## ORIGINAL ARTICLE

# Renal resistance thresholds during hypothermic machine perfusion and transplantation outcomes – a retrospective cohort study

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#### **SUMMARY**

Renal resistance (RR), of allografts undergoing hypothermic machine perfusion (HMP), is considered a measure of organ quality. We conducted a retrospective cohort study of adult deceased donor kidney transplant (KT) recipients whose grafts underwent HMP. Our aim was to evaluate whether RR is predictive of death-censored graft failure (DCGF). Of 274 KT eligible for analysis, 59% were from expanded criteria donor. RR was modeled as a categorical variable, using a previously identified terminal threshold of 0.4, and 0.2 mmHg/ml/min (median in our cohort). Hazard ratios (HR) of DCGF were 3.23 [95% confidence interval (CI): 1.12-9.34, P = 0.03] and 2.67 [95% CI: 1.14–6.31, P = 0.02] in univariable models, and 2.67 [95% CI: 0.91–7.86, P = 0.07] and 2.42 [95% CI: 1.02–5.72, P = 0.04] in multivariable models, when RR threshold was 0.4 and 0.2, respectively. Increasing risk of DCGF was observed when RR over the course of HMP was modeled using mixed linear regression models: HR of 1.31 [95% CI: 1.07-1.59, P < 0.01] and 1.25 [95% CI: 1.00–1.55, P = 0.05], in univariable and multivariable models, respectively. This suggests that RR during HMP is a predictor of long-term KT outcomes. Prospective studies are needed to assess the survival benefit of patients receiving KT with higher RR in comparison with staving wait-listed.

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#### Key words

graft failure, hypothermic machine perfusion, kidney transplantation, renal resistance

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## Introduction

Hypothermic machine perfusion (HMP) is a preservation technique that has been shown to decrease ischemia reperfusion injury and delayed graft function (DGF) and improve graft outcomes in kidney transplant (KT) recipients [1–5]. Parameters such as pressure, resistance, time, and flow are continuously measured during HMP, and those have been linked to short-term graft outcomes. Although, lower flow and higher renal resistance (RR) were associated with higher DGF [6– 20], some publications suggest that RR during HMP demonstrated a poor predictive performance for the same outcome [8,11]. Table 1 summarizes the evidence, to date, on the relationship between pump parameters and KT outcomes.

Despite contradictory reports on the relationship between RR and short-term graft outcomes, RR has

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| Reference   | Machine used                              | Parameter                       | Threshold                                  | Unit employed<br>for RR                        | size*                  | type                           | Follow up                     | Outcome  |
|---|---|---------------------------------|--|--|------------------------|--------------------------------|-------------------------------|--|
| Sandal e <i>t al.</i> [5]                               | LifePort IL                               | Resistance                      | 0.4 and 0.2                                | mmHg/ml/min                                    | 274                    | 59% ECD                        | 5 years                       | RR during HMP is associated with   |
| Parikh e <i>t al.</i> [7]                               | LifePort IL                               | Flow and                        | None                                       | mmHg/ml/min                                    | 671                    | 31% ECD,                       | 6 months                      | 1-h flow associated with DGF and   |
| Burgos Revilla <i>et al.</i> [8]                        | LifePort IL                               | resistance<br>Resistance        | 0.3 and 0.4                                | mmHg/ml/min                                    | 06                     | 26% UCD<br>100% ECD            | 1 year                        | Iast KK with higher 6-month eGFK<br>RR not predictive of graft survival      |
| Yushkov <i>et al.</i> [9]                               | LifePort IL                               | Resistance                      | <0.2, 0.2–0.3,<br>>0 3                     | ?mmHg/ml/min                                   | 454                    | 41% ECD,<br>10% DCD            | 1 year                        | RR >0.3 was predictive of 1-year<br>graft survival                           |
| Patel <i>et al.</i> (pg 2207) [19]                      | LifePort IL                               | Flow and                        | None                                       | mmHg/ml/min                                    | 190                    | N/A                            | 1 year                        | Initial resistance associated with 1-  |
| Patel et al. (pg 2202) [19]                             | LifePort IL                               | resistance<br>Pressure          | 23 mmHg                                    | mmHg/ml/min                                    | 73                     | 27% ECD,                       | 1 year                        | year graft survival<br>Higher pressure associated with an                    |
|   |   |                                 |  |  |                        | 4%DCD                          |                               | increased risk for DGF but similar 1-<br>year graft survival                 |
| de Vries et al. [10]                                    | Gambro PF-3B                              | Resistance                      | ≤1, 1–1.99, ≥2                             | mmHg/ml/min                                    | 440                    | All DCD                        | 15 years                      | Initial RR associated with PNF and   |
|   |   |                                 |  | per 100 g kidney<br>weight                     |                        |                                |                               | DGF but not graft survival   |
| Jochmans <i>et al.</i> [11]                             | LifePort IL                               | Resistance                      | None                                       | mmHg/ml/min                                    | 336                    | 27% ECD,<br>12% DCD            | 1 year                        | Terminal RR predictive of graft<br>failure                                   |
| Matsuno et al. [12]                                     | LPS-2, Nikiso,                            | Flow                            | 0.45-0.90                                  | ml/min/g machine                               | 88                     | 84% DCD                        | Immediate                     | Low flow associated with PNF and   |
|   | Japan                                     |                                 |  | perfusion flow                                 | Ĺ                      |                                |                               | ATN  |
| Nyberg <i>et al.</i> [14]                               | MUX100, MIN                               | index                           | cc.0-7.0                                   | renal mass                                     | G24                    | MA                             | 4 year                        | kk associated with interior graft<br>function                                |
| Mozes <i>et al.</i> [13]                                | Waters RM-3                               | Resistance                      | ≤0.4 vs. 0.4–0.6                           | mmHg/ml/min                                    | 280                    | N/A                            | Immediate                     | RR not predictive of immediate   |
| Kwiatkowski <i>et al.</i> [15]                          | MOX 100, MN                               | Resistance                      | None                                       | mmHg/ml/min/g                                  | 260                    | N/A                            | Immediate                     | RR associated with DGF   |
| Polyak et al. [17]                                      | MOX 100, MN                               | Resistance                      | None                                       | NA   | 111                    | All ECD†                       | Immediate                     | Higher RR predictive of early graft<br>dvsfunction                           |
| Tesi <i>et al.</i> [18]                                 | MOX 100, MN                               | Resistance<br>and flow          | ≤0.4, ≥70<br>for flow                      | mmHg/ml/min                                    | 82                     | N/A                            | 2 year                        | Pump parameters not predictive of DGF  |
| Henry <i>et al.</i> [16]<br>Sampson <i>et al.</i> [6]   | MOX 100, MN<br>Belzer LI-400 &<br>MOX 100 | Resistance<br>Flow              | None<br><80, 80–100,<br>>100               | WA<br>ml/min                                   | 254<br>100             | N/A<br>N/A                     | lmmediate<br>1 year           | RR >0.289 predictive of DGF<br>Flow not associated with creatinine<br>values |
| ATN, acute tubular necrosi<br>eGFR, estimated glomerula | is; DCD, donation<br>r filtration rate; E | n after cardiac<br>CD, expandec | : death; DCGF, dea<br>1 criteria donor; N/ | ath censored graft fa<br>A, not available; PNF | ilure; DC<br>, primary | GS, death cer<br>non function; | sored graft :<br>RR, renal re | survival; DGF, delayed graft function; sistance.                             |

