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Prognostic value of bone marrow MUC4 expression in acute myeloid leukaemia

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ABSTRACT

Background: Aberrant expression of mucin-4 (MUC4) is present in a variety of solid cancers, but the expression pattern of *MUC4* and its clinical relevance in acute myeloid leukaemia (AML) is unknown. We aimed to evaluate the expression level of *MUC4* and explore its prognostic value in newly diagnosed adult patients with AML.

Methods: Bone marrow from 70 AML patients and 26 healthy donors was obtained. *MUC4* levels were quantified by quantitative real-time PCR. Routine blood indices were measured by standard techniques.

Results: Bone marrow *MUC4* expression levels were significantly elevated in AML patients compared to controls at median (range) 2.77 (0.7–16.6) and 1.14 (0.5–1.99) respectively (p = 0.005). Moreover, lower *MUC4* expression was strongly associated with persistent remission (p = 0.001) while higher *MUC4* levels were associated with worse overall as well as disease-free survival (p = 0.011 and p = 0.006, respectively). Thus, its level may act as an indicator of disease progression. High *MUC4* expression was identified as an independent prognostic predictor for both overall survival and disease-free survival.

Conclusion: *MUC4* over-expression is an independent predictor of a poor prognosis in AML patients.

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KEYWORDS

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Introduction

Acute myeloid leukaemia (AML) is a common acute leukaemia that affects adults, it is an aggressive haematological malignancy caused by several factors including environmental factors, chromosomal aberrations, and gene mutations. AML is a cytogenetically, and molecularly heterogeneous disease characterized by overproliferation and accumulation of myeloid blasts in the bone marrow and blood [1,2]. Though the survival rate in AML has improved, relapse remains a major obstacle towards treatment [3] and the prognosis assessment of AML is still difficult. Therefore, identifying an effective and novel marker for the diagnosis and prognosis of clinical outcome and treatment response is vital.

Mucins are a heterogeneous family of high molecular weight glycoproteins that have been subdivided into three types: secretory (gel-forming), membranebound, and soluble mucins [4]. They are produced from various types of epithelial cells and leukocytes and play an important role in lubrication and protection from microbial pathogens [5,6]. Additionally, mucins participate in several signalling pathways to regulate cellular renewal, cell–cell interactions, differentiation, and apoptosis [7,8].

MUC4 is located at chromosome locus 3q29 and encodes the multi-domains transmembrane protein mucin-4 (MUC4) [9]. With its epidermal growth factor (EGF)-like domains, MUC4 binds to erythroblastic oncogene B2 (ErbB2) receptor tyrosine kinase, also known as epidermal growth factor receptor (HER2), to facilitate signal transduction, cell proliferation, and cell survival [10–12].

Although several studies on solid cancers have implicated MUC4 expression in diagnosis and disease progression, data about its levels and prognostic value are largely inconclusive. MUC4 is over-expressed in breast and pancreatic cancers, while its expression level is decreased in prostatic adenocarcinoma and bladder cancer [13–16]. Although the prognostic significance of MUC4 varies with the cancer type, MUC4 expression is associated with tumour aggressiveness and poor survival in lung and ovarian cancers [17,18], whilst in mucoepidermoid carcinoma of salivary glands, MUC4 expression is related to better survival [19].

Although *MUC4* is significantly mutated in wholeexome analysis of normal karyotypes, indicating that *MUC4* might be a predictor for the risk of normal karyotype AML [20], the prognostic utility of *MUC4* in AML is unknown. We, therefore, hypothesised the alteration of *MUC4* expression level in the bone marrow of adult AML patients and its potential utility as a prognostic marker.

Materials and methods

We recruited 70 newly diagnosed adult AML patients from the medical oncology department, National Cancer Institute, Cairo University, Cairo, Egypt, who

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were treated and followed up between May 2016 and October 2019, while acute promyelocytic leukaemia (M3) subtype patients received different treatment protocols; thus, they were excluded from this study. All AML cases were diagnosed according to criteria of the French-American-British (FAB) classification and WHO combined with immunological and cytogenetic analyses [21]. Twenty-six age and sex-matched healthy donors (donors of a bone marrow transplant) with no clinical symptoms of haematologic or other types of cancer and other diseases were enrolled. Bone marrow (BM) aspiration specimens were collected into EDTA from all participants. This study was approved by the Ethics Committee and Institutional Review Board of National Cancer Institute, Cairo University (201617027--4). All participants signed written informed consent following the Declaration of Helsinki guidelines.

