	54.7	45.3
6511	G	A	222	12695	8850	58.9	41.1
6551	C	T	222	15251	11238	57.6	42.4

^aSequence of the assembled contig; ^bAlternative sequence observed in RNA-seq reads; ^cPhred-scaled quality score for the assertion made in Alt by BCFtools.

Supplementary Table S2. Sequence variation of ObRV2

Position	Con ^a	Alt ^b	Quality ^c	Con #	Alt #	Con %	Alt %
335	CAA	CAAA	183	21	18	53.8	46.2
352	A	T	181	21	15	58.3	41.7
355	A	T	179	21	15	58.3	41.7
442	C	T	222	35	32	52.2	47.8
445	T	C	222	39	30	56.5	43.5
451	A	G	222	42	33	56.0	44.0
454	T	C	222	47	35	57.3	42.7
583	C	T	212	42	22	65.6	34.4
601	T	C	222	41	25	62.1	37.9
676	C	T	222	39	30	56.5	43.5
691	C	T	222	40	22	64.5	35.5
715	C	T	222	36	24	60.0	40.0
736	A	G	222	30	27	52.6	47.4
763	G	A	222	38	27	58.5	41.5
776	C	G	222	40	30	57.1	42.9
835	C	T	222	31	26	54.4	45.6
836	G	C	222	30	26	53.6	46.4
889	G	A	222	45	52	46.4	53.6
911	C	A	222	48	43	52.7	47.3
912	G	A	222	47	43	52.2	47.8
926	A	C	222	52	44	54.2	45.8
940	C	T	222	55	39	58.5	41.5
961	T	C	222	56	31	64.4	35.6
967	G	A	222	50	25	66.7	33.3
1006	C	G	219	34	21	61.8	38.2
1007	A	C	217	34	21	61.8	38.2
1065	C	G	222	30	20	60.0	40.0
1066	C	T	221	31	20	60.8	39.2
1085	G	A	222	32	26	55.2	44.8
1102	C	T	222	30	29	50.8	49.2
1178	C	A	222	38	24	61.3	38.7
1267	A	T	222	47	35	57.3	42.7
1276	G	A	222	49	33	59.8	40.2
1321	A	G	222	50	32	61.0	39.0
1330	T	C	222	49	36	57.6	42.4
1465	T	C	222	25	21	54.3	45.7
1486	C	T	12.9425	45	6	88.2	11.8
1504	G	A	222	26	26	50.0	50.0
1510	T	C	222	25	28	47.2	52.8
1594	A	T	222	23	21	52.3	47.7
1600	C	T	221	22	17	56.4	43.6
1732	T	C	222	52	42	55.3	44.7
1775	G	A	172	48	40	54.5	45.5
1777	A	C	157	49	40	55.1	44.9
1810	G	A	222	30	32	48.4	51.6
1837	C	T	222	34	31	52.3	47.7
1939	T	G	174	32	13	71.1	28.9
1996	C	T	203	26	28	48.1	51.9
2024	T	G	222	29	37	43.9	56.1
2047	A	G	222	36	36	50.0	50.0
2062	G	A	222	31	34	47.7	52.3
2074	T	C	222	35	37	48.6	51.4
2092	C	T	222	32	41	43.8	56.2
2125	T	C	222	46	44	51.1	48.9
2161	G	A	222	51	41	55.4	44.6
2170	G	A	222	49	46	51.6	48.4
2239	T	C	222	36	31	53.7	46.3
2248	A	G	222	33	29	53.2	46.8
2251	C	T	222	33	30	52.4	47.6
2266	A	G	222	31	32	49.2	50.8
2449	C	T	123	49	19	72.1	27.9

^aSequence of the assembled contig; ^bAlternative sequence observed in RNA-seq reads; ^cPhred-scaled quality score for the assertion made in Alt by BCFtools.

