

Gamma-delta T cell expansion is closely associated with cytomegalovirus infection in all solid organ transplant recipients

doi:10.1111/j.1432-2277.2010.01181.x

We read with interest the manuscript published by Puig-Pey *et al.* entitled “Characterisation of gamma-delta T cells in organ transplantation” [1]. For a better reading of this paper, we would like to add several comments on the basis of our experience in $\gamma\delta$ T cell

immunomonitoring in kidney transplant recipients (KTR), to enlighten their role in the context of cytomegalovirus (CMV) infection. Indeed, in KTR, we demonstrated, 10 years ago, that CMV infection is the sole parameter independently associated with a persistent

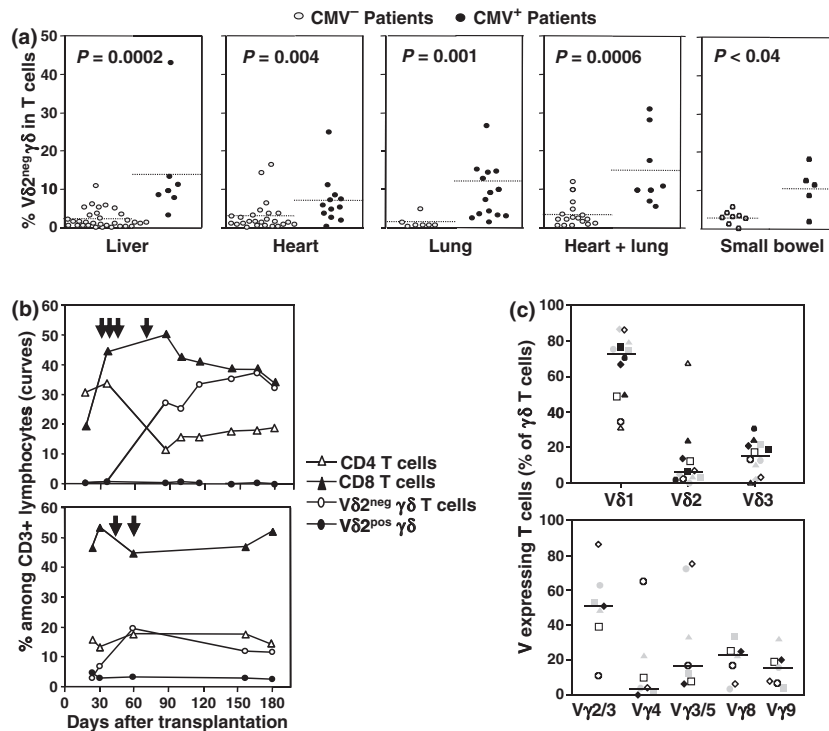


Figure 1 (a) High percentages of $V\delta 2^{neg}$ $\gamma\delta$ T cells are associated with CMV infection in the heart, liver, lung, lung + heart and small bowel transplant recipients. The percentage of $V\delta 2^{neg}$ $\gamma\delta$ T cells in the peripheral blood of liver, heart, lung, lung + heart and small bowel transplant recipients was determined by flow cytometry and compared between patients who suffered from CMV infection post-transplantation (CMV^+ patients, full circles) and those who did not (CMV^- patients, empty circles). Dashed lines represent the mean value for each group of patients. The statistical differences between groups of CMV^- and CMV^+ donors were tested using the unpaired Mann–Whitney *U*-test. (b) $V\delta 2^{neg}$ $\gamma\delta$ T cell expansion occurs during CMV infection in transplant patients. The course of CD4+, CD8+, $V\delta 2^{neg}$ $\gamma\delta$ T cells and $V\delta 2^{pos}$ $\gamma\delta$ T cell subsets was longitudinally followed post-transplantation and are represented in one liver-recipient (upper panel) and in one lung-recipient (lower panel) suffering from CMV infection, as detected by pp65 antigenemia or CMV DNA in bronchoalveolar liquid (black arrows), respectively. It is noteworthy that $V\delta 2^{neg}$ $\gamma\delta$ T cell expansion was followed in both patients by the resolution of CMV infection. $\gamma\delta$ T cell percentages remained stable at least 6 months post-transplantation. (c) Combinatorial diversity of $\gamma\delta$ T cells in CMV^+ lung- and/or heart-transplanted patients. The $V\delta 1,2,3$ T-cell receptor variable region expression by $\gamma\delta$ T cells was analyzed by flow cytometry in three lung, one heart and eight lung + heart recipients (upper panel), and $V\gamma 2,3,4,5,8,9$ expression was analyzed in seven of these recipients (lower panel). Analysis was carried out in whole blood by triple labeling with anti- $V\delta$ or γ , anti-pan δ and anti-CD3 monoclonal antibodies. $\gamma\delta$ mainly expressed the $V\delta 1$ and $V\delta 3$ variable regions of the TCR.