AML patients received induction chemotherapy consisting of the standard 3 + 7 regimen (Doxorubicin 30 mg/m² on 1–3 days; Cytarabine 100 mg/m² on 1–7 days). By the end of first induction therapy, complete remission (CR) was defined by normalization of bone marrow (BM) elements; neutrophil count of 1×10^9 /L and platelet count of 100×10^9 /L, and BM examination showing a normocellular marrow containing less than 5% blasts. Resistant disease was defined as the presence of more than 25% blasts in BM. Overall survival (OS) was the time from the entry into the study to death and disease-free survival (DFS) was the time from the date of CR achievement to death or relapse.

Total RNA was extracted from bone marrow cells using QIAamp[®] RNA Blood Mini Kits (Cat# 52304, Qiagen, Germany) based on the manufacturer's instructions. The purity and the concentration of the purified RNA were detected using Nano-Drop (Quawell, Q-500, Scribner, USA). The synthesis of cDNA was performed by reverse transcription using High-Capacity cDNA Reverse Transcription Kit (Cat# 4368814, Thermo Fisher) and stored at – 20°C till performing quantitative real-time PCR.

Real-time quantitative PCR was carried out to evaluate the expression of MUC4 and GAPDH (as internal control) using Step OneTM Real-Time PCR System (Applied Biosystems). The real-time PCR reactions were performed in a 20 μ L volume using TaqManTM Universal Master Mix II (Cat# 4440043, Applied Biosystems) and TaqManTM Gene Expression Assays for MUC4 and GAPDH were (Assay ID: Hs00366414_m1; Cat# 4331182, and Assay ID: Hs03929097_g1; Cat# 4331182, Applied Biosystems, Foster City, USA). RQ-PCR reactions were performed as follows: one cycle at 95°C for 10 min, followed by 40 cycles at 95°C for 15 s and at 60°C for 1 min. Relative MUC4 expression level was calculated using 2^{- $\Delta\Delta$ Ct} method [22].

IBM-SPSS version 20.0 software was used to analyse the data (IBM Corp, NY, US). Categorical data were expressed as frequencies and percentages. Shapiro– Wilk test was performed to determine the distribution for numerical data; normally distributed data were expressed as mean±SD while non-normally distributed data were described using median and interquartile range (25th and 75th percentile). For continuous variables, independent Student's t-test and Mann– Whitney's U test were used to compare the difference between the two groups. Chi-square analysis or Fisher exact test were carried out for categorical variables. The prognostic value of *MUC4* for OS and DFS of AML patients was estimated *via* Kaplan–Meier survival curves using log-rank test and Cox regression analysis. p-Value < 0.05 was considered significant.

Results

Baseline characteristics of the 70 AML patients are summarized in Table 1. Median (range) of the patient and controls were 33 (26-45 years) and 35 (29-50), respectively (p = 0.191). The number of men and women was 46/ 34 patients and 15/11 controls (p = 0.468). The majority of the patients were classified as acute myeloblastic leukaemia with maturation/acute myelomonocytic leukaemia (M2/M4) subtypes. MUC4 expression level was significantly increased in AML bone marrow samples compared to healthy donors at median (range) 2.77 (0.7-16.6) and 1.14 (0.5-1.99) respectively (p = 0.005). Of the 70 patients, 45 (64%) achieved CR after the first induction, but 15 (33% of CR; 21% of total) relapsed later. Resistant and relapsed patients were collectively labelled as 'poor responders' (47.4%), while patients with persistent remission (52.6%) were labelled as 'good responders'.

Patients were assigned to two groups based on the median expression level of *MUC4*. Table 2 shows that *MUC4* expression level is significantly lower in patients with persistent remission, being alive or good responders. The level of *MUC4* expression is strongly associated with FAB subtypes, treatment response, survival status and final outcome. There were no links between *MUC4* expression and other clinical variables, including gender, age, white blood cells count, haemoglobin, platelet count, BM blasts, karyotypic classifications and FLT3 mutation.