Supplementary Fig. S1

Multiple alignment of the amino acid sequences of the RdRp regions of ObRV1 and related viruses

ObrV1:ASL68496.1:1274-1651	PKHSNGDSLTFFAAVKRLKFSPPQIEREKFEKVRHLGSEMFELEKLIPLDNKNDLMMQICVNEYIERKVKSPAGTIK	80
HarVA:YP_004376201.1:1274-1651	PKHSNQDSLTFFAAVKRLKFSPPHVEREKFEKVRHLGSEMFDVFLSKINLNSYNHLMQMSVNEYIEKVKSTNTIK	80
YP_006905850.1:1274-1651	PKHQNSDSLTFLAAVKRLKFSPPSVERERFEKVRHLGNEMLDIFLDKIKIDNKLNSEMMARSYNEYLVKVKSTANTIA	80
DiVB:YP_006905848.1:1274-1651	PKHSNSDTLTFFAAVKRLKFSPPQIEREKFEKVRHLGQEMLELFLEKVKVSNHYDKQEMLSYNEYIEKVKVKTGNTIA	80
CVA:ANE06570.1:1246-1621	PRHFANDDLTFWSAVKRLVFNKPLNNVHDFEKAKPFGKEMLDIFLKKVPLIPSPDQRMYYEISEFEEKIKSNAAMIG	80
CVA:APG53775.1:1246-1621	PRHFANDDLTFWSAVKRLVFNKPLNSAHDFEKAKPFGKELNIFLRKVPKPNFDQRMYSVSEFEKIKSNAAMIG	80
PCMV:YP_001497153.1:1748-2123	PRHKGDTSVTFMAVKKRSLFSPQIESAKLRRAMPFGKFLQTFVLYIPLCGKHDSLDMSKAVDFEKKLKSAAITIE	80
ApLV:ADT91605.1:1791-2166	PRHKGDTATFMAVKKRSLFSSPAAEHAKLRRARPFGKFLDFTFLKRIPLNCRHDCMMSAVHAFEEKLSKSMATIE	80
NeLV:YP_009174681.1:1486-1861	PRHKGDDTATFMAVKKRSLFSPQAEVAFRAAEPFGEYMLGVFLKHKLKLNKHEPQKMAADAKREFEKKTSAAVIE	80
NSV:YP_842438.1:1484-1859	PRHKGDDTATFMAVKKRSLFSPQAEVAFRAAEPFGEYMLGIFLKHKLKLNKHEPLKMAAAKREFEKKTSAAVIE	80
CTLaV:AHJ80270.1:1639-2014	PRHKGDTATFVMAARKLRFSPANERQKFLSAIPYGDMLRVFLEKVKLKNPFDHRLFEARNDFEKKLQKSMATLE	80
CTLaV:AHJ80314.1:1641-2016	PRHRAGDTATFVMAARKLRFSLPANEQQKFLSAIPYGDMLRVFLEKVKLKNPFDYRLFEEARNDFEKKLQKSMATLE	80
CNRMV:ALP45953.1:1640-2015	PRHKSQDTATFMAARKLRFSPANERQKFLSAIPYGDMLRVFLEKVKLKNPFDHLLDFEARNDFFEKKLQKSMATLE	80
CRMaV:AHA59466.2:1631-2006	PRHKGDTATFVMAARKLRFSPANERQKFLSAIPYGDMLRVFLEKVKLKNPFDHLLFEESRDFEKKLQKSMATLE	80
CRMaV:AHJ80324.1:1628-2003	PRHKGDTATFVMAARKLRFSPANERQKFLSAIPYGDMLRVFLEKVKLKNPFDHLLFEESRDFEKKLQKSMATLE	80
CRMaV:AKN20442.1:1631-2006	PRHKGDTATFVMAARKLRFSPAREKQKFLSAIPYGESMLQVFLKRVKLNPNFDYRLFEEARNDFEKKLQKSMATLE	80
CGRMV:AFU54620.1:1628-2003	PRHKGDTATFVMAARKLRFSPAREKQKFLSAIPYGESMLQVFLKRVKLNPNFDHLLFEAEVSDFEKKLQKSMATLE	80
BanMMV:NP_112029.1:1372-1747	PRHRNSDVTTFMAVKKRSLFSPQRKEMQRYLMMNRKKEEMADAFEKVPKSNFSREKFIKMEFESKLEKSKATIE	80
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ObrV1:ASL68496.1:1274-1651	SHSGRSDCDWKLNDVFLFKTQLCTKYEKRFSDAKAGQTLACFHVILNRFAPARYIEKIKLSLCLPDNYYIHQKKNFDM	160
