



# Abnormally Low HbA<sub>1c</sub> Caused by Hemolytic Anemia, a Case Report and Literature Review

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Hemoglobin A<sub>1c</sub> is a widely used diagnostic tool for monitoring glycemic control in diabetes management. However, its accuracy can be influenced by various factors. We present a case of a 17-year-old boy with abnormally low Hemoglobin A<sub>1c</sub> levels caused by warm autoantibody-induced hemolytic anemia. This case highlights the importance of considering conditions that may affect erythrocyte survival, and the potential interferences when interpreting Hemoglobin A<sub>1c</sub> results to ensure accurate diagnosis and effective management of diabetes.

**Keywords:** glycosylated hemoglobin, HbA<sub>1c</sub>, hemolytic anemia, enzymatic method, diabetes

## INTRODUCTION

Hemoglobin (Hb) is the protein contained in red blood cells (RBCs) that is responsible for delivery of oxygen to the tissues. Hb is composed of two pairs of dissimilar chains,  $\alpha$  and  $\beta$ , each defined by a specific amino acid sequence and incorporating an iron-containing heme group. Two  $\alpha$ - $\beta$  dimers combine to form a hemoglobin tetramer. In adults, Hemoglobin A (HbA) is the dominant type, accounting for around 97% of all hemoglobin. Minor variations of HbA can arise through post-translational modifications. These modified HbA include A<sub>1a</sub>, A<sub>1b</sub>, and A<sub>1c</sub>, with A<sub>1c</sub> being the most prevalent of these minor components [1].

Hemoglobin A<sub>1c</sub> (HbA<sub>1c</sub>) is formed through a non-enzymatic process called glycation, where glucose molecules bind to the amino groups of proteins. Specifically, glucose reacts with the N-terminal amino group of the hemoglobin beta-chain, resulting in the formation of a Schiff base. This reaction then undergoes a rearrangement to form HbA<sub>1c</sub>. Notably, this process is irreversible and depends on both the average glucose levels in the blood and the age of RBCs [2]. RBCs typically have a lifespan of approximately 120 days. Therefore, glycosylated hemoglobin reflects the average glucose levels over the past 60–90 days [3].

HbA<sub>1c</sub> concentration is a useful tool for monitoring glycemic control over time, as well as establishing treatment goals and decision boundaries [4, 5]. The HbA<sub>1c</sub> test indicates the average blood glucose level for the last 8–12 weeks [6]. The American Diabetes Association (ADA) has recommended an HbA<sub>1c</sub> level of  $\geq 6.5\%$  (47.5 mmol/mol) as the diagnostic threshold for diabetes since 2010 [7]. The recommendation to use HbA<sub>1c</sub> as a diagnostic test is based on its advantages over traditional glucose tests. HbA<sub>1c</sub> provides a better overall picture of glycemic exposure and long-term complication risk, and it is less susceptible to variations in biological and preanalytical factors. Moreover, HbA<sub>1c</sub> levels are not affected by sudden changes in glucose levels caused by acute illnesses or stress, making it a more reliable indicator of glycemic control in these situations [8].

Currently, various HbA<sub>1c</sub> assay methods are used in clinical practice, and significant efforts have been made towards global standardization. This standardization ensures good performance and

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reproducibility across different assays. It is achieved through traceability to the National Glycohemoglobin Standardization Program (NGSP) and the reference materials and methods of the International Federation of Clinical Chemistry (IFCC) [9, 10]. Recent guidelines recommend intra-laboratory and inter-laboratory coefficients of variation (CV) of <1.5% and <2.5%, respectively, which are achievable due to the efforts of global standardization programs [3].