During 41 months of follow-up, the mean (95% Cl) OS and DFS intervals were 18.9 months (14.3–23.3) and 21.0 (16.8–25.1), respectively. Kaplan-Meier analysis highlighted that high *MUC4* expressers suffered from significantly worse OS (p = 0.011, Figure 1(a)) and shorter DFS (p = 0.006, Figure 1(b)) than low expressers.

In Cox proportional hazard model, univariate analysis showed that high bone marrow *MUC4* expression level was an independent prognostic indicator for predicting poorer OS and DFS (Table 3). In adjusting for age and sex, hazard ratios for OS and DFS were 2.9 (1.5–5.9) (p = 0.002), and 4.3 (1.4–13.9) (p = 0.014) respectively.

Table 1. Baseline characteristics of AML patients.

Variables		N (%)
Gender	Male	46 (65.7)
	Female	24 (34.3)
Age, median (range), years		33 (26–45)
FAB classification	M0	1 (1.4)
	M1	5 (7.1)
	M2	33 (47.1)
	M4	23 (32.9)
	M5	7 (10)
	M6	1 (1.4)
Organomegaly	Hepatomegaly (present vs absent)	12/58
	Splenomegaly (present vs absent)	11/59
	Lymphadenopathy (present vs absent)	21/49
FLT3 mutation	Wild	59 (84.3)
	Mutant	11 (15.7)
Karyotype	Favourable	24 (34.3)
classifications	Intermediate	33 (47.1)
	Adverse	13 (18.6)
Treatment response	Induction death	13 (18.6)
	Resistant	12 (17.1)
	Relapse	15 (21.4)
	Persistent remission	30 (42.9)
Survival status	Dead	39 (55.7)
	Alive	31 (44.3)
Final outcome	Poor (Resistant+ Relapse)	27 (47.4)
	Good (Persistent remission)	30 (52.6)

Categorical data are expressed as number (percentage). AML, acute myeloid leukaemia; FAB, French-American-British; M0, acute myeloblastic leukaemia with minimal differentiation; M1, acute myeloblastic leukaemia without maturation; M2, acute myeloblastic leukaemia with maturation; M4, acute myelomonocytic leukaemia; M5, acute monocytic leukaemia; M6, acute erythroid leukaemia

Discussion

Despite numerous studies on the oncogenic potential of *MUC4* in different malignancies, there is no detailed study on the expression of *MUC4* in AML. Thus, we explored the *MUC4* expression level in order to evaluate for the first time its clinical significance as a potential prognostic tool for adult AML. The bone marrow expression level of *MUC4* was significantly elevated in AML compared to the control group. Additionally, the high *MUC4* expression level was associated with worse survival and shorter DFS. Finally, *MUC4* expression level was found to be an independent predictor for AML.

Several researches have supported a role for MUC4 in various cancers, where it is involved in cellular functions as tumour growth, proliferation, adhesion, invasion, inhibition of apoptosis, and chemo-resistance [23]. However, there is contradictory data about its expression level; Sadras et al. reported that *MUC4* expression was up-regulated in Ph-like CRLF2-rearranged acute lymphoblastic leukaemia compared to non- Ph-like CRLF2-rearranged acute lymphoblastic leukaemia group [24]. In addition, MUC4 over-expression was found in lung adenocarcinoma [17], ovarian cancer [18], breast cancer [13], pancreatic cancer [14], and mucoepidermoid carcinomas of the salivary gland [19]. This is supported by the results of the current work. In contrast, a reduced *MUC4* expression was exhibited in

Table 2.	Link	between	clinical	features	of	AML	patients	with
aberrant	expre	essed MU	C4.					