HarVA:YP_004376201.1:1274-1651	SHSSRSEPDWKLNDVFLFMKTLCTKYEKRFSDAKAGQTLACFHVILNRFAPARYIEKIKLSQCLPSNYYIHQKKNFDC	160
YP_006905850.1:1274-1651	SHSSRSEPDWKLNEIFLFMKTQLCTKYEKRFSDAKAGQTLACFHVILNRFAPARYIEKIKLSQCLPSNYYIHQKKNFDC	160
DiVB:YP_006905848.1:1274-1651	AHSNRSEPDWNLNEIFLFMKTQLCTKYEKRFCEAKAGQTLACFHVILNRFAPAGTRYIEKIKSSYLPNNYYIHQKKNFDC	160
CVA:ANE06570.1:1246-1621	AHHRSTTDWPINEIFLFIKSQLCTKKEKMFCDKAGQTLACFHVILNRFAPLNRVIEKIKVQCLPGNFIYHQQKNFDE	160
CVA:APG53775.1:1246-1621	AHHRSTTDWPTNEIFLFIKSQLCTKKEKMFCDKAGQTLACFHVILNRFAPLNRVIEKIKVQCLPGNFIYHQQKNFDE	160
PCMV:YP_001497153.1:1748-2123	NHSGRSSRDWVVKALIFMKSQCLCTKFDNFRFSAKAGQTLACFQHSVLCRFAPYMYRIEAKLFNALPDRFYIHSKNIIDD	160
ApLV:ADT91605.1:1791-2166	NHSGRSCDWVVKALIFMKSQCLCTKFDNFRFSAKAGQTLACFQHSVLCRFAPYMYRIEAKLVVLEALPNLYIHSKNIIEE	160
NeLV:YP_009174681.1:1486-1861	NHSGRSCRDWLDLGLFIFKNQCTKFEKRFADAKAAQTLICFQHEVLCRFAPYVRYIEKLNDAIPAKYIHSKNIIDD	160
NSV:YP_842438.1:1484-1859	NHSGRSCRDWLDLGLFIFKNQCTKFEKRFADAKAAQTLICFQHEVLCRFAPYVRYIEKLNDAIPAKYIHSKNIIDD	160
CTLaV:AHJ80270.1:1639-2014	NHSGRSDPDWVEKALIFMKSQCLCTKFDNFRFSAKAGQTLACFHHNVLCRLAPYIRYIEKIKVFAALPKNLYIHSKNIIDD	160
CTLaV:AHJ80314.1:1641-2016	NHSGRSDPDWVEKALIFMKSQCLCTKFDNFRFSAKAGQTLACFHHNVLCRLAPYIRYIEKIKVFNLPKNLYIHSKNIIDD	160
CNRMV:ALP45953.1:1640-2015	NHSGRSDPDWVEKALIFMKSQCLCTKFDNFRFSAKAGQTLACFHHNVLCRLAPYVRYIEKIKVFNLPKNLYIHSKNIIDD	160
CRMaV:AHA59466.2:1631-2006	NHSGRSDPDWVEKALIFMKSQCLCTKFDNFRFSAKAGQTLACFHHNVLCRLAPYIRYIEKIKVFNLPKNLYIHSKNIIDD	160
CRMaV:AHJ80324.1:1628-2003	NHSGRSDPDWVEKALIFMKSQCLCTKFDNFRFSAKAGQTLACFHHNVLCRLAPYIRYIEKIKVFNLPKNLYIHSKNIIDD	160
CRMaV:AKN20442.1:1631-2006	NHSGRSDPDWVENASIFMKSQCLCTKFDNFRFSAKAGQTLACFHHNVLCRLAPYIRYIEKIKVFNLPKNLYIHSKNIIDD	160
CGRMV:AFU54620.1:1628-2003	NHSGRSDPDWVEKALIFMKSQCLCTKFDNFRFSAKAGQTLACFHHNVLCRLAPYIRYIEKIKVFNLPKNLYIHSKNIIDD	160
BanMMV:NP_112029.1:1372-1747	NHAQRSNREWKIDEAMIFMKSQCLCTKFEKRFADAKAGQTLACFHVILNRFAPYVRYIEKIKVFNLPKNLYIHSKNIIDD	160
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ObrV1:ASL68496.1:1274-1651	LNAWVVRNDFSDCELESYAEAFDSSQDCLILAFEYELLKYMGSQSLDDYLDLDFNLGCRNLGNLAVMRFTGEFTFLFN	240
HarVA:YP_004376201.1:1274-1651	LNEWVIRNDFSESCLESYAEAFDSSQDCLILAFEYELLKYMGSQSLDDYLDLDFNLGCRNLGNLAVMRFTGEFTFLFN	240