expansion of circulating $\gamma\delta$ T cells [2]. This expansion is associated with the resolution of the infection, and late $\gamma\delta$ T cell expanders suffer from longer infections, suggesting an anti-viral role for these cells *in vivo* [3]. Since then, CMV-associated expansion of $\gamma\delta$ T cells has also been observed by other groups in immunodeficient children [4,5], neonates [6], stem cell transplant recipients [7], and healthy volunteers [8].

As confirmed by Dr Sanchez-Fueyo's group, we have demonstrated that only $V\delta 2^{neg}$ $\gamma\delta$ T cells (including $V\delta 1$ and $V\delta 3$ T cells) are involved in this stable expansion over-time [9], and that they display a highly biased repertoire suggestive of antigen-driven selection or CMV-mediated amplification of an initially restricted $V\delta 2^{neg}$ $\gamma\delta$ T cell population [9]. The $V\delta 2^{neg}$ $\gamma\delta$ T cells expand concomitantly with CMV-specific CD8+ T cells in the blood of KTR and share a similar effector/memory T_{EMRA} phenotype [8,10]. *In vitro*, they display T-cell receptor-dependent cytotoxicity against CMV-infected cells and inhibit CMV multiplication through interferon- γ production [11].

Puig-Pey *et al.* show that HCV infection is also associated with an increase in the $V\delta 1/V\delta 2$ ratio, but the significance is low ($P = 0.046$). Moreover, they do not mention whether a significant increase in $V\delta 1$ $\gamma\delta$ T cells is observed in HCV-infected liver transplant recipients (LTR). This is an important issue because in our previous study in KTR, the infections with other viruses than CMV (including HCV) were conversely associated with a reduced number of $\gamma\delta$ T cells. If this decrease concerns only $V\delta 2$ T cells, this could explain the increase in the $V\delta 1/V\delta 2$ ratio without the need for a rise in $V\delta 1$ number. Consequently, we think that further investigations such as multivariate analysis are mandatory to confirm a relationship between HCV and $V\delta 2^{neg}$ $\gamma\delta$ T cell expansion.

Previous studies by Dr Sanchez-Fueyo's group have concluded that $V\delta 1$ $\gamma\delta$ T cell expansions in LTR are associated with operational tolerance [12,13]. However, in their present paper, they demonstrate that quantification of $V\delta 1$ $\gamma\delta$ T cells does not allow for accurate discrimination between tolerant LTR and recipients requiring maintenance immunosuppression. On the contrary, they confirmed that "CMV infection constitutes the main force shaping the repertoire of peripheral blood $V\delta 1$ $\gamma\delta$ T cells" in LTR and KTR. We have also shown that CMV-mediated expansion of $V\delta 2^{neg}$ $\gamma\delta$ T cells is a general phenomenon observed in all CMV-infected solid-organ transplant recipients (Fig. 1). The percentage of $V\delta 1$ $\gamma\delta$ T cells they found in CMV-negative LTR is equivalent to that of healthy individuals (compare figure 4 and figure 1 of their paper). This result confirms that "transplantation procedure per se" or "the chronic exposure to pharmacological IS" cannot induce $V\delta 2^{neg}$ $\gamma\delta$ T cells expansion in the absence of CMV infection. The high prevalence of

CMV infection in the LTR cohort of Dr Sanchez-Fueyo's group (90%) may explain why they missed the critical link between this virus and $V\delta 1$ $\gamma\delta$ T cell expansion previously.