	MUC4 ex		
Variables	Low (n = 35)	P-value	
Sex, male/female	21/14	25/10	0.314
Age (years)	34.3 ± 9.4	36.5 ± 11.3	0.386
WBC, ×10 ⁹ /L	27.0 (4–98)	14.8 (5–62)	0.707
Hb, g/L	77 ± 17	80 ± 21	0.466
PLT, ×10 ⁹ /L	29 (13–62)	31 (18–57)	0.617
BM blasts, %	64.8 ± 22.5	56.1 ± 22.1	0.109
FAB classifications			0.021
M0	0 (0%)	1 (2.9%)	
M1	4 (11.4%)	1 (2.9%)	
M2	12 (34.4%)	21 (60%)	
M4	12 (34.3%)	11 (31.4%)	
M5	7 (20%)	0(0%)	
M6	0 (0%)	1 (2.9%)	
FLT3 mutation			0.324
Wild	28 (80%)	31 (88.6%)	
Mutant	7 (20%)	4 (11.4%)	
Karyotype classification			0.216
Favourable	9 (25.7%)	15 (42.9%)	
Intermediate	20 (57.1%)	13 (37.1%)	
Adverse	6 (17.1%)	7 (20%)	
Treatment response			0.001
Induction death	6 (17.1%)	7 (20%)	
Resistant	2 (5.7%)	10 (28.6%)	
Relapse	4 (11.4%)	11 (31.4%)	
Persistent remission	23 (65.7%)	7 (20%)	
Survival status			0.002
Dead	13 (37.1%)	26 (74.3%)	
Alive	22 (62.9%)	9 (25.7%)	
Final outcome			<0.001
Poor	6 (20.7%)	21 (75%)	
Good	23 (79.3%)	7 (25%)	

Data are expressed as mean±SD, median (interquartile range) and frequency (percentage) categorical data. AML, acute myeloid leukaemia; WBC, White blood cells; HB, haemoglobin; PLT, platelet, BM, bone marrow; FAB, French-American-British; M0, acute myeloblastic leukaemia with minimal differentiation; M1, acute myeloblastic leukaemia without maturation; M2, acute myeloblastic leukaemia with maturation; M4, acute myelomonocytic leukaemia; M5, acute monocytic leukaemia; M6, acute erythroid leukaemia

prostatic adenocarcinoma tissues compared to the adjacent benign tissue [15]. Additionally, the expression of MUC4 is lower in urothelial carcinoma [16].

This aberrant over-expression may be attributed to different mechanisms. Firstly, an association between alterations of *MUC4* copy number and *MUC4* expression was found [25]; therefore, amplification of the *MUC4* locus may contribute to this over-expression.

Secondly, regulators such as growth factors, cytokines, and miRNA may affect the alteration of *MUC4* expression [26–29]. Mejías-Luque et al. demonstrated that IL-6 is responsible for *MUC4* up-regulation in gastric cancer cell lines through the STAT pathway [30]. In addition, Stevens et al. showed that IL-6 activates STAT3 signalling in paediatric AML [31]. Further, bone marrow and plasma IL-6 levels are elevated in AML and this elevation correlates with shorter survival [32,33]. Collectively, we suggest that *MUC4* over-expression might be mediated *via* IL-6 induced STAT3 activity. This is speculative and further molecular mechanisms should be involved.

Although MUC4 is considered as a prognostic marker in various cancers, its exact potential in AML is still blurred. We found that *MUC4* over-expression

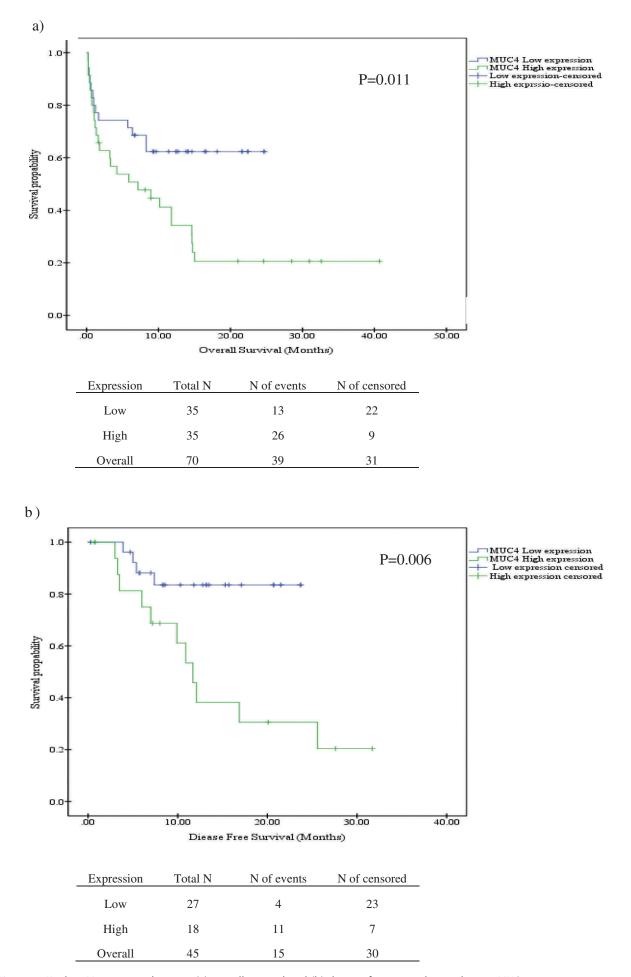


Figure 1. Kaplan–Meier survival curves: (a) overall survival and (b) disease-free survival according to MUC4 expression.