YP_006905850.1:1274-1651	LNDWVVANNFDSYCESYAEAFDSSQDCLILAFEYELLKYLWGDQSLDDYLDLDFNLGCRNLGNLAVMRFTGEFTFLFN	240
DiVB:YP_006905848.1:1274-1651	LNEWVIENKFTGESLESYAEAFDSSQDCLILAFEYELLKYLWGNQSLDDYLDLDFNLGCRNLGNLAVMRFTGEFTFLFN	240
CVA:ANE06570.1:1246-1621	LEKWKVSYDFSGVCTESDYAEADASQDSYTLAFEYELLMKYLGVSNMSIEDYLYLKMHLNCKLGNLAINRFTGEFTFLFN	240
CVA:APG53775.1:1246-1621	LEKWKVSYDFSGVCTESDYAEADASQDSYTLAFEYELLMKYLGVSNMSIEDYLYLKMHLNCKLGNLAINRFTGEFTFLFN	240
PCMV:YP_001497153.1:1748-2123	LGNWVKKQNFSGECTESDYAEAFDASQDHFILAFEIEIMRHLLGPEGLINDYIFIKCNLGSKLGSAIMRFTGEASTFLFN	240
ApLV:ADT91605.1:1791-2166	LADWVKNKFNVCCTESDYAEAFDASQDHFILAFELVEMKHLGLPADLADYTFIKTHLGSKLGSAIMRFTGEASTFLFN	240
NeLV:YP_009174681.1:1486-1861	LNDWVKDNDVSGICTESDYAEAFDASQDHFILAFEIVAMKYLGLPRDLINDYIFIKTHLGSKLGSAIMRFTGEASTFLFN	240
NSV:YP_842438.1:1484-1859	LNDWVKANNFSGVCTESDYAEAFDASQDHYIMAFEVAMKYLGLPRDLVNDYIFIKTHLGSKLGSAIMRFTGEASTFLFN	240
CTLaV:AHJ80270.1:1639-2014	LRDWVIAHRFSGVCTESDYAEAFDSSQDANILAFEVSLMNYLNLPRDLIEDYKHLKFHTHSLKLGQFVAVMRFTGEAGTFLFN	240
CTLaV:AHJ80314.1:1641-2016	LRDWIKSNFSGMCTESDYAEAFDSSQDANILAFEVSLMNYLNLPRDLIEDYKHLKFHTHSLKLGQFVAVMRFTGEAGTFLFN	240
CNRMV:ALP45953.1:1640-2015	LQEWVIQNNFVGTCTESDYAEAFDSSQDANILAFEVSLMNYLNLPRDLIEDYKHLKFHTHSLKLGQFVAVMRFTGEAGTFLFN	240
CRMaV:AHA59466.2:1631-2006	LRDWVIENNFTGCTESDYAEAFDSSQDANILAFEVSLMEYLRLPRDLIEDYKHLKFHTHSLKLGQFVAVMRFTGEAGTFLFN	240
CRMaV:AHJ80324.1:1628-2003	LRDWVIRNNTGCTESDYAEAFDSSQDANILAFEVSLMEYLRLPRDLIEDYKHLKFHTHSLKLGQFVAVMRFTGEAGTFLFN	240
CRMaV:AKN20442.1:1631-2006	LRDWVIANNFTGLCTESDYAEAFDSSQDANILAFEVSLMNYLNLPRDLIEDYKHLKFHTHSLKLGQFVAVMRFTGEAGTFLFN	240
CGRMV:AFU54620.1:1628-2003	LRDWVKNFSGMCTESDYAEAFDSSQDANILAFEVSLMNYLNLPRDLIEDYKHLKFHTHSLKLGQFVAVMRFTGEAGTFLFN	240
BanMMV:NP_112029.1:1372-1747	LNDYVKNNSFGRICLESYAEAFDSSQDANILAFEVSLMNYLNLPRDLIEDYKHLKFHTHSLKLGQFVAVMRFTGEAGTFLFN	240
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ObrV1:ASL68496.1:1274-1651	TLANMVFTFMSYDLTGKEAICFAGDDMCCKNGIRRRDGRFDHILNRLSKAKAVITTEPTFCGWRLTKYGIFFKPELVL	320
HarVA:YP_004376201.1:1274-1651	TLANMVFTFMSYELNGKESICFAGDDMCCKNGIKRIRIDGKFDHILKRLSKAKAVITTEPTFCGWRLTPFGYIKPELVL	320