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FECD defined as all grafts needing a biopsy.

\*Some kidneys were discarded.

been used as an indicator of graft quality, at times to guide decisions on organ utilization [21,22]. It has been reported that more than 15% of kidneys undergoing HMP are discarded annually in the USA, partly based on elevated RR [22,23]. This is despite a paucity of evidence on the predictive value of these parameters in relation to graft outcomes beyond 1 year. Hence, many have advocated against discarding kidneys strictly based on parameters during HMP [11,13,22–24]. In the light of these knowledge gaps, the aim of our study was to evaluate RR as an independent predictor of long-term graft survival in KT recipients whose allografts underwent HMP.

# Methods

# Study design and population

We conducted a retrospective cohort study in adult, deceased donor KT recipients at the McGill University Health Centre (MUHC) between November 1, 2007 and October 31, 2013. Multi-organ transplant recipients and grafts with missing perfusion parameters were excluded. The Research Ethics Board at the MUHC approved this study.

# Data sources

Recipient and donor demographics, histocompatibility data, laboratory investigations, treatments employed, histology data, and clinical outcomes were obtained from our in-centre transplant database. Pump parameters measured at the time of HMP were downloaded from the LifePort apparatus.

# Immunosuppression

Until 2011, the induction immunosuppression protocol consisted of anti-thymocyte globulin (Thymoglobulin<sup>®</sup>; Genzyme Canada, Mississauga, ON, Canada) and intraoperative methylprednisolone. Maintenance regimen included tacrolimus (Prograf®; Astellas, Mississauga, ON, Canada), mycophenolate mofetil (CellCept<sup>®</sup>; Hoffmann-LaRoche, ON, Canada), and prednisone. From November 2011 onwards, the induction immunosuppression protocol consisted of alemtuzumab (Campath<sup>®</sup>; Genzyme Canada) and intraoperative methylprednisolone. Maintenance regimen was long-acting tacrolimus (Advagraf<sup>®</sup>; Astellas Pharmaceuticals, Markham, ON, Canada) and mycophenolate sodium (Myfortic<sup>®</sup>; Novartis, Dorval, QC, Canada).

# Organ preservation

Kidnevs were recovered from adult deceased donors and flushed with SPS-1 (Belzer solution; Organ Recovery Systems (ORS), Itasca, IL, USA). Kidneys were shipped on ice. On arrival to our centre, grafts were placed on the LifePort Kidney Transporter device (ORS), and perfused with KPS-1 solution (ORS) supplemented with mannitol (2.5 g/l). Initially, the device was set for a systolic pressure of 30 mmHg. At the discretion of the surgeon on call, occasionally, this was reduced to 25 mmHg when flow rates were extremely high. HMP was continued until transplantation. At our center, HMP is considered the standard of care and is used whenever possible for deceased donor KT. By agreement of all surgeons and nephrologists involved in the transplant program, organs that were deemed clinically acceptable for transplantation were not discarded based on pump parameters alone.

# Exposure and outcome measurements

The exposure of interest was RR calculated using the formula pressure (mmHg)/flow (ml/min). Terminal RR, representing the mean RR measured over the last 15 min of HMP, was modeled as a categorical variable using a threshold of 0.4 mmHg/ml/min, based on prior publications [11]. We also used a threshold of 0.2 mmHg/ml/min, representing the median terminal RR in our study sample. Finally, to flexibly capture the relationship between our study endpoints and RR over the entire course of HMP, we used mixed linear regression models assuming a random slope and intercept [25,26]. The main outcome of interest was death-censored graft failure (DCGF). All-cause graft failure, patient death with function, DGF, and primary nonfunction (PNF) were secondary endpoints.

# Definitions

Expanded criteria donor (ECD) refers to all brain-dead donors aged 60 years or older, and donors aged 50– 59 years with two or more of the following comorbidities: history of hypertension, death resulting from cerebrovascular accident, and terminal serum creatinine  $\geq$ 1.5 mg/dl or 133 µmol/l. Standard criteria donors (SCD) included all non-ECD donors. Donation after cardiac death (DCD) donors was those designated by the United Network for Organ Sharing as nonheartbeating donors. DGF was defined as the need for dialysis within the first week post-transplant. PNF was defined as the failure of the transplanted kidney to function within the first 3 months post-transplant. Estimated GFR (eGFR) was calculated using the chronic kidney disease epidemiology collaboration equation [27].