 Table 3. Univariate analysis for overall survival and disease-free survival.

	Univariate analysis		
Variables	HR (95% CI)	P-value	
Overall survival			
MUC4	2.3 (1.2-4.5)	0.014	
Sex	1.2 (0.6–2.2)	0.650	
Age (years)	0.9 (0.5–1.7)	0.730	
FAB classifications	1 (0.8–1.3)	0.950	
FLT3 mutation	0.6 (0.2–1.6)	0.311	
Karyotype classification	1.0 (0.7–1.6)	0.834	
Disease-free survival			
MUC4	4.4 (1.4–14.2)	0.012	
Sex	1.1 (0.4–3.1)	0.881	
Age (years)	1.5 (0.5-4.1)	0.485	
FAB classifications	0.7 (0.5-1.1)	0.131	
FLT3 mutation	0.7 (0.2-2.6)	0.638	
Karyotype classification	1.2 (0.6–2.4)	0.589	

AML, acute myeloid leukaemia; FAB, French-American-British; HR, hazard ratio; 95% CI: 95% confidence interval.

predicted a poorer OS and shorter DFS of AML patients. Moreover, multivariate Cox analysis highlighted the clinical significance of MUC4 levels for the AML prognosis independently of patients' clinicopathological data. Notably, Rakha et al. highlighted that the up-regulated levels of MUC4 in breast cancer patients are associated with worse overall survival [34]. Moreover, increased MUC4 expression was associated with a decreased survival rate among patients with cholangiocarcinoma and with colorectal adenocarcinoma [35,36]. Additionally, MUC4 is over-expressed in biliary tract cancer and this increase is associated with poor survival rate [37]. Conversely, MUC4 overexpression is associated with improved patient survival and prolonged time to relapse in mucoepidermoid carcinoma of salivary glands [19].

MUC4 has been implicated in suppression of apoptosis and stimulation of proliferation in several cancers; Workman et al. demonstrated that MUC4 overexpression suppressed apoptosis through augmentation of ErbB2/HER2 signalling to regulate antiapoptotic Bcl-2 proteins in breast cancer cells [38]. Further, Skrypek et al. revealed that gemcitabine efficacy was improved in MUC4 knockdown pancreatic cancer cell lines. The authors showed this knockdown decreased the activation of the mitogen-activated protein kinases (MAPK), c-Jun-NH2-kinase (JNK), and nuclear factor-kappa beta (NF-KB) pathways [39]. In ovarian cancer cells, MUC4 over-expression induced the phosphorylation of focal adhesion kinase (FAK), a non-receptor tyrosine kinase, and the activation of Akt and extracellular signal-regulated kinase (ERK) pathways [40]. These events can promote cell migration and proliferative responses that may contribute to the severity of the disease and in turn a worse OS.

Due to the limited number of cases in our study, further studies are needed using a larger sample size to determine the mechanism of aberrant *MUC4* overexpression and to elucidate the molecular pathway linking MUC4 to leukaemic progression and resistance to therapy. Nevertheless, our data offers convincing evidence that bone marrow *MUC4* expression is markedly elevated in AML and is closely associated with poor clinical outcome. Therefore, bone marrow *MUC4* might serve as a novel prognostic biomarker for AML and may provide a new target for therapy.

This work represents an advance in biomedical science because it links *MUC4* expression level with prognosis in AML.

Summary table

What is known about this topic:

- In solid malignancies, MUC4 expression is associated with chemoresistance.
- MUC4 expression may induce invasion and metastasis in various types of cancer.

What this work adds:

- Increased levels of MUC4 gene expression in de novo AML patients compared to healthy controls.
- MUC4 gene level acts as a novel prognostic marker in AML.

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Disclosure statement

The authors declare no conflict of interest.