Methods for measuring HbA<sub>1c</sub> can be broadly categorized into three groups based on their assay principles. The first group includes methods that detect the charge differences between glycosylated and non-glycosylated hemoglobin, such as ion-exchange high-performance liquid chromatography (HPLC). This method is widely used due to its high precision and ability to provide a complete hemoglobin profile. However, it can be affected by hemoglobin variants, leading to potential inaccuracies in patients with hemoglobinopathies [11]. The second group of methods separates glycosylated from non-glycosylated hemoglobin based on structural differences, as seen in affinity chromatography or immunoassays. Affinity chromatography specifically binds glycosylated hemoglobin, making it less affected by hemoglobin variants compared to ion-exchange HPLC [12]. Immunoassays, which use antibodies to recognize glycosylated hemoglobin, are both rapid and suitable for high-throughput laboratories. However, these methods can be prone to interference from endogenous antibodies, such as heterophile antibodies, which may result in inaccurate measurements [13, 14]. The third group of methods measures HbA<sub>1c</sub> based on its chemical reactivity, such as enzymatic assays. These methods offer the advantage of being less affected by hemoglobin variants or other structural alterations, and they tend to be quicker and simpler to perform than chromatographic methods [15].

The accuracy of HbA<sub>1c</sub> measurements can be affected by various factors. Pre-analytical factors can be classified into four primary categories: 1) erythropoiesis factors, such as folate and vitamin B12 deficiency; 2) hemoglobinopathies; 3) factors influencing glycation, such as alcoholism, renal failure, and the consumption of vitamins C and E; and 4) factors related to erythrocyte destruction, such as hemolytic anemia and certain drugs. Analytical factors include variations in reagent lot or those specific to antibody-based methods such as the presence of heterophile antibodies [9, 16]. Heterophile antibodies are non-specific antibodies that may bind to reagents used in the assay, such as monoclonal or polyclonal antibodies, leading to false results. Depending on the specific assay technique, these antibodies can either enhance or suppress the signal, resulting in inaccurate HbA<sub>1c</sub> measurements [14].

These factors may influence the measurement of HbA<sub>1c</sub> based on the method principle and it is essential to consider these factors when interpreting HbA<sub>1c</sub> results to ensure accurate conclusions. In this study, we present a case of abnormally low HbA<sub>1c</sub> caused by hemolytic anemia in a 17-year-old boy.

## CASE DESCRIPTION

Following a clinical examination that revealed abdominal pain, lateral edema, and splenomegaly, a 17-year-old boy presented to

**TABLE 1 |** Hematology findings of the patient.

Test	Result	Reference range
RBC (x10 <sup>12</sup> /L)	<b>2.56</b>	4.0–5.5
Hb (g/L)	<b>81</b>	120–160
HCT (L/L)	<b>0.25</b>	0.37–0.49
MCV (fL)	90.8	80–100
MCH (pg)	31.5	27–34
MCHC (g/L)	347	320–360
WBC (x10 <sup>9</sup> /L)	6.7	3.5–11.0
PLT (x10 <sup>9</sup> /L)	234	180–345
Reticulocyte (%)	<b>9.8</b>	0.5–2.5

Values in bold indicate results outside the reference interval.

**TABLE 2 |** Biochemistry findings of the patient.

Test	Result	Reference range
HbA <sub>1c</sub> (%)	<b>2.8</b>	4.0–6.0
FPG (mmol/L)	5.1	3.9–5.9
Urea (mmol/L)	7.1	3.0–7.5
Creatinine (μmol/L)	88.5	61.9–123.9
ALT (U/L)	<b>62</b>	<40
AST (U/L)	<b>78</b>	<37
LDH (IU/L)	<b>436</b>	<280
Total bilirubin (μmol/L)	<b>116.2</b>	1.7–20.5
Direct bilirubin (μmol/L)	<b>10.2</b>	1.7–5.1
TSH (mIU/L)	2.2	0.5–5.0
T4 (nmol/L)	79.3	60–150
Ferritin (μg/L)	71.7	16–220
25 (OH)D <sub>3</sub> (nmol/L)	126.3	75–250 (sufficient)