## Model covariates

Recipient characteristics that were considered for inclusion in the model included recipient age, sex, race, BMI, peak panel reactive antibody, cause of end-stage renal disease, and dialysis modality. Donor characteristics included age, sex, race, BMI, type of donor (ECD, DCD), and terminal eGFR. Other transplant variables included cold ischemia time (CIT), human leukocyte antigen mismatch, and, induction and maintenance immunosuppression regimens. Dialysis modality was missing in 6% of the final cohort, and a missing indicator was used in the multivariable analysis. Other variables were missing in  $\leq$ 5% of the analytic cohort.

## Statistical analysis

The distributions of recipient, donor, and transplant characteristics at the time of transplantation were evaluated by RR categories using the t-test or Kruskal-Wallis test for continuous variables, and chi-square or Fisher's exact test for categorical variables. The relationships between terminal RR thresholds, modeled as categorical variables, (using thresholds of 0.4 and 0.2 mmHg/ml/min) and DCGF as well as all-cause graft failure and death with function, were estimated using the Kaplan-Meier method. Univariable and multivariable Cox proportional hazards models were employed to assess the independent relationship between terminal RR and time-to-event outcomes. RR estimates from the mixed linear regression models were similarly incorporated into univariable and multivariable Cox proportional hazards models [25,26]. To ensure the relationship was robust, we conducted sensitivity analyses whereby the models were also fit in a subcohort excluding patients with PNF. We were planning to introduce an interaction term and assess whether the effect of RR on study endpoints was modified within prespecified subgroups; however, the number of events was too small to conduct such an analysis. Statistical analyses were performed using STATA/IC 12.0 (StataCorp, College Station, TX, USA, www.stata.com) and R version 3.4.2 (R Foundation for Statistical Computing, Vienna, Austria, www.R-projects. org). Missing covariate data were handled by multiple imputations using STATA's "ice" command and R's "mice" package. A two-tailed P value of <0.05 was considered statistically significant.

## Results

During the study period, 422 deceased donor KT were performed. Of these, 274 were eligible for analysis. The study flow diagram is presented in Fig. 1. In our cohort, 59% of the KT were from ECD and 8% from DCD donors. Mean donor eGFR was  $98.30 \pm 29.32$  ml/min/ 1.73 m<sup>2</sup>. Allografts underwent HMP over a median of 8.5 h [95% confidence interval (95% CI): 6.1–11.5]. In most allografts, RR measurements decreased to a trough corresponding to the terminal RR. In some allografts, however, temporal changes in RR from initiation of HMP to transplantation demonstrated other patterns (e.g. U-shaped curve or rising to a peak corresponding to terminal RR measurement). Figure 2 presents changes in RR measurements over the course of HMP in eight allografts from our cohort.

## Baseline characteristics by terminal RR category

Baseline donor, recipient, and transplant characteristics of the analytic cohort are presented in Table 2. Of the 274 kidneys, 259 (94.5%) had a RR of <0.4 mmHg/ml/min and 15 (5.5%) had a RR of  $\geq$ 0.4 mmHg/ml/min. Donor eGFR was lower in allografts with a higher RR measuring 99.47  $\pm$  28.72 in those with RR <0.4 mmHg/ml/min vs. 78.43  $\pm$  33.14 in those with RR  $\geq$ 0.4 mmHg/ml/min (P < 0.01). Otherwise, donor and recipient characteristics were equally distributed. When applying the median



Figure 1 Study flow diagram.



Figure 2 Patterns of change in renal resistance over the course of hypothermic machine perfusion.

terminal RR in our population as a threshold (0.2 mmHg/ml/min), kidneys with higher RRs tended to be from female donors (50% vs. 38%, P = 0.01) and had a lower eGFR (102.72 ± 27.51 vs. 94.49 ± 30.36, P = 0.01).

#### Death-censored graft failure

Median follow-up in our cohort was 2.9 years [interguartile range (IQR): 1.3-4.4]. By the end of the study, 80% of the recipients were alive with a functioning graft, 10% experienced graft loss, and 10% died with a functioning graft. Kaplan-Meier curves of DCGF by terminal RR thresholds are depicted in Fig. 3. Results of univariable and multivariable Cox proportional hazards models evaluating the association between terminal RR and DCGF are presented in Table 3. An increased risk of DCGF was observed in univariable models with hazard ratio (HR) of 3.23 [95% CI: 1.12–9.34, P = 0.03] and 2.67 [95% CI: 1.14–6.31, P = 0.02] when applying terminal RR thresholds ≥0.4 and ≥0.2 mmHg/ml/min, respectively. In multivariable models (adjusted for ECD, CIT, DGF, and recipient age), HRs of 2.67 [95% CI: 0.91-7.86, P = 0.07] and 2.42 [95% CI: 1.02–5.72, P = 0.04] were observed when applying terminal RR thresholds of 20.4 and ≥0.2 mmHg/ml/min, respectively. An increasing risk of DCGF was also observed when repeated measurements of RR over the course of HMP were modeled using mixed linear regression models: HR 1.31 [95% CI: 1.07-1.59, P < 0.01] and 1.25 [95% CI: 1.00–1.55, P = 0.05] in univariable and multivariable analysis, respectively.