YP_006905850.1:1274-1651	TLANMVFTFMTYDNLGTEISCFAGDDMCCKNGIKARVDGKYDHLKRLTLKAKAVITTEPTFCGWRLTKYGIFFKPELVL	320
DiVB:YP_006905848.1:1274-1651	TLANMFTFMSYDLDDKEAICFAGDDMCCKNPVKKRSQGRYDHLKRLTLKAKAVSYTREPTFCGWRLTKHGIFFKPELVL	320
CVA:ANE06570.1:1246-1621	TLTNMLFTFMKYDVRKTHAICFAGDDMCANVRLPENLQ--HTNLKKFSLKAKAVDFTRSPFCGWNLTRYIVKPELIA	318
CVA:APG53775.1:1246-1621	TLTNMLFTFMKYDVRKTHAICFAGDDMCANVRLPENHQ--YSDLKRSLLKAKAVDFTRSPFCGWNLTRYIVKPELIA	318
PCMV:YP_001497153.1:1748-2123	TMANMFTFLRYELSGHESIFAGDDMCANRRLRVRST--YKSFLEKIRLAKAVQFTNPTFCGWGLCPGVFKKPDLLV	318
ApLV:ADT91605.1:1791-2166	TMANMFTFLRYDLNGKEAICFAGDDMCANRRLKVTQT--HSKFLDKIKLAKAVQFTNPTFCGWGLCEHGVFKKPDLLV	318
NeLV:YP_009174681.1:1486-1861	TMSNMLFTFMKYEINGESICFAGDDMCASKLRLSDE--HSSYLEKRLKAKAVCFTRPTFCGWNLTRYIVKPELVL	318
NSV:YP_842438.1:1484-1859	TMSNMLFTFMKYEITGKESICFAGDDMCASKLRLSTV--HSSYLEKRLKAKAVCFTRPTFCGWNLTRYIVKPELVL	318
CTLaV:AHJ80270.1:1639-2014	TLANMVFTFMRYETNGREIACFAGDDMCANLRLRKSSE--FEHVLDRMTLAKAVQHTTEPTFCGWRLNGFIVKRPQLVQ	318
CTLaV:AHJ80314.1:1641-2016	TLANMVFTFMRYETNGREIACFAGDDMCANLRLRKSSE--FEHVLDRMTLAKAVQHTTEPTFCGWRLNGFIVKRPQLVQ	318
CNRMV:ALP45953.1:1640-2015	TLANMVFTFMRYETNGREIACFAGDDMCANLRLRKSSE--FEHVLDRMTLAKAVQHTTEPTFCGWRLNGFIVKRPQLVQ	318
CRMaV:AHA59466.2:1631-2006	TLANMVFTFMRYEYNGREIACFAGDDMCANLRLRKSSE--FEHILDRMTLAKAVQHTTEPTFCGWRLNGFIVKRPQLVQ	318
CRMaV:AHJ80324.1:1628-2003	TLANMVFTFMRYEYNGREIACFAGDDMCANLRLRKSSE--FEHILDRMTLAKAVQHTTEPTFCGWRLNGFIVKRPQLVQ	318
CRMaV:AKN20442.1:1631-2006	TLANMVFTFMRYEYNGREIACFAGDDMCANLRLRKSSE--FEHILDRMTLAKAVQHTTEPTFCGWRLNGFIVKRPQLVQ	318
CGRMV:AFU54620.1:1628-2003	TLANMVFTFMRYDNLGKEAICFAGDDMCANLRLRKSSE--FEHILDRMTLAKAVQHTTEPTFCGWRLNGFIVKRPQLVQ	318
BanMMV:NP_112029.1:1372-1747	TMANIVFTIMAYDLKGDCEILFAGDDMCMTVRRANNS--YTHILKLNKLLKAVGITEEPTFCGWRLTIHGIVKRPQLVQ	318
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ObRV1:ASL68496.1:1274-1651	ERFLIAIEKNNLKDVIDSYYLECSYAYSLGERLFECEKDFVAHYCCIRLVHLHKDL	378
HarVA:YP_004376201.1:1274-1651	ERFLIAIEKGRLLDVIDSYYLECSYAYSLGERLSKCFSEKDFVAHYCCVRLVHKHKSL	378
YP_006905850.1:1274-1651	ERFLIAIEKGRLLDVIDSYYIECSYAYNLGERLFECEKDFSAHYCCIRIVHKHKSL	378
DiVB:YP_006905848.1:1274-1651	ERFLIAIEKGRLLKEVIDSYFIECSYAYNLGERLYECEFSEKDFIAHYCCVRIVHENS	378