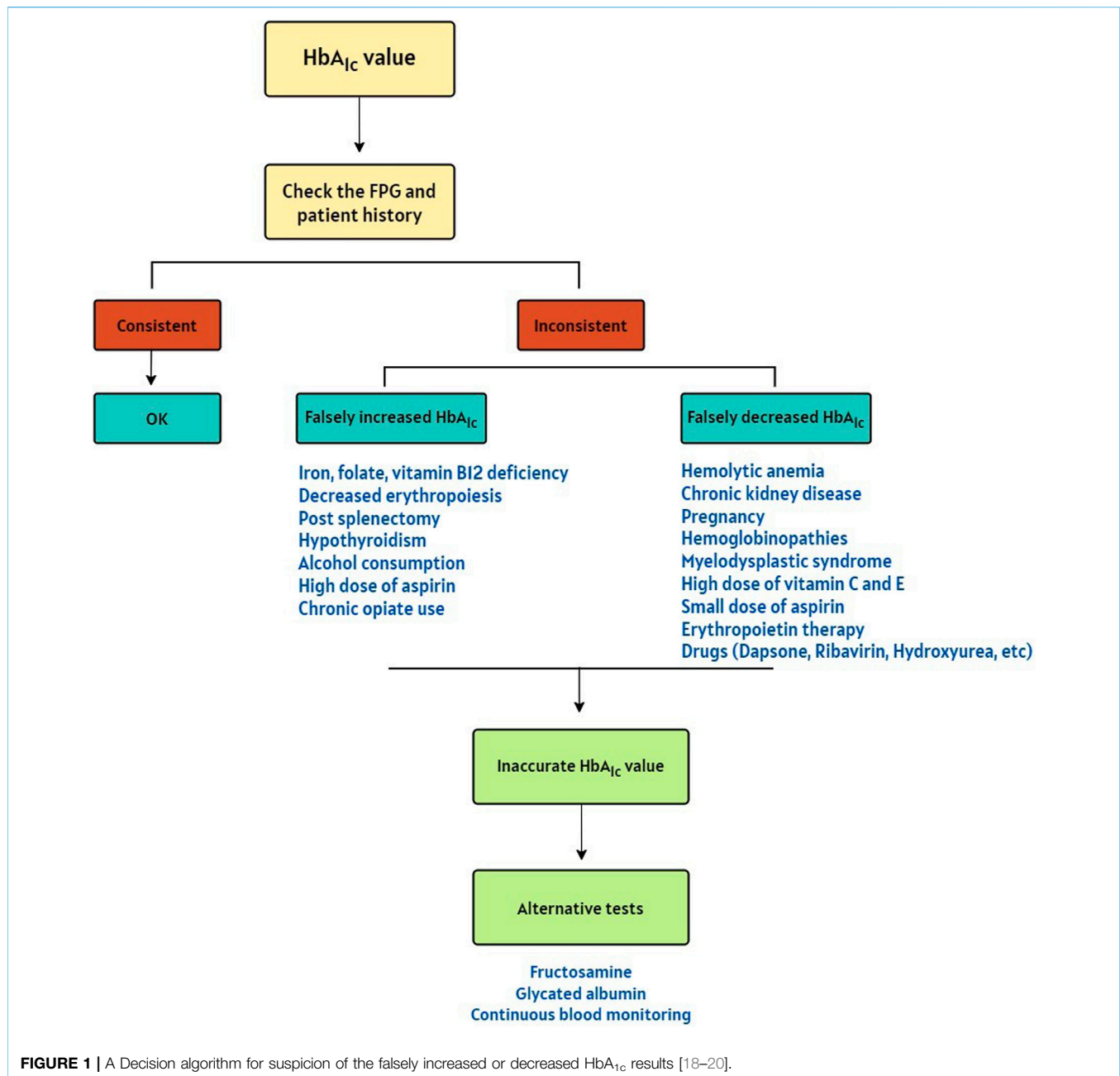
Values in bold indicate results outside the reference interval.