## Secondary endpoints

Kaplan–Meier curves for all-cause graft failure and death with function by terminal RR thresholds are

depicted in Fig. 3. When RR representing measurements over the course of HMP were modeled using mixed linear regression models, HRs for all-cause graft failure and death with function of 1.20 [95% CI: 0.98–1.45, P = 0.07] and 1.04 [95% CI: 0.75–1.45, P = 0.79] were observed in univariable analysis, and HR of 1.08 [95% CI: 0.88–1.34, P = 0.49] and 0.91 [95% CI: 0.64–1.28, P = 0.59] were observed in multivariable analysis (adjusted for ECD, DGF, and recipient age), respectively.

Delayed graft function was observed in 66 (24.1%) and PNF in 8 (2.9%) of the KT recipients. When applying a threshold of 0.4 mmHg/ml/min, 23.5% with RR <0.4 and 33.3% with RR  $\geq$ 0.4 experienced DGF. When applying a threshold of 0.2 mmHg/ml/min, a 23.2% with RR <0.2 and 24.8% with RR  $\geq$ 0.2 experienced DGF. An adjusted odds ratio for DGF, when RR measurements over the course of HMP were modeled using mixed linear regression models, was 1.01 [95% CI: 0.77–1.32, *P* = 0.95].

#### Sensitivity analysis

When repeating the analysis in a subcohort that excluded patients with PNF, univariable analysis demonstrated HR of 2.28 [95% CI: 0.53–9.84, P = 0.27] and 3.64 [95% CI: 1.21–10.89, P = 0.02], and multivariable analysis (adjusted for ECD, CIT, DGF, and recipients age) demonstrated HR of 1.34 [95% CI: 0.28–6.31, P = 0.71] and 3.42 [95% CI: 1.16–10.48, P = 0.03] for DCGF when applying terminal RR thresholds  $\geq$ 0.4 and  $\geq$ 0.2 mmHg/ml/min, respectively. When RR representing measurements over the course of HMP were modeled using mixed linear regression models, HRs for DCGF of were 1.29 [95% CI: 1.01–1.66, P = 0.05] and 1.24 [95% CI: 0.93–1.66, P = 0.15] in univariable analysis, respectively.

#### Discussion

This study sought to assess whether RR measured during HMP is an independent predictor of long-term graft outcomes in KT recipients. We found that terminal RR using a threshold of 0.2 mmHg/ml/min was associated with DCGF. Although underpowered to detect a statistically significant difference, when applying a threshold of 0.4 mmHg/ml/min, a greater effect size for risk of DCGF was observed. Finally, an increased risk of DCGF was also observed in analyses considering RR measured repeatedly over the entire course of HMP. Taken together, these findings suggest that RR is an independent predictor of long-term graft survival.

| Table 2. Baseline characte                      | eristics of total cohort | : and subgroups by ter | rminal renal resistance  | threshold of h | ypothermic machine β      | berfused kidney allograf | ts.     |
|---|--------------------------|------------------------|--------------------------|----------------|---------------------------|--------------------------|---------|
| Variable  | Total cohort<br>n = 274  | RR <0.4<br>n = 259     | RR ≥0.4<br><i>n</i> = 15 | P value        | RR <0.2<br><i>n</i> = 125 | RR ≥0.2<br>n = 149       | P value |
| Age (years)                                     | $56.08 \pm 12.69$        | $55.86 \pm 12.80$      | 59.93 ± 10.23            | 0.89           | $54.63 \pm 12.96$         | 57.30 ± 12.37            | 0.96    |
| Sex n (%)<br>Female                             | 102 (37.2)               | 94 (36.3)              | 8 (53.3)                 | 0.18           | 51 (40.8)                 | 51 (34.2)                | 0.26    |
| Male  | 172 (62.8)               | 165 (63.7)             | 7 (46.7)                 | r<br>r         | 74 (59.2)                 | 98 (65.8)                |         |
| BIVII (Kg/m <sup>-</sup> )<br>Race <i>n</i> (%) | 07.6 ± 04.77             | $21.2 \pm 5.12$        | 20.// ± 0.80             | 0.33           | $77.2 \pm 20.87$          | $26.84 \pm 5.10$         | 0.03    |
| Caucasian                                       | 182 (66.4)               | 173 (66.8)             | 6(0.0)                   | 0.59           | 88 (70.4)                 | 94 (63.1)                | 0.2     |
| Non-Caucasian                                   | 92 (33.6)                | 86 (33.2)              | 6 (40.0)                 |                | 37 (29.6)                 | 55 (36.9)                |         |
| Pre-emptive                                     | 21 (7.7)                 | 20 (7.7)               | 1 (6.7)                  | 0.48           | 10 (8)                    | 11 (7.4)                 | 0.29    |
| Hemodialysis                                    | 225 (82.1)               | 213 (82.2)             | 12 (80.0)                |                | 107 (85.6)                | 118 (79.2)               |         |
| Peritoneal dialysis                             | 22 (8.0)                 | 21 (8.1)               | 1 (6.7)                  |                | 6 (4.8)                   | 16 (10.7)                |         |
| Missing   | 6 (2.2)                  | 5 (1.9)                | 1 (6.7)                  |                | 2 (1.6)                   | 4 (2.7)                  |         |
| Cause of ESRD <i>n</i> (%)                      |                          |                        |                          |                |                           |                          |         |
| Diabetes  | 79 (28.8)                | 75 (29)                | 4 (26.7)                 | 0.63           | 40 (32)                   | 39 (26.2)                | 0.53    |
| Polycystic kidney                               | 34 (12.4)                | 32 (12.4)              | 2 (13.3)                 |                | 13 (10.4)                 | 21 (14.1)                |         |
|   |                          |                        | ĺ                        |                |                           |                          |         |
| Glomerulonephritis                              | 71 (25.9)                | 67 (25.9)              | 4 (26.7)                 |                | 28 (22.4)                 | 43 (28.9)                |         |
| Pyelonephritis/<br>interctitial nenhritic       | 49 (17.9)                | 48 (18.5)              | 1 (6. /)                 |                | 23 (18.4)                 | 26 (17.4)                |         |
|   | 11 (1E O)                | 11/2/                  | 17 3C1 N                 |                | 71 /16 8/                 | 113 11                   |         |
| Utilei<br>HLA mismatch <i>n</i> (%)             | (0.01) 14                | (C. <del>1</del> 1) (C | 4 (20.7)                 |                | (10.0)                    | (4.01) 02                |         |
| 0   | 1 (0.4)                  | 1 (0.4)                | 0                        | 0.41           | 0                         | 1 (0.67)                 | 0.91    |
| 1   | 11 (4.0)                 | 10 (3.9)               | 1 (6.7)                  |                | 6 (4.8)                   | 5 (3.4)                  |         |
| 2   | 24 (8.8)                 | 22 (8.5)               | 2 (13.3)                 |                | 9 (7.2)                   | 15 (10.1)                |         |
| m   | 79 (28.8)                | 72 (27.8)              | 7 (46.7)                 |                | 37 (29.6)                 | 42 (28.2)                |         |
| 4   | 89 (32.5)                | 86 (33.2)              | 3 (20.0)                 |                | 39 (31.2)                 | 50 (33.6)                |         |
| Ū   | 51 (18.6)                | 50 (19.3)              | 1 (6.7)                  |                | 24 (19.2)                 | 27 (18.1)                |         |
| 6   | 18 (6.6)                 | 17 (6.6)               | 1 (5.6)                  |                | 10 (8)                    | 8 (5.4)                  |         |
| Missing   | 1 (0.4                   | 1 (0.4)                | 0                        |                | 0                         | 1(0.7)                   |         |
| Donor age (years)<br>Donor sex n (%)            | $51.27 \pm 16.00$        | $50.91 \pm 16.98$      | $57.6 \pm 15.41$         | 0.94           | $48.21 \pm 15.67$         | $53.84 \pm 15.87$        | -       |
| Female  | 123 (44.9)               | 112 (43.2)             | 11 (73.3)                | 0.08           | 48 (38.4)                 | 75 (50.3)                | 0.01    |
| Male  | 147 (53.6)               | 143 (55.2)             | 4 (26.7)                 |                | 77 (61.6)                 | 70 (47)                  |         |
| Missing   | 4 (1.5)                  | 4 (1.5)                | 0                        |                | 0                         | 4 (2.7)                  |         |