References

- Prada-Arismendy J, Arroyave JC, Rothlisberger S. Molecular biomarkers in acute myeloid leukemia. Blood Rev. 2017;31:63–76.
- [2] Estey E, Döhner H. Acute myeloid leukemia. Lancet. 2006;368:1894–1907.
- [3] Breems DA, Van Putten WL, Huijgens PC, et al. Prognostic index for adult patients with acute myeloid leukemia in first relapse. J Clin Oncol. 2005;23:1969–1978.
- [4] Jonckheere N, Van Seuningen I. The membranebound mucins: from cell signaling to transcriptional regulation and expression in epithelial cancers. Biochimie. 2010;92:1–11.
- [5] Pelaseyed T, Bergström JH, Gustafsson JK, et al. The mucus and mucins of the goblet cells and enterocytes provide the first defense line of the gastrointestinal tract and interact with the immune system. Immunol Rev. 2014;260:8–20.
- [6] Kufe DW. Mucins in cancer: function, prognosis, and therapy. Nat Rev Cancer. 2009;9:874–885.
- [7] Moniaux N, Escande F, Porchet N, et al. Structural organization and classification of the human mucin genes. Front Biosci. 2001;6:D1192– D1206.
- [8] Andrianifahanana M, Moniaux N, Batra SK. Regulation of mucin expression: mechanistic aspects and implications for cancer and inflammatory diseases. Biochim Biophys Acta. 2006;1765:189–222.

- [9] Byrd JC, Bresalier RS. Mucins and mucin binding proteins in colorectal cancer. Cancer Metastasis Rev. 2004;23:77–99.
- [10] Singh PK, Hollingsworth MA. Cell surface associated mucins in signal transduction. Trends Cell Biol. 2006;16:467–476.
- [11] Jonckheere N, Skrypek N, Merlin J, et al. The mucin MUC4 and its membrane partner ErbB2 regulate biological properties of human CAPAN-2 pancreatic cancer cells via different signalling pathways. PLoS One. 2012;7:e32232.
- [12] Karg A, Dinc ZA, Basok O, et al. MUC4 expression and its relation to ErbB2 expression, apoptosis, proliferation, differentiation, and tumor stage in non-small cell lung cancer (NSCLC). Pathol Res Pract. 2006;202:577.
- [13] Mukhopadhyay P, Lakshmanan I, Ponnusamy MP, et al. MUC4 overexpression augments cell migration and metastasis through EGFR family proteins in triple negative breast cancer cells. PLoS One. 2013;8:e54455.
- [14] Saitou M, Goto M, Horinouchi M, et al. MUC4 expression is a novel prognostic factor in patients with invasive ductal carcinoma of the pancreas. J Clin Pathol. 2005;58:845–852.
- [15] Singh AP, Chauhan SC, Bafna S, et al. Aberrant expression of transmembrane mucins, MUC1 and MUC4, in human prostate carcinomas. Prostate. 2006;66:421–429.
- [16] Kaur S, Momi N, Chakraborty S, et al. Altered expression of transmembrane mucins, MUC1 and MUC4, in bladder cancer: pathological implications in diagnosis. PLoS One. 2014;9:e92742.
- [17] Tsutsumida H, Goto M, Kitajima S, et al. MUC4 expression correlates with poor prognosis in small-sized lung adenocarcinoma. Lung Cancer. 2007;55:195–203.
- [18] Chauhan SC, Singh AP, Ruiz F, et al. Aberrant expression of MUC4 in ovarian carcinoma: diagnostic significance alone and in combination with MUC1 and MUC16 (CA125). Mod Pathol. 2006;19:1386–1394.
- [19] Alos L, Lujan B, Castillo M, et al. Expression of membrane-bound mucins (MUC1 and MUC4) and secreted mucins (MUC2, MUC5AC, MUC5B, MUC6 and MUC7) in mucoepidermoid carcinomas of salivary glands. Am J Surg Pathol. 2005;29:806–813.
- [20] Heo SG, Hong EP, Park JW. Genetic risk prediction for normal-karyotype acute myeloid leukemia using whole-exome sequencing. Genomics Inform. 2013;11:46–51.
- [21] Arber DA, Orazi A, Hasserjian R, et al. The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. Blood. 2016;127:2391–2405.
- [22] Livak KJ, Schmittgen TD. Analysis of relative gene expression data using realtime quantitative PCR and the 2(-Delta Delta C (T)) Method. Methods. 2001;25:402–408.
- [23] Yonezawa S, Higashi M, Yamada N, et al. Mucins in human neoplasms: clinical pathology, gene expression and diagnostic application. Pathol Int. 2011;61:697–716.
- [24] Sadras T, Heatley SL, Kok CH, et al. Differential expression of MUC4, GPR110 and IL2RA defines two groups of CRLF2-rearranged acute lymphoblastic leukemia patients with distinct secondary lesions. Cancer Lett. 2017;408:92–110.