the laboratory for a blood test under a physician's prescription. The patient denied any significant medical history or recent drug use. He underwent routine laboratory tests, including screening for glucose metabolism, with HbA<sub>1c</sub> included as part of the standard check-up. During the investigation, an abnormally low HbA<sub>1c</sub> level, despite a normal fasting plasma glucose (FPG) level, was noted. HbA<sub>1c</sub> was measured using a modified enzymatic method, showing a significant decrease to 2.8% (7.0 mmol/mol), while the FPG level was 92 mg/dL (reference range: 70–106 mg/dL), which was discordant with the HbA<sub>1c</sub> result. The patient had no family history of hyper- or hypoglycemia.

Hematological findings revealed normocytic, normochromic anemia, with platelet (PLT) and white blood cell (WBC) counts remaining within the reference range (Table 1). Examination of the peripheral blood smear, stained with new methylene blue, demonstrated significant reticulocytosis at 9.8%.

The biochemical findings revealed mild elevations in serum alanine aminotransferase (ALT), aspartate transaminase (AST), and lactate dehydrogenase (LDH) levels and a significant increase in indirect bilirubin. Other biochemistry tests were in the reference range as shown in Table 2.

In this patient, the diagnosis of hemolytic anemia was unexpected. The direct antiglobulin test (DAT) revealed the presence of warm autoimmune hemolytic anemia. After exclusion of lymphoproliferative disorders, rheumatic disorders, non-lymphoid malignancies, and drug-induced



**FIGURE 1** | A Decision algorithm for suspicion of the falsely increased or decreased HbA<sub>1c</sub> results [18–20].

autoimmune hemolytic anemia, it was classified as idiopathic [17].

## DISCUSSION

HbA<sub>1c</sub> is a useful tool for glycemic control in individuals with diabetes mellitus. Discrepancies between HbA<sub>1c</sub> value, FPG levels, and patient history should prompt consideration of underlying factors that could lead to inaccurate HbA<sub>1c</sub> results. While HbA<sub>1c</sub> is generally a reliable indicator of glycemia, there are specific circumstances where its reliability is compromised

(Figure 1). Interfering factors are typically method-dependent and may lead to overestimation or underestimation of HbA<sub>1c</sub> in various mechanisms.

As in this case, hemolytic anemia is a condition marked by the premature destruction of erythrocytes, which can occur either intravascular or extravascular, primarily within the reticuloendothelial system [21]. This normocytic normochromic anemia is usually characterized by elevated indirect bilirubin, elevated LDH, decreased haptoglobin, increased reticulocyte count, and the presence of spherocytes in blood smear [22]. Warm autoantibody-induced hemolytic anemia can affect people of all ages, but this condition is more prevalent among those over

40 years old, with the peak incidence typically occurring in the 70s [23]. However, a study conducted at the Mayo Clinic from 1994 to 2014 on 35 pediatric patients (median age, 10 years old) with autoimmune hemolytic anemia revealed that warm antibodies were the underlying cause of hemolytic anemia in approximately 80% of the patients, consistent with our case [24]. Warm antibody hemolytic anemia is the primary type of autoimmune hemolytic anemia, accounting for approximately 80%–90% of cases. In this condition, warm-reactive antibodies, which are most active at body temperature (37°C), bind to RBCs and initiate a complement-mediated process that damages the cell membranes [25].

Hemoglobinopathies are the leading global cause of inherited single-gene disorders worldwide that are often associated with artificially altered HbA<sub>1c</sub> levels [26]. If a patient does not have HbA, as is the case in individuals with homozygous variants such as sickle cell disease, HbA<sub>1c</sub> testing should be avoided, and alternative tests should be used. For other variants, guidelines recommend that laboratories should be aware of the potential effects of hemoglobinopathies on their selected methods [3]. Some methods used to measure HbA<sub>1c</sub> can produce inaccurate results in patients with hemoglobinopathies, as these disorders can cause variant hemoglobin molecules to migrate similarly to HbA<sub>1c</sub> during testing, leading to co-elution and falsely increased or decreased results. Hemoglobin variants can also alter the glycation sites and therefore interfere with HbA<sub>1c</sub> assays [27]. The impact of hemoglobin variants on HbA<sub>1c</sub> measurements is method-dependent, and since they are present in nearly one-third of patients with diabetes, it is essential to acknowledge their influence [28].