| Table 2. Continued.            |                         |                           |                          |                 |                           |                           |             |
|--------------------------------|-------------------------|---------------------------|--------------------------|-----------------|---------------------------|---------------------------|-------------|
| Variable                       | Total cohort<br>n = 274 | RR <0.4<br><i>n</i> = 259 | RR ≥0.4<br><i>n</i> = 15 | P value         | RR <0.2<br><i>n</i> = 125 | RR ≥0.2<br><i>n</i> = 149 | P value     |
| Donor BMI (kg/m <sup>2</sup> ) | 28.00 ± 12.49           | 28.14 ± 12.81             | 25.47 ± 3.19             | 0.21            | 27.99 ± 6.11              | 28.00 ± 16.00             | 0.5         |
| Donor type <i>n</i> (%)<br>ECD | 162 (59.1)              | 151 (58.30)               | 11 (73.33)               | 0.19            | 66 (52.8)                 | 96 (64.4)                 | 0.05        |
| DCD                            | 22 (8.0)                | 22                        | 0                        | 0.28            | 12 (9.6)                  | 10 (6.7)                  | 0.38        |
| Donor eGFR* (ml/min            | $98.30 \pm 29.32$       | $99.47 \pm 28.72$         | $78.43 \pm 33.14$        | <0.01           | $102.72 \pm 27.51$        | $94.49 \pm 30.36$         | 0.01        |
| per 1.73 m <sup>2</sup> )      |                         |                           |                          |                 |                           |                           |             |
| CIT (h)                        | $18.31 \pm 6.14$        | $18.23 \pm 6.00$          | $19.65 \pm 8.40$         | 0.8             | $17.85 \pm 6.07$          | $18.69 \pm 6.19$          | 0.89        |
| Induction immunosuppressic     | (%) <i>u</i> u          |                           |                          |                 |                           |                           |             |
| Anti-thymocyte                 | 167 (61)                | 162 (62.5)                | 5 (33.3)                 | 0.023           | 79 (63.2)                 | 88 (59.1)                 | 0.4         |
| globulin                       |                         |                           |                          |                 |                           |                           |             |
| Alemtuzumab                    | 66 (24.1)               | 62 (23.9)                 | 4 (26.7)                 |                 | 31 (24.8)                 | 35 (23.5)                 |             |
| IL2R inhibitor                 | 25 (9.1)                | 21 (8.1)                  | 4 (26.7)                 |                 | 11 (8.8)                  | 14 (9.4)                  |             |
| Missing                        | 16 (5.8)                | 14 (5.4)                  | 2 (13.3)                 |                 | 4 (3.2)                   | 12 (8.1)                  |             |
| Maintenance immunosuppre       | ssion <i>n</i> (%)      |                           |                          |                 |                           |                           |             |
| Tacrolimus                     | 264 (96.3)              | 251 (96.9)                | 13 (86.7)                | 0.1             | 123 (98.4)                | 141 (94.6)                | 0.1         |
| Mycophenolate                  | 268 (97.8)              | 254 (98.1)                | 14 (93.3)                | 0.29            | 125 (100)                 | 143 (96)                  | 0.02        |
| Prednisone                     | 205 (74.8)              | 197 (76.1)                | 8 (53.3)                 | 0.05            | 99 (79.2)                 | 106 (71.1)                | 0.13        |
| DCD, donation after cardiac    | c death; CIT, cold isch | iemia time; eGFR, estir   | mated glomerular filtra  | tion rate; ESRD | , end stage renal disea   | se; ECD, expanded crite   | eria donor; |

HLA, human leukocyte antigen; RR, renal resistance. \*Calculated using the chronic kidney disease epidemiology collaboration equation.