- [25] Jonckheere N, Seuningen IV. Integrative analysis of the cancer genome atlas and cancer cell lines encyclopedia large-scale genomic databases: MUC4/MUC16/ MUC20 signature is associated with poor survival in human carcinomas. J Transl Med. 2018;16:259.
- [26] Andrianifahanana M, Singh AP, Nemos C, et al. IFNgamma-induced expression of MUC4 in pancreatic cancer cells is mediated by STAT-1 up regulation: a novel mechanism for IFN-gamma response. Oncogene. 2007;26:7251–7261.
- [27] Jonckheere N, Perrais M, Mariette C, et al. A role for human MUC4 mucin gene, the ErbB2 ligand, as a target of TGF beta in pancreatic carcinogenesis. Oncogene. 2004;23:5729–5738.
- [28] Yamada N, Nishida Y, Tsutsumida H, et al. Promoter CpG methylation in cancer cells contributes to the regulation of MUC4. Br J Cancer. 2009;100:344–351.
- [29] Lahdaoui F, Delpu Y, Vincent A, et al. miR-219-1-3p is a negative regulator of the mucin MUC4 expression and is a tumor suppressor in pancreatic cancer. Oncogene. 2015;34:780–788.
- [30] Mejías-Luque R, Peiró S, Vincent A, et al. IL-6 induces MUC4 expression through gp130/STAT3 pathway in gastric cancer cell lines. Biochim Biophys Acta. 2008;1783:1728–1736.
- [31] Stevens AM, Miller JM, Munoz JO, et al. Interleukin-6 levels predict event-free survival in pediatric AML and suggest a mechanism of chemotherapy resistance. Blood Adv. 2017;1:1387–1397.
- [32] Sanchez-Correa B, Bergua JM, Campos C, et al. Cytokine profiles in acute myeloid leukemia patients at diagnosis: survival is inversely correlated with IL-6 and directly correlated with IL-10 levels. Cytokine. 2013;61:885–891.
- [33] Han Y, Ye A, Bi L, et al. Th17 cells and interleukin-17 increase with poor prognosis in patients with acute myeloid leukemia. Cancer Sci. 2014;105:933–942.
- [34] Rakha EA, Boyce RW, Abd El-Rehim D, et al. Expression of mucins (MUC1, MUC2, MUC3, MUC4, MUC5AC and MUC6) and their prognostic significance in human breast cancer. Mod Pathol. 2005;18:1295–1304.
- [35] Shibahara H, Tamada S, Higashi M, et al. MUC4 is a novel prognostic factor of intrahepatic cholangiocarcinoma-mass forming type. Hepatology. 2004;39:220–229.
- [36] Shanmugam C, Jhala NC, Katkoori VR, et al. Prognostic value of mucin 4 expression in colorectal adenocarcinomas. Cancer. 2010;116:3577–3586.
- [37] Matull WR, Andreola F, Loh A, et al. MUC4 and MUC5AC are highly specific tumor-associated mucins in biliary tract cancer. Br J Cancer. 2008;98:1675–1681.
- [38] Workman HC, Sweeney C, Carraway KL. The membrane mucin Muc4 inhibits apoptosis induced by multiple insults via ErbB2-dependent and ErbB2-independent mechanisms. Cancer Res. 2009;69:2845–2852.
- [39] Skrypek N, Duchene B, Hebbar M, et al. The MUC4 mucin mediates gemcitabine resistance of human pancreatic cancer cells via the Concentrative Nucleoside Transporter family. Oncogene. 2013;32:1714–1723.
- [40] Ponnusamy MP, Singh AP, Jain M, et al. MUC4 activates HER2 signalling and enhances the motility of human ovarian cancer cells. Br J Cancer. 2008;99:520–526.