In addition to Hb variants, chemically modified derivatives of Hb can impact the accuracy of these measurements, though the extent of this effect varies widely across different commercially available methods [29]. Carbamylated Hemoglobin (CarbHb) forms through a non-enzymatic reaction and increases with higher levels of urea-derived cyanate. This primarily occurs in patients with chronic kidney disease or those undergoing dialysis. CarbHb can co-elute with HbA<sub>1c</sub> during assays, potentially leading to falsely elevated results in some methods [30]. The effects of sulfhemoglobin, methemoglobin (MetHb), and acetylated hemoglobin on HbA<sub>1c</sub> measurements have been studied among chemically modified derivatives. However, these substances interfere less with newer methods. Sulfhemoglobin alters the absorption spectrum, leading to either falsely low or high results in spectrophotometric methods, depending on the assay type. This alteration can occur during the administration of sulfonamides [31]. MetHb, due to its oxidized iron, alters hemoglobin's optical properties and can interfere with HbA<sub>1c</sub> assays that use spectrophotometric or colorimetric techniques. Individuals exposed to oxidizing substances or with conditions like methemoglobinemia are more likely to experience interference with their HbA<sub>1c</sub> measurements from MetHb [32]. Acetylated hemoglobin, formed through reactions with acetyl groups (such as those from acetylated drugs like aspirin) can interfere with certain HbA<sub>1c</sub> assays by altering the charge of hemoglobin. This change may cause it to be misidentified as glycated hemoglobin, resulting in falsely elevated HbA<sub>1c</sub> levels [33].

Drugs can affect HbA<sub>1c</sub> levels in multiple ways such as oxidation of hemoglobin, subclinical hemolysis, shortened

survival of erythrocytes, etc. Despite the theoretical possibility, there have been only a few documented instances of drug-induced variability in HbA<sub>1c</sub> levels reported in the scientific literature, including dapsone, ribavirin, antiretrovirals, aspirin, hydroxyurea, and Trimethoprim-Sulfamethoxazole [18].

Pregnancy, splenectomy, myelodysplastic syndrome, blood loss, folate, and B12 deficiency can affect the accuracy of the HbA<sub>1c</sub> test by impacting the survival and lifespan of RBCs as another mechanism in which interference can occur. Additionally, circumstances like iron deficiency anemia, and consumption of alcohol, small doses of aspirin, vitamins C and E can influence the glycation process, potentially leading to erroneous HbA<sub>1c</sub> results [18, 19].

Notably, population data have shown that HbA<sub>1c</sub> results may be affected by some patient variables. Age-related increases of approximately 0.1% per decade after 30 years of age have been observed in individuals without diabetes [34]. Although research on the impact of ethnicity on HbA<sub>1c</sub> results is inconsistent, some evidence suggests that Black and Hispanic populations may have higher HbA<sub>1c</sub> values than White populations at the same level of glycemia [35, 36]. A meta-analysis of 12 studies involving 49,238 individuals has shown that mean HbA<sub>1c</sub> levels are 0.26%, 0.24%, and 0.08% higher in Black, Asian, and Hispanic individuals, respectively, compared to White individuals [37]. However, another study did not find any difference between Black and White individuals [38]. While seemingly modest (<1%), these differences could have significant clinical implications if these populations' decision thresholds are not appropriately adjusted. For instance, the observed disparities may lead to an overestimation of HbA<sub>1c</sub> levels in Black and Asian individuals, potentially resulting in the over-diagnosis of diabetes mellitus. This is particularly critical when HbA<sub>1c</sub> values are near clinical decision limits, where small differences can influence diagnostic or treatment decisions. Furthermore, elevated HbA<sub>1c</sub> levels in older individuals without diabetes mellitus may similarly lead to over-diagnosis, potentially exposing individuals to unnecessary treatment and causing increased financial burdens for healthcare systems. Further work is required to elucidate these relationships, but clinical laboratories should be aware of the potential clinical significance of these factors.