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Our study provides the longest follow-up to date when evaluating the transplant outcomes in relation to pump parameters measured using the LifePort Kidney Transporter preservation system. In a study using the Gambro PF-3B preservation system and using three different RR thresholds in recipients who received grafts from DCD donors, no difference was observed in graft survival over a 15-year follow-up [10]. In a study using the MOX 100 preservation system, with a 4-year followup, higher RR was found to correlate with inferior graft outcomes [14]. Table 1 outlines the variability across studies with regard to the choice of kidney transport preservation system, the units of pump parameters considered, and the thresholds applied when verifying a relationship between pump parameters and graft outcomes. Six different devices were used across different studies. Yushkov et al. [9] suggested that thresholds for RR differ between different machines. For example, machines using a pressure controlled roller pump to deliver the perfusate create sinusoidal flow curves, and others that use a flow controlled pumping system give rise to alternate pressure waveforms [11]. When the effect of the device used for HMP was evaluated in relation to graft outcomes by Wszola et al. [28], a flow driven device was found to result in inferior graft outcomes when compared to a pressure driven device. Hence, caution should be exercised when comparing the relationship between HMP parameters and outcomes across studies.

When using the LifePort Kidney Transporter preservation system, two studies reported no relationship between RR and graft survival, and three studies showed an association; however, follow-up was limited to 1 year or less in all these five studies [8,9,11,19,20]. Jochmans et al. [11] showed that RR was an independent risk factor for 1-year graft failure in both unadjusted and adjusted Cox proportional hazards models; however, the authors only considered donor age as a covariate. When applying the RR threshold of ≥0.4 mmHg/ml/min identified by Jochmans et al., the relationship between RR and DCGF in our sample did not reach statistical significance. However, our analysis was likely underpowered to detect a difference, considering the small number of participants with RR exceeding 0.4 mmHg/ml/min. When using the median RR in our study sample (0.2 mmHg/ml/min) as a threshold, on the other hand, we did find a statistically significantly increased risk for DCGF as a function of RR with an expected lower effect size than that estimated for organs with RR greater than 0.4 mmHg/ml/min.

Although these analyses confirm that RR is an independent risk factor for DCGF, our findings do not suggest that organs with terminal RR exceeding either of these thresholds should influence decisions on organ utilization or discard. This is also supported by the fact that we observe a gradual increase in the risk for DCGF when considering changes across the range of RR measurements over the course of HMP. Evidence suggests that multiple intrinsic qualities of an allograft determine graft survival, such as donor's gender, age, height, and weight [7,11,29]. For instance, in our cohort, allografts with RR ≥0.2 mmHg/ml/min demonstrated a lower eGFR and were more likely to originate from female donors; however, our sample was too small to pursue subgroup analyses by donor sex. In addition to donor characteristics, recipient and transplant characteristics are also important determinants of graft outcomes. For example, donor-recipient sex mismatch, in particular, KT from female donors to male recipients, has been linked to inferior graft outcomes [30]. Realizing the multifactorial nature of predictors of kidney transplant outcomes, the United Network of Organ Sharing is currently using a new allocation policy that risk-stratifies deceased donors using 10 different donor factors, that are associated with all-cause graft survival [29,31]. Thus, a higher RR is unlikely to be the sole determinant of kidney transplant outcomes but rather one of several characteristics that inform on the quality of the graft.

In contrast to prior studies, in our cohort, RR was not associated with DGF (Table 1). The reasons for this could be manifold. First, our donor population included a large proportion of grafts from ECD. In a large registry analysis, we have recently shown that in some high-risk grafts from ECD, HMP use did not lower the odds of DGF [5]. The mechanism of DGF is not limited to ischemia-induced damage to the kidney tubules, but a constellation of thrombotic and inflammatory effects mediated by cytotoxins, and both the innate and adaptive immune responses [32]. This may also serve to explain why RR measured during HMP in our population with a relatively large proportion of ECD kidneys is not associated with DGF. Second, while the standard definition of DGF warrants administration of at least one dialysis session during the first week post-transplant, the number of dialysis sessions administered may vary. Consequently, the presence of DGF may not necessarily gauge the severity of graft dysfunction [4]. Third, and as mentioned previously, in addition to the considerable variability in terms of the preservation system used, no uniform practices exist across physicians and centers in terms of perioperative care including that



Figure 3 Death-censored graft failure, all-cause graft failure and death with function by terminal renal resistance threshold of 0.4 and 0.2 mmHg/ml/min.

| Variable                     | Univariable Cox pro<br>hazards model | portional | Multivariable Cox p<br>hazards model* | roportional |
|------------------------------|--------------------------------------|-----------|---------------------------------------|-------------|
| Death censored graft failure |                                      |           |                                       |             |
| Terminal RR ≥0.4             | 1.12 <b>3.23</b> 9.34                | 0.03      | <sub>0.91</sub> 2.67 <sub>7.86</sub>  | 0.07        |
| Terminal RR ≥0.2             | 1.142.67 <sub>6.31</sub>             | 0.02      | 1.022.425.72                          | 0.04        |
| RR over course of HMP        | <sub>1.07</sub> 1.31 <sub>1.59</sub> | < 0.01    | 1.00 <b>1.25</b> 1.55                 | 0.05        |
| All-cause graft failure      |                                      |           |                                       |             |
| Terminal RR ≥0.4             | <sub>0.55</sub> 1.53 <sub>4.24</sub> | 0.82      | <sub>0.34</sub> 1.00 <sub>2.85</sub>  | 0.98        |
| Terminal RR ≥0.2             | 1.142.02 <sub>3.59</sub>             | 0.02      | <sub>0.95</sub> 1.70 <sub>3.05</sub>  | 0.07        |
| RR over course of HMP        | <sub>0.98</sub> 1.20 <sub>1.45</sub> | 0.07      | <sub>0.88</sub> 1.08 <sub>1.34</sub>  | 0.49        |
| Death with function          |                                      |           |                                       |             |
| RR over course of HMP        | <sub>0.75</sub> 1.04 <sub>1.45</sub> | 0.79      | <sub>0.64</sub> 0.91 <sub>1.29</sub>  | 0.59        |

 Table 3. Hazard ratio for graft outcomes by renal resistance measured during hypothermic machine perfused kidney allografts.