As $V\delta 2^{neg}$ $\gamma\delta$ T cell expansion is a cell signature of CMV infection, monitoring of these cells after transplantation holds promising interest. Moreover, their ability to kill both CMV-infected cells and carcinoma cells *in vitro* [11], and their association with reduced cancer risk in CMV-infected KTR [14], make their close monitoring of particular relevance to post-transplant outcomes.

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Conflict of interest

The authors have no conflicting financial interests. There is no study sponsor.

References

1. Puig-Pey I, Bohne F, Benitez C, *et al.* Characterization of gammadelta T cell subsets in organ transplantation. *Transpl Int* 2010.
2. Dechanet J, Merville P, Berge F, *et al.* Major expansion of gammadelta T lymphocytes following cytomegalovirus infection in kidney allograft recipients. *J Infect Dis* 1999; **179**: 1.
3. Lafarge X, Merville P, Cazin MC, *et al.* Cytomegalovirus infection in transplant recipients resolves when circulating gammadelta T lymphocytes expand, suggesting a protective antiviral role. *J Infect Dis* 2001; **184**: 533.
4. de Villartay JP, Lim A, Al-Mousa H, *et al.* A novel immunodeficiency associated with hypomorphic RAG1 mutations and CMV infection. *J Clin Invest* 2005; **115**: 3291.
5. Ehl S, Schwarz K, Enders A, *et al.* A variant of SCID with specific immune responses and predominance of gamma delta T cells. *J Clin Invest* 2005; **115**: 3140.
6. Vermijlen D, Brouwer M, Donner C, *et al.* Human cytomegalovirus elicits fetal gammadelta T cell responses *in utero*. *J Exp Med* 2010; **207**: 807.

7. Knight A, Madrigal AJ, Grace S, *et al.* The role of Vdelta2-negative gamma-delta T cells during cytomegalovirus reactivation in recipients of allogeneic stem cell transplants. *Blood* 2010; **116**: 2164.
8. Pitard V, Roumanes D, Lafarge X, *et al.* Long term expansion of effector/memory V{delta}2neg {gamma}{delta} T cells is a specific blood signature of CMV infection. *Blood* 2008; **112**: 1317.
9. Dechanet J, Merville P, Lim A, *et al.* Implication of gammadelta T cells in the human immune response to cytomegalovirus. *J Clin Invest* 1999; **103**: 1437.
10. Couzi L, Pitard V, Netzer S, *et al.* Common features of gammadelta T cells and CD8(+) alphabeta T cells responding to human cytomegalovirus infection in kidney transplant recipients. *J Infect Dis* 2009; **200**: 1415.
11. Halary F, Pitard V, Dlubek D, *et al.* Shared reactivity of V{delta}2(neg) {gamma}{delta} T cells against cytomegalovirus-infected cells and tumor intestinal epithelial cells. *J Exp Med* 2005; **201**: 1567.
12. Li Y, Koshiba T, Yoshizawa A, *et al.* Analyses of peripheral blood mononuclear cells in operational tolerance after pediatric living donor liver transplantation. *Am J Transplant* 2004; **4**: 2118.
13. Martinez-Llordella M, Puig-Pey I, Orlando G, *et al.* Multi-parameter immune profiling of operational tolerance in liver transplantation. *Am J Transplant* 2007; **7**: 309.
14. Couzi L, Levaillant Y, Jamaï A, *et al.* Cytomegalovirus-induced gammadelta T cells associate with reduced cancer risk after kidney transplantation. *J Am Soc Nephro* 2010; **21**: 181.