The case highlights a key limitation of relying on HbA<sub>1c</sub> as a standalone diagnostic test for diabetes, as recommended by the ADA and other guidelines. While HbA<sub>1c</sub>'s long-term glycemic representation makes it a valuable diagnostic tool, conditions such as hemolytic anemia that alter red blood cell turnover can lead to misleading results. In this case, the abnormal HbA<sub>1c</sub> value was identified because it was compared with FPG and clinical findings. Without these additional measures of glycemic control, the low HbA<sub>1c</sub> value could have been misinterpreted, potentially leading to underdiagnosis or inappropriate clinical decisions. This underscores the importance of integrating HbA<sub>1c</sub> with other diagnostic approaches. In cases where the HbA<sub>1c</sub> value does not align with the FPG and patient history, interfering factors may be responsible for over- or under-estimation. These factors are summarized in the decision algorithm depicted in **Figure 1**. In such situations, alternative methods

can be employed to assess glycemic control. Fructosamine and glycosylated albumin testing measure average blood glucose levels over the past 2-3 weeks, reflecting short-term glucose control more accurately. Self-monitoring of blood glucose also offers a snapshot of blood glucose levels at a specific point in time, allowing patients and healthcare providers to track changes in glucose levels [39]. By combining these methods, healthcare providers can gain a more comprehensive understanding of a patient's glucose levels and make informed treatment decisions.

This study has limitations that should be acknowledged. While direct insights from the patient regarding their perspective on the condition were not available, it is well-documented that individuals diagnosed with autoimmune hemolytic anemia often experience significant physical and emotional challenges. However, this case report is limited by the lack of follow-up data, including details on therapeutic interventions and patient outcomes. Consequently, we could not evaluate the effectiveness of standard treatments in this specific case. Future studies or reports with comprehensive follow-up are necessary to provide a more holistic understanding of patient management and perspectives in similar contexts.

## CONCLUSION

In conclusion, HbA<sub>1c</sub> measurement is a widely used diagnostic tool for monitoring glycemic control in diabetes management. However, it is essential to consider the various influencing and interfering factors that can affect its accuracy. To ensure accurate diagnosis and effective management of diabetes, it is crucial to consider these factors when interpreting HbA<sub>1c</sub> results. Alternate methods, such as fructosamine, glycosylated albumin, and glucose monitoring should be used to assess glycemic control when HbA<sub>1c</sub> results are inaccurate.

## Take-home messages and learning points

- HbA<sub>1c</sub> is a useful tool for glycemic control
- The ADA has considered HbA<sub>1c</sub> ≥ 6.5% diagnostic criteria for diabetes
- Hemolytic anemia is normocytic normochromic anemia that results in an inaccurate HbA<sub>1c</sub> measurement
- Alternative methods in case of inaccurate HbA<sub>1c</sub> results should be considered

## DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding author.

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## ETHICS STATEMENT

The study protocol was approved by the Ethical Committee of Shahid Beheshti University of Medical Sciences in Tehran, Iran, and all procedures followed were in accordance with the ethical standards of the responsible committee. Written informed consent was obtained from the legal guardian of the patient for the publication of any potentially identifiable images or data included in this article.

## AUTHOR CONTRIBUTIONS

SB: conceptualization; writing - original draft; writing - review and editing. NT and SD'A: writing - review and editing. All authors contributed to the article and approved the submitted version.

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## CONFLICT OF INTEREST

Author SD'A was employed by Viollier AG.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## GENERATIVE AI STATEMENT

The author(s) declare that no Generative AI was used in the creation of this manuscript.

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