CIT, cold ischemia time; DGF, delayed graft function; ECD, expanded criteria donor; HMP, hypothermic machine perfusion; RR, renal resistance.

\*Multivariable Cox proportional hazards models of graft outcomes by terminal renal resistance were adjusted for ECD, CIT, DGF, and recipient age and multivariable models of death censored graft failure by renal resistance over the course of HMP modeled using mixed linear regression models was adjusted for ECD, DGF, and recipient age.

may affect the risk of experiencing reversible and short-term graft outcomes like DGF [33,34].

As is apparent from Table 1, our study provides information on the risk of adverse transplant outcomes associated with increasing RR measures in allografts undergoing HMP using the LifePort Kidney Transporter preservation system over the longest post-transplant follow-up to date. Our study population of KT recipients includes a large proportion of ECD allowing the evaluation of the predictive role of RR in a relatively high risk donor population. Moreover, rather than relying on terminal RR alone, we apply mixed linear regression models to better capture variability in RR both between allografts and within allografts over time. Despite these strengths, some limitations need to be considered. First, this is a single-center study, the findings of which may not be applicable to all centers. As with any study evaluating the effect of HMP parameters on kidney transplant outcomes, our study is also prone to inherent bias due to selection. Mitigating this risk, however, is the fact that it is the standard practice at our center to use HMP for all deceased donor kidneys. Furthermore, if the clinical characteristics of the graft are deemed acceptable, grafts are transplanted regardless of RR. The observational nature of this study makes it vulnerable to residual confounding. For example, previous studies found histological findings such as glomerulosclerosis, tubular interstitial scarring, and arterial disease in kidney allografts demonstrating lower flow rates and higher RR [9,35]. However, histological findings on

procurement biopsies are not available for all our study participants, and, consequently, could not be accounted for in multivariable analyses. Finally, the small sample size and number of events limits our ability to adjust for all relevant confounders in the multivariable analysis and may compromise the power to detect associations between RR and study secondary endpoints.

In conclusion, our findings suggest that terminal RR and repeated measurements of RR in kidney allografts undergoing HMP are associated with DCGF. These findings, however, should not be interpreted to suggest that a particular RR measurements should be used for decisions on organ utilization or discard but rather that this is one of the several graft characteristics that need to be accounted for in organ allocation. Our observations warrant evaluation in larger, prospective, multicenter cohort studies (e.g. trial ISRCTN15821205 underway). These studies, in addition to informing on the added risk of graft failure, must also evaluate on how ascending RR may affect not only allograft survival but also the survival of kidney transplant recipients in comparison with end-stage renal disease patients remaining on dialysis.

# Authorship

SS: contributed to conception and design of the study, acquisition and interpretation of data, drafting and revising the article, providing intellectual content, final approval of the version to be published. SP and MC: contributed to conception and design of the study, interpretation of data, revising the article, providing intellectual content, final approval of the version to be published. DB, PC, JT: performed conception of the study, interpretation of data, revising the article, providing intellectual content, final approval of the version to be published. RS-P: performed conception and design of the study, analysis and interpretation of data, revising the article, providing intellectual content, final approval of the version to be published.

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# **Conflict of interest**

The authors have declared no conflicts of interest.

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# REFERENCES

- Moers C, Pirenne J, Paul A, Ploeg RJ. Machine perfusion or cold storage in deceased-donor kidney transplantation. *N Engl J Med* 2012; 366: 770.
- Maathuis MH, Manekeller S, van der Plaats A, et al. Improved kidney graft function after preservation using a novel hypothermic machine perfusion device. *Ann Surg* 2007; 246: 982; discussion 9– 91.
- 3. Moers C, Smits JM, Maathuis MH, *et al.* Machine perfusion or cold storage in deceased-donor kidney transplantation. *N Engl J Med* 2009; **360**: 7.
- Gill J, Dong J, Eng M, Landsberg D, Gill JS. Pulsatile perfusion reduces the risk of delayed graft function in deceased donor kidney transplants, irrespective of donor type and cold ischemic time. *Transplantation* 2014; 97: 668.
- Sandal S, Luo X, Massie AB, Paraskevas S, Cantarovich M, Segev DL. Machine perfusion and long-term kidney transplant recipient outcomes across allograft risk strata. *Nephrol Dial Transplant* 2018; doi: 10.1093/ndt/gfy010. [Epub ahead of print].
- Sampson D, Jun HM, Walczak P. Flow and function in machine-preserved kidneys. Br J Surg 1978; 65: 37.
- Parikh CR, Hall IE, Bhangoo RS, et al. Associations of perfusate biomarkers and pump parameters with delayed graft function and deceased-donor kidney allograft function. Am J Transplant 2015; 16: 1526.
- 8. Burgos Revilla FJ, Hevia V, Diez V, et al. Machine perfusion: initial results in an expanded criteria donor kidney transplant program. *Transplant Proc* 2015; **47**: 19.
- 9. Yushkov YY, Stern J, Ying A, et al. Identifying risk factors in renal

allografts before transplant: machinemeasured renal resistance and posttransplant allograft survival. *Prog Transplant* 2012; **22**: 175.