CVA:ANE06570.1:1246-1621	ARLAVAKQKGEVNLVDSYFLEHLYAYNKGDLFEILSEKELEHHYNLTRFFVKHGNL	376
CVA:APG53775.1:1246-1621	ARLAVAKQKGEINLVDSYFLEHLYAYNKGDLFEILSEKELEHHYNLTRFFVKHNSL	376
PCMV:YP_001497153.1:1748-2123	ERLQIAVETNNLQNCIDNYAIEVSYAYSMGESLSKYLSEEMDAHYNCVRFIVKHSHL	376
ApLV:ADT91605.1:1791-2166	ERLQIARETRNLENCIDNYAIEVSCAYRMGENLNLYLSPNEMDAHYNCVRFIILHNHL	376
NeLV:YP_009174681.1:1486-1861	ERMCAIEKNNLQNCIDNYAIEVSYAYLMGENALVLMDEEIEHNNHYMCVRTIVQNKNL	376
NSV:YP_842438.1:1484-1859	GRMCAIEKNNLQNCIDNYAIEVSYAYLMGENALVLMDEEIEHNNHYMCVRTIVQNKNL	376
CTLaV:AHJ80270.1:1639-2014	ERILIAIEKGNFNECIDNYAIEVSYAYNLGERLVSIMSEKELDAHFCVTRTFLQHKNL	376
CTLaV:AHJ80314.1:1641-2016	ERILIAIEKGNFNECIDNYAIEVSYAYNLGERLVSIMSEKELDAHFCVTRTFLQHKNL	376
CNRMV:ALP45953.1:1640-2015	ERILIAIEKGNFNECIDNYAIEVSYAYNLGERLVSIMSEKELDAHFCVTRTFLQNKKL	376
CRMaV:AHA59466.2:1631-2006	ERILIAIEKGNFNECIDNYAIEVSYAYNLGERLVSIMSEKELDAHFCVTRTFLQNKKL	376
CRMaV:AHJ80324.1:1628-2003	ERILIAIEKGNFNECIDNYAIEVSYAYNLGERLVSIMSEKELDAHFCVTRTFLQNKKL	376
CRMaV:AKN20442.1:1631-2006	ERILIAIEKGNFNECIDNYAIEVSYAYNLGERLVSIMSEKELDAHFCVTRFLRNKNL	376
CGRMV:AFU54620.1:1628-2003	ERILIAIEKGNFNECIDNYAIEVSYAYNLGERLVSIMSEKELDAHFCVTRTFLQNKCL	376
BanMMV:NP_112029.1:1372-1747	ERFMIAIENGLENLENCIDNYAIECSYAYKLGDRLVSMFSEEENCAHYILVRYIVKKRHL	376
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Asterisks (*), fully conserved residues; colons (:), residues with strongly similar properties; and periods (.), residues with weakly similar properties. See Table 2 for full names of viruses.

Supplementary Fig. S2

Multiple alignment of the amino acid residues of the RdRp regions of ObrV2 and related viruses

Table with multiple columns showing amino acid alignments for various viruses including ObrV2, BcMV1, FpMV4, OMV3a, SsMV3, MpmV3, RsmV, RsmV-15, RsmV-14, HetMV1, COV, BcMV3, FpMV3, S1aMV5, S1aMV5-19, SsMV16, SsMV16-19, CpMV1, RcmV, SsMV15, HfmV1, SsMV16, SsMV2, S1aMV3, S1aMV3-19, SsMV16, SsMV2, FgMV1, and TbmV. The table shows conserved residues with asterisks and similar residues with colons.

Asterisks (*), fully conserved residues; colons (:), residues with strongly similar properties; and periods (.), residues with weakly similar properties. See Table 2 for virus names.