- de Vries EE, Hoogland ER, Winkens B, Snoeijs MG, van Heurn LW. Renovascular resistance of machine-perfused DCD kidneys is associated with primary nonfunction. *Am J Transplant* 2011; 11: 2685.
- 11. Jochmans I, Moers C, Smits JM, *et al.* The prognostic value of renal resistance during hypothermic machine perfusion of deceased donor kidneys. *Am J Transplant* 2011; **11**: 2214.
- Matsuno N, Konno O, Mejit A, et al. Application of machine perfusion preservation as a viability test for marginal kidney graft. *Transplantation* 2006; 82: 1425.
- Mozes MF, Skolek RB, Korf BC. Use of perfusion parameters in predicting outcomes of machine-preserved kidneys. *Transplant Proc* 2005; 37: 350.
- Nyberg SL, Baskin-Bey ES, Kremers W, Prieto M, Henry ML, Stegall MD. Improving the prediction of donor kidney quality: deceased donor score and resistive indices. *Transplantation* 2005; 80: 925.
- Kwiatkowski A, Danielewicz R, Kosieradzki M, *et al.* Six-year experience in continuous hypothermic pulsatile perfusion kidney preservation. *Transplant Proc* 2001; 33: 913.
- Henry ML, Sommer BG, Ferguson RM. Improved immediate function of renal allografts with Belzer perfusate. *Transplantation* 1988; 45: 73.
- Polyak M, Boykin J, Arrington B, Stubenbord WT, Kinkhabwala M. Pulsatile preservation characteristics predict early graft function in extended

criteria donor kidneys. *Transplant Proc* 1997; 29: 3582.

- Tesi RJ, Elkhammas EA, Davies EA, Henry ML, Ferguson RM. Pulsatile kidney perfusion for evaluation of highrisk kidney donors safely expands the donor pool. *Clin Transplant* 1994; 8(2 Pt 1): 134.
- Patel SK, Pankewycz OG, Nader ND, Zachariah M, Kohli R, Laftavi MR. Prognostic utility of hypothermic machine perfusion in deceased donor renal transplantation. *Transplant Proc* 2012; 44: 2207.
- Patel SK, Pankewycz OG, Weber-Shrikant E, et al. Effect of increased pressure during pulsatile pump perfusion of deceased donor kidneys in transplantation. Transplant Proc 2012; 44: 2202.
- Kozaki K, Sakurai E, Kubota K, *et al.* Prediction of kidney nonfunction after transplantation with machine perfusion preservation. *Transplant Proc* 2000; 32: 275.
- Jochmans I, O'Callaghan JM, Pirenne J, Ploeg RJ. Hypothermic machine perfusion of kidneys retrieved from standard and high-risk donors. *Transpl Int* 2015; 28: 665.
- 23. Sung RS, Christensen LL, Leichtman AB, *et al.* Determinants of discard of expanded criteria donor kidneys: impact of biopsy and machine perfusion. *Am J Transplant* 2008; **8**: 783.
- 24. Guarrera James VJ, Goldstein MJ, Samstein B, Henry S, Reverte C. 'When good kidneys pump badly': outcomes of deceased donor renal allografts with poor pulsatile perfusion characteristics. *Transplant Int* 2010; 23: 444.
- 25. Cnaan A, Laird NM, Slasor P. Using the general linear mixed model to analyse unbalanced repeated measures and

longitudinal data. Stat Med 1997; 16: 2349.

- Rizopoulos D. Joint Models for Longitudinal and Time-to-Event Data: With Applications in R. Boca Raton, FL: Chapman and Hall/CRC, 2012.
- Kidney Disease: Improving Global Outcomes (KDIGO) Anemia Work Group. KDIGO 2012 clinical practice guideline for the evaluation and management of chronic kidney disease. *Kidney Int Suppl* 2013; 3: 1.
- Wszola M, Kwiatkowski A, Diuwe P, et al. One-year results of a prospective, randomized trial comparing two machine perfusion devices used for kidney preservation. *Transpl Int* 2013; 26: 1088.
- 29. Rao PS, Schaubel DE, Guidinger MK, et al. A comprehensive risk quantifica-

tion score for deceased donor kidneys: the kidney donor risk index. *Transplantation* 2009; **88**: 231.

- 30. Zhou JY, Cheng J, Huang HF, Shen Y, Jiang Y, Chen JH. The effect of donorrecipient gender mismatch on shortand long-term graft survival in kidney transplantation: a systematic review and meta-analysis. *Clin Transplant* 2013; 27: 764.
- Israni AK, Salkowski N, Gustafson S, et al. New national allocation policy for deceased donor kidneys in the United States and possible effect on patient outcomes. J Am Soc Nephrol 2014; 25: 1842.
- Nashan B, Abbud-Filho M, Citterio F. Prediction, prevention, and management of delayed graft function: where are we now? *Clin Transplant* 2016; **30**: 1198.

- Jochmans I, Akhtar MZ, Nasralla D, et al. Past, present, and future of dynamic kidney and liver preservation and resuscitation. Am J Transplant 2016; 16: 2545.
- 34. Sandal S, Bansal P, Cantarovich M. The evidence and rationale for the perioperative use of loop diuretics during kidney transplantation: a comprehensive review. *Transplant Rev (Orlando)* 2017; pii: S0955-470X(17)30084-8. doi: 10.1016/j.trre.2017. 11.002. [Epub ahead of print]
- 35. Patel SK, Pankewycz OG, Weber-Shrikant E, *et al.* Graft arteriosclerosis and glomerulosclerosis correlate with flow and resistance to machine perfusion in kidney transplantation. *Transplant Proc* 2012; **44**: